



# Return of the ghosts of dispersal past: historical spread and contemporary gene flow in the blue sea star *Linckia laevigata*

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**ABSTRACT.**—Marine animals inhabiting the Indian and Pacific oceans have some of the most extensive species ranges in the world, sometimes spanning over half the globe. These Indo-Pacific species present a challenge for study with both geographic scope and sampling density as limiting factors. Here, we augment and aggregate phylogeographic sampling of the iconic blue sea star, *Linckia laevigata* Linnaeus, 1758, and present one of the most geographically comprehensive genetic studies of any Indo-Pacific species to date, sequencing 392 base pairs of mitochondrial COI from 791 individuals from 38 locations spanning over 14,000 km. We first use a permutation based multiple-regression approach to simultaneously evaluate the relative influence of historical and contemporary gene flow together with putative barriers to dispersal. We then use a discrete diffusion model of phylogeography to infer the historical migration and colonization routes most likely used by *L. laevigata* across the Indo-Pacific. We show that estimates of genetic structure have a stronger correlation to geographic distances than to “oceanographic” distances from a biophysical model of larval dispersal, reminding us that population genetic estimates of gene flow and genetic structure are often shaped by historical processes. While the diffusion model was equivocal about the location of the mitochondrial most recent common ancestor (MRCA), we show that gene flow has generally proceeded in a step-wise manner across the Indian and Pacific oceans. We do not find support for previously described barriers at the Sunda Shelf and within Cenderwasih Bay. Rather, the strongest genetic disjunction is found to the east of Cenderwasih Bay along northern New Guinea. These results underscore the importance of comprehensive range-wide sampling in marine phylogeography.

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Marine biodiversity is concentrated in the Indo-Pacific region, with species diversity reaching its highest values in the Coral Triangle, a region centered in the Indo-Malay-Philippines archipelago (Roberts et al. 2002, Carpenter and Springer 2005, Tittensor et al. 2010). The Coral Triangle has a complex geological history with much tectonic activity, including substantial reconfigurations of landmasses due to moving plates (Hall 2002). Fluctuating sea levels have also substantially restructured land and sea configurations, as the shallow continental shelf is extensive in this region (Voris 2000). For example, sea levels 18,000 yrs ago are thought to have been about 130 m lower than present day levels, resulting in greatly reduced area for most shallow marine habitats and thus severe reductions in local population sizes (Crandall et al. 2012a). At that time, the Makassar Strait was much narrower than it is today, almost completely blocking the marine connection between the Pacific and Indian oceans (Chappell and Shackleton 1986). As sea levels rose over the next 10,000 yrs, continental shelf habitat of an area slightly smaller than the land area of the country of India (approximately  $3.16 \times 10^6 \text{ km}^2$ ) would have resubmerged and become available again to local marine species (Voris 2000).

Perhaps unsurprisingly, phylogeographic studies have revealed the imprint of these sea level changes on population genetic patterns of Coral Triangle species (reviewed in Carpenter et al. 2011). Some taxa show nearly reciprocal monophyly in mtDNA over relatively short distances, probably as a consequence of isolation during the Pleistocene (e.g., Barber et al. 2000, 2002). For many taxa, interestingly, the locations of likely vicariance due to Pleistocene sea level change are also associated with regions of persistent population genetic structure, most notably the Sunda shelf (McMillan et al. 1999, DeBoer et al. 2008, Ackiss et al. 2013), Torres Strait (Mirams et al. 2011), and Halmahera Eddy (Barber et al. 2006, 2011), although there is also evidence that this structure is being eroded by contemporary dispersal in some species (Gaither et al. 2011, Liu et al. 2012, DeBoer et al. 2014). Finally, nearly all loci and taxa that have been examined to date show signatures of Pleistocene-era population expansions onto newly submerged continental shelf habitats (e.g., Chenoweth et al. 1998, Lind et al. 2007, Crandall et al. 2008a,b, 2012a, Gaither et al. 2010).

Given the globally acknowledged value of the Coral Triangle in terms of marine biodiversity as well as the multitude of current threats to the region (Roberts et al. 2002, Burke et al. 2011), it is of practical importance to understand both the history of its marine communities (how and from where did genetic and species diversity arise? e.g., Renema et al. 2008, Williams and Duda 2008, reviewed in Bowen et al. 2013), as well as how the regional seascape is presently constructed (how are different parts of the region connected demographically by larval dispersal? reviewed in Riginos and Liggins 2013). Previous phylogeographic attempts to answer these questions have focused on the measurement of genetic structure ( $F_{ST}$  and its analogues, Wright 1950), and testing specific hypotheses of population structure primarily with analyses of molecular variance (AMOVA, Excoffier et al. 1992). However, these approaches based on allele frequencies are poorly suited to organisms with evolutionarily high levels of gene flow (10–100 migrants per generation) and large coalescent effective population sizes (partially arising from high gene flow), which depress traditional estimates of population structure (such as  $F_{ST}$ , Hedrick 2005). Furthermore, because  $F_{ST}$  and AMOVA summarize a combined model of gene flow and effective population size (Whitlock 2011), these frequency-based approaches allow estimation of marine population structure only at a very coarse resolution, often resulting

in substantial bias when parameters such as effective population size and timing of population divergence are not considered (Bird et al. 2011, Marko and Hart 2011, Faurby and Barber 2012). As an alternative, one might consider the inverse approach: testing explicit hypotheses of gene flow rather than genetic structure (Crandall et al. 2012b).

The substitution of gene flow for genetic structure as the parameter of interest in seascape genetics makes intuitive sense. Gene flow in most marine organisms is mediated by the planktonic larval stage, where millions of larvae disperse through a complex milieu of currents and environmental conditions. Thus, there are few impermeable barriers to larval dispersal in the ocean: for almost every physical or oceanographic entity that is thought to impede larval dispersal for some species (see examples in Rocha et al. 2007) one can find several species that show no evidence of isolation whatsoever (Lessios and Robertson 2006, Carpenter et al. 2011, Toonen et al. 2011). Although ocean currents and land masses may effectively act as barriers to gene flow, it is more appropriate to think in terms of probabilistic larval dispersal kernels for which the probability of a successful dispersal event (and therefore of gene flow) declines sharply with distance due to larval diffusion, behavior and mortality (Cowen et al. 2000, Gerlach et al. 2007, Buston et al. 2012). The most informed hypotheses of gene flow therefore come from modeling such dispersal through oceanographic current vectors while taking these additional factors into account (Kool et al. 2011, Trembl et al. 2012). Empirical testing with genetic data has shown this to be the case using coalescent estimates of gene flow (Crandall et al. 2012b), assignment tests (Fievet et al. 2007), parentage-based tagging (Saenz-Agudelo 2012) or  $F_{ST}$  (Galindo and Palumbi 2006, White et al. 2010, Alberto et al. 2011, Foster et al. 2012). Until recently, gene flow estimates have been constrained to the unrealistic assumptions of Wright's island model by relying on the  $F_{ST}$  summary statistic (Whitlock and McCauley 1999), which assumes equal levels of gene flow throughout the sampled area and does not take historical factors (such as lineage sorting) into account. However, coalescent modeling approaches now allow flexible evaluation and selection of specific models of gene flow, which are estimated simultaneously with the genealogical history of genetic sequence data (Hey and Nielsen 2007, Lemey et al. 2009, Beaumont 2010, Beerli and Palczewski 2010).

A further challenge to understanding genetic diversity in the Coral Triangle arises from its location: it exists near the junction of the Indian and Pacific Oceans, at the center of the Indo-Pacific region, which is the largest biogeographic region on Earth (Spalding et al. 2007). Many species found in the Coral Triangle have vast ranges that may include large portions of the Indian and Pacific oceans. Although there have been considerable recent efforts to document genetic patterns within the Coral Triangle, there are few studies with dense sampling (many locations, many individuals per location) that include both the Coral Triangle and surrounding regions of the Indo-Pacific (see Keyse et al. 2014). Although logistically challenging, large-scale geographic coverage is necessary to determine the context of genetic variation of marine species. Without broad-scale sampling that includes both the Coral Triangle and other parts of the species' range, it is not possible to completely resolve the extent of divergent genetic lineages (Manel and Holdregger 2013).

The sea star *Linckia laevigata* Linnaeus, 1758, easily recognizable for its striking blue coloration, is one of the best-studied species in the Indo-Pacific region. Besides the well-known royal blue phenotype, several color variations are reported

from different geographic regions without apparent differentiation by morphotype (Williams 2000). Like many other marine benthic taxa, adult *L. laevigata* are sedentary, but the larvae have a moderate (at least 22 d before metamorphosis, Yamaguchi 1973) pelagic larval duration (PLD) after external fertilization.

Early genetic surveys of *L. laevigata* were based on allozymes (Williams and Benzie 1993, Williams and Benzie 1996, 1998, Williams et al. 2002) or mtDNA (Williams and Benzie 1997, 1998, Williams et al. 2002, Crandall et al. 2008b, Kochzius et al. 2009) and have had broad coverage from the western Pacific to the Indian ocean without much sampling in the Coral Triangle (e.g., Williams and Benzie 1998) or have exclusively focused sampling within the Coral Triangle (Crandall et al. 2008b, Kochzius et al. 2009). At the broadest scale, support was found for differentiation between the Indian and Pacific oceans based on allozymes, albeit with individuals from Western Australia (Ningaloo) showing greater affinity to western Pacific populations than to western Thailand and South Africa (Williams and Benzie 1998, Williams et al. 2002, see also Vogler et al. 2013 for similar results in crown-of-thorns starfish). For mtDNA COI sequences, Williams et al. (2000) described two major clades: an "Indian Ocean" clade, which included both Pacific and Indian Ocean individuals and a "Pacific Ocean" clade that contains only Pacific Ocean individuals with the exception of a few western Australian individuals. Crandall et al. (2008b) as well as Kochzius et al. (2009) sampled comprehensively within the Coral Triangle and found that the Indian clade haplotypes dominate most Indonesian populations, declining in frequency from Aceh in the west to Jayapura in the east. Within the Coral Triangle, the greatest population structure was found by grouping Aceh with Krakatau against a cluster of remaining locations, whereas less support was found for a west-east delineation defined by the Sunda Shelf, and there was modest support for distinctiveness of Teluk Cenderawasih (Crandall et al. 2008b). Kochzius et al. (2009) suggested that *L. laevigata* has historically expanded populations into the western Pacific from eastern Indian Ocean origins, a route of colonization running counter to the Indonesian Throughflow (Kochzius et al. 2009).

In the present study we combine mtDNA data from some of the previous studies cited above with new sampling from 18 additional locations to evaluate the influence of putative historical and contemporary gene flow and spatial features on genetic patterns within *L. laevigata*. Although data from additional loci are desirable for future studies, the rich genealogical information in this mtDNA-only data set allows inference from the recent (approximately 100 kya) matrilineal history of this species and the cross compatibility of this DNA sequence data facilitates synergism across research groups (Bowen et al. 2014). The data set analyzed here represents the single most extensive population genetic survey both in geographic extent (diameter >14,000 km) and density for any species to date from the Indo-Pacific region (Keyse et al. 2014); its compilation was only possible by cooperation and collaboration among research groups.

We use this data set to consider the genealogical history of *L. laevigata* in terms of inferred mitochondrial gene flow in addition to estimates of genetic structure. We begin by parameterizing a biophysical model of contemporary larval dispersal to predict mean dispersal distances for *L. laevigata* larvae among sampling sites. We then use a multiple regression approach based on permutation (Legendre et al. 1994) to ask whether measurements of genetic structure in this species are better explained by the mean dispersal distances from the biophysical model, or by geographic

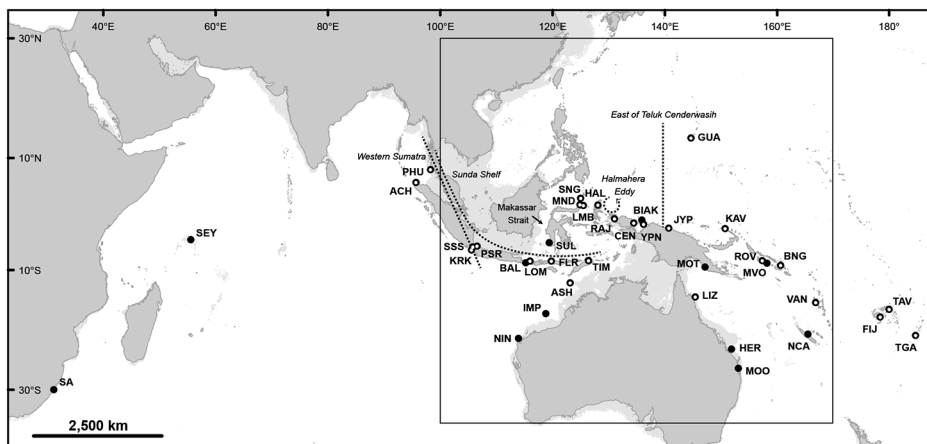


Figure 1. Sampling locations; see Table 1 for abbreviations. Open circles represent locations with sample size  $>10$ . The domain for the biophysical model is represented by a box. Four putative barriers to gene flow that were evaluated under the MRDM and AMOVA approaches are denoted as dotted lines.

distances. The biophysical model assesses whether mitochondrial patterns are best explained by equilibrium gene flow from contemporary larval dispersal whereas the geographic distance model assesses the relative importance of colonization processes and historical gene flow. We also use the biophysical model output to define modular geographic regions with greater larval connectivity within each region than between regions. These regions are then used in a phylogeographical model that reconstructs historical gene flow through time by treating each region as a character state that can be inferred for each node on the genealogy through ancestral state reconstruction (Lemey et al. 2009). This approach allows us to infer the vectors of gene flow that were important in the spread of this lineage throughout the Indo-Pacific region.

## METHODS

**STUDY SITES AND COI SEQUENCING.**—Mitochondrial cytochrome oxidase I sequences from blue morph *L. laevigata* were obtained from both newly sampled individuals and from previous studies (Williams 2000, Crandall et al. 2008b). New samples were obtained from locations in western and eastern Australia, East Timor, Papua New Guinea, the Solomon Islands, Vanuatu, Fiji, and Tonga (see Fig. 1 and Table 1 for more details), using tube feet preserved in ethanol. New sequences were also obtained from locations in Thailand, Indonesia, Guam, and New Caledonia using old pyloric caecum samples preserved in DMSO, which were previously analyzed in an allozyme study (Williams et al. 2002). DNA preparation was undertaken at the University of Queensland (including NIN, ASH, TIM, KAV, MVO, MOT, LIZ, HER, MOO, TGA, ROV; see Table 1 for abbreviations), Boston University (including ROV, BNG, VAN, FIJ, and TAV) and Ludwig-Maximilians-Universität München (including LOM, PHU, NCA, and GUA). At the University of Queensland, genomic DNA was extracted using a modified salt extraction protocol (Aljanabi and Martinez 1997). The cytochrome oxidase I (COI) gene was amplified using polymerase chain reaction (PCR), with each reaction containing approximately 1  $\mu$ l of DNA and using Titanium

Table 1. Sampling locations and summary statistics. Number of samples (*n*), Haplotype diversity (*H*), percent segregating sites (*S*), nucleotide diversity ( $\pi$ ), Tajima's *D*, and Fu's *F<sub>s</sub>*

Location	CODE	Latitude	Longitude	Source	<i>n</i>	<i>H</i>	<i>S</i> (%)	$\pi$ (%)	<i>D</i>	<i>F<sub>s</sub></i>
South Africa	SA	-30.1	31.2	Williams 2000, 2002	5	1.00	1.8	0.9	0.50	-1.90
Seychelles	SEY	-4.6	55.6	Williams 2000, 2002	4	0.25	0.0	0.0	0.00	0.00
Ningaloo Reef, Australia	NIN	-21.7	114.0	Present study: Riginos	7	0.86	2.8	1.2	0.44	-1.19
Imperieuse Reef, Australia	IMP	-17.5	118.8	Williams 2000, 2002	3	1.00	2.0	1.4	0.00	0.46
Ashmore Reef, Australia	ASH	-12.2	123.1	Present study: Riginos	15	0.80	6.1	1.9	-0.04	-3.12
East Timor	TIM	-8.3	126.4	Present study: Riginos	19	0.68	4.8	1.1	-0.76	-4.90*
Aceh, Indonesia	ACH	5.6	95.7	Crandall et al. 2008	15	0.73	4.3	1.0	-0.96	-4.31**
Phuket, Thailand	PHU	7.9	98.3	Present study: Yasuda	28	0.75	6.9	1.0	-1.59*	-15.47
Krakatau	KRK	-6.1	105.5	Crandall et al. 2008	48	0.65	9.4	1.0	-1.79*	-25.36
Sebesi/Sebuku/Sangiang	SSS	-5.9	105.5	Crandall et al. 2008	51	0.51	9.4	1.4	-1.19	-11.13**
Pulau Seribu	PSR	-5.7	106.6	Crandall et al. 2008	79	0.48	10.7	1.4	-1.19	-22.60
South Sulawesi	SUL	-5.1	119.4	Crandall et al. 2008	7	0.86	3.8	1.5	-0.28	-0.84
Bali	BAL	-8.7	115.3	Crandall et al. 2008	5	0.80	3.6	1.7	0.09	0.98
Lombok	LOM	-8.4	116.0	Crandall et al. 2008, Present study: Yasuda;	23	0.70	4.3	1.2	0.05	-7.10***
Flores	FLR	-8.4	119.8	Crandall et al. 2008	14	0.79	4.8	1.4	-0.39	-3.53*
Manado	MND	1.6	124.9	Crandall et al. 2008	76	0.43	10.5	1.3	-1.21	-16.03***
Lembah	LMB	1.5	125.2	Crandall et al. 2008	20	0.60	6.1	1.4	-0.70	-2.25
Sangihe	SNG	2.8	125.4	Crandall et al. 2008	17	0.47	3.1	0.8	-0.48	-1.20
Halmahera	HAL	1.5	128.0	Crandall et al. 2008	75	0.41	11.2	1.3	-1.37	-13.45***
Raja Ampat	RAJ	-0.9	131.1	Crandall et al. 2008	31	0.42	5.4	1.2	-0.39	-1.85
TelukCenderawasih	CEN	-1.7	134.5	Crandall et al. 2008	22	0.68	5.9	1.6	-0.10	-4.34*
Biak	BIAK	-1.1	136.0	Crandall et al. 2008	7	0.86	4.8	1.9	-0.24	-0.38
Yapan	YPN	-1.9	136.2	Crandall et al. 2008	19	0.42	3.8	0.9	-0.79	-0.62
Jayapura	JYP	-2.5	140.7	Crandall et al. 2008	19	0.58	4.1	1.1	-0.23	-2.54
Guam	GUA	13.5	144.7	Present study: Yasuda; Williams 2000, 2002	25	0.68	5.4	1.3	-0.21	-6.82**
Kavieng, PNG	KAV	-2.6	150.8	Present study: Riginos	16	0.69	4.1	1.3	0.06	-2.97



Table 1. Continued.

Location	CODE	Latitude	Longitude	Source	<i>n</i>	H	S (%)	$\pi$ (%)	D	$F_s$
Roviana, SOL	ROV	-8.3	157.4	Present study: Riginos	18	0.72	5.9	1.3	-0.91	-4.46*
Marovo, SOL	MVO	-8.8	158.3	Present study: Riginos	5	1.00	2.8	1.3	-0.38	-1.35
Boneagi, SOL	BNG	-9.2	160.7	Present study: Crandall	10	0.60	2.8	1.1	0.59	0.09
Motupore, PNG	MOT	-9.4	147.2	Present study: Riginos	8	0.88	3.6	1.3	-0.11	-1.78
Lizard Island, GBR	LIZ	-14.7	145.5	Present study: Riginos	16	0.81	6.6	1.5	-0.92	-4.75*
Heron Island, GBR	HER	-23.5	151.9	Present study: Riginos	9	0.89	5.1	1.6	-0.70	-2.10
Mooloolaba, QLD	MOO	-26.6	153.1	Present study: Riginos	3	1.00	0.8	0.5	0.00	-0.69
New Caledonia	NCA	-21.0	165.6	Present study: Yasuda	5	0.80	2.0	1.0	0.29	0.13
Vanuatu	VAN	-15.6	167.0	Present study: Crandall	16	0.63	5.4	1.1	-1.31	-2.35
Viti Levu, Fiji	FIJ	-18.1	178.4	Present study: Crandall	20	0.40	4.1	1.0	-0.39	0.08
Taveuni, Fiji	TAV	-16.8	180.0	Present study: Crandall	10	0.80	3.8	1.5	0.37	-1.62
Tonga	TGA	-21.2	-175.3	Present study: Riginos	12	0.75	2.6	0.8	-0.42	-4.06**

\*  $P < 0.05$ ; \*\*  $P < 0.01$ ; \*\*\*  $P < 0.001$

*Taq* polymerase (Clontech Laboratories, Inc.) and the universal COI primers from Folmer et al. (1994). Amplicons were purified with an exo/sap procedure (New England Biolabs) and sent to Macrogen (Korea) for capillary sequencing. At Boston University, sequencing protocols followed those outlined in Crandall et al. 2008b. At Ludwig-Maximilians-Universität München, newly developed *L. laevigata*-specific primers (LL-F1, 5'-ACCACCGGCTGGGTCGAA-3' and LL-R1, 5'-TAATCTTTGGGGCGTGAGC-3') were used for PCR and sequencing for three populations (PHU, GUA, and NCA), to increase PCR efficiency. The amplifications were made in 10  $\mu$ l reactions with a final concentration of 3mM of  $MgCl_2$ , 1 $\times$  buffer, 0.3  $\mu$ M of each primer, 0.8 mM of dNTP, 0.07 units of Kapa *Taq* DNA Polymerase (Kapa biosystems) and 1  $\mu$ l of DNA template. Thermocycling consisted of denaturation of DNA at 94 °C for 3 min, 35 cycles of 94 °C for 30 s, 50 °C for 30 s and 72 °C for 30 s, followed by final extension of DNA at 72 °C for 7 min and cooling at 4 °C. The amplification of PCR products was confirmed by gel electrophoresis in a 1% agarose gel. Sequencing reactions were conducted on an ABI 3730 capillary sequencer using BigDye v.3.1. Sequence was determined from both directions using LL-F1 and LL-R1 primers. Trace files were manually checked and edited using CodonCode v3.0.2. These newly generated sequences were aligned against published sequences from Williams (2000), Williams et al. (2002), and Crandall et al. (2008b). Individual haplotypes from Crandall et al. (2008b) were assigned in the same manner as the original paper.

In total, we aggregated mtDNA haplotypes from 38 localities (Fig. 1) and data from all locations were represented in the haplotype network (Fig. 2). To obtain an accurate and unbiased estimate of pairwise genetic relationships between locations ( $\Phi_{ST}$ ,  $F_{ST}$ , and  $D_{est}$  with respect to Euclidean distance and overwater distances), we included only those locations which had at least 10 individuals sampled, leaving 26 populations for these analyses (Fig. 1, sites with white centers). Those locations within the domain of the biophysical model (box in Fig. 1) were used for two purposes. First, those sample sites with more than 10 individuals sampled, that also contained substantial reef habitat were used for reanalyzing the pairwise genetic relationships between locations with respect to dispersal distances derived from the biophysical model (20 total sites; all white sites within box in Fig. 1 excluding Jayapura, which does not have substantial reefs, and is thus not included in the biophysical model). Second, all collection locations within the model domain were used, except Jayapura and Mooloolaba, due to reef representation to extract the geographic cluster membership of the remaining 29 sample locations.

**GENETIC DIVERSITY AND DIFFERENTIATION.**—To visualize the total COI diversity, a median joining haplotype network was constructed in Network 4.611 and edited in Network Publisher 2.0 (fluxus-engineering.com, Bandelt et al. 1999). To reduce complexity, non-parsimonious links were deleted using the maximum parsimony calculation option (MP, Polzin and Daneshmand 2003). Colors across the haplotype network represented different biophysically derived clusters (see below) and other locations/regions beyond the scope of the biophysical model, but assumed to be distinct demographically due to their geographic isolation, and significant genetic structure (Online Table S1).

Standard population summary statistics were calculated in Arlequin v3.5.1.3 (Excoffier and Lischer 2010). Statistics calculated included Watterson's  $\theta$  (Watterson



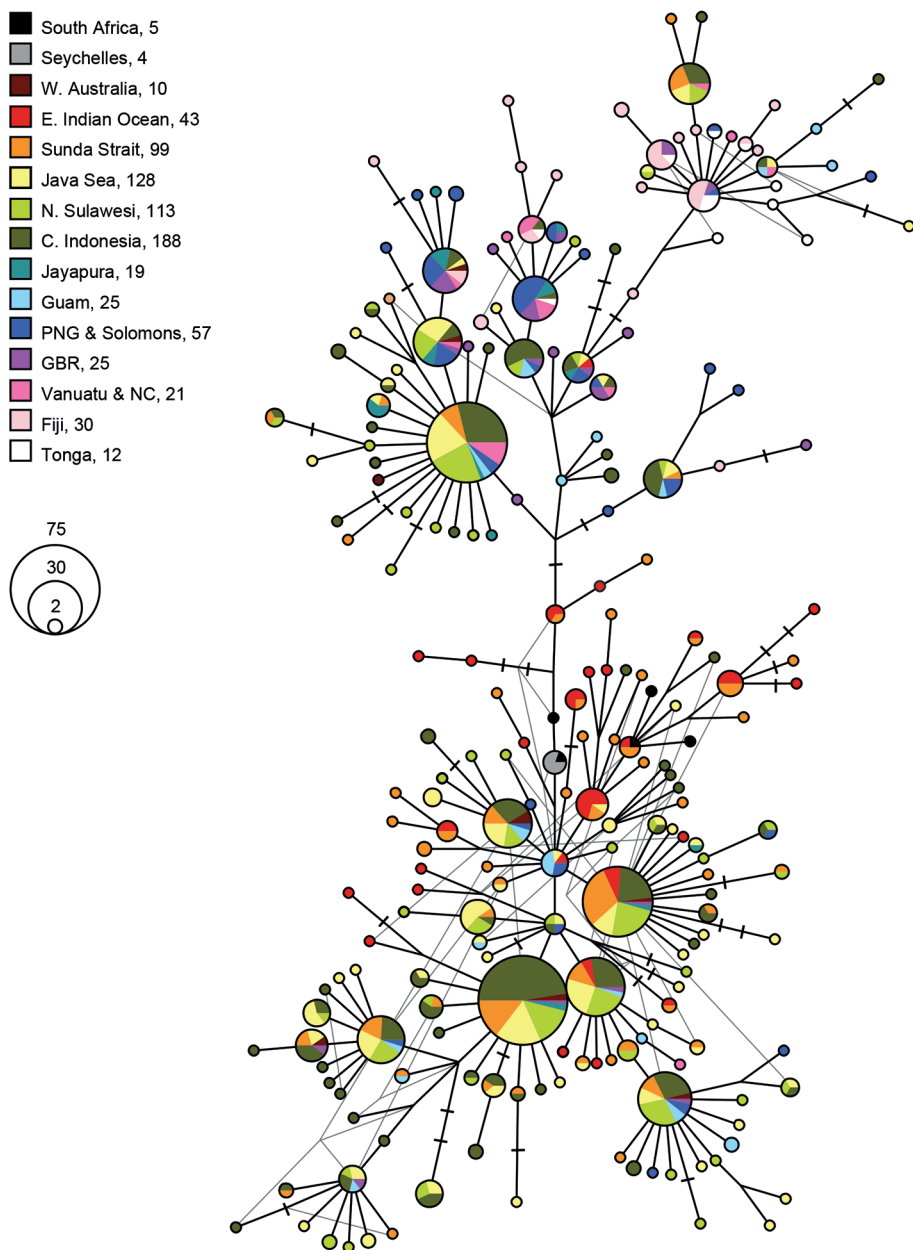


Figure 2. Median-joining haplotype network for *Linckia laevigata* colored according to modular clusters determined from the biophysical model. The number of individual sequences included in each cluster follow the name in the key. The frequency of each haplotype is indicated by size (see key, bottom right). Edges between haplotypes or small cross-bars indicate a mutational step. Black edges represent one of the maximum parsimony networks chosen at random; grey edges represent alternate relationships among haplotypes found in 29 other equally parsimonious networks.

1975), which estimates the average number of polymorphic sites, and  $\pi$ , which estimates the average number of differences between two random sequences from the same population. Tajima's  $D$  (Tajima 1989) and Fu's  $F_s$  (Fu 1997) were also measured to compare *L. laevigata* COI diversity against neutral, equilibrium expectations.

We used a variety of approaches to summarize genetic differentiation. Pairwise relationships among populations (for sampling locations including 10 individuals or more: 26 populations) were described by  $\Phi_{ST}$  (based on the Tamura-Nei distance as selected by jModeltest; Posada 2009) as well as  $F_{ST}$  (based on haplotype identities) in Arlequin. To reduce the effects of high allelic variability, we also used  $D_{est}$  (Jost 2008) calculated in Genodive (Meirmans and Van Tienderen 2004) by reducing infiles to haplotype identity. Relationships among populations based on these pairwise statistics were visualized using non-metric multidimensional scaling (vegan package in R, Oksanen et al. 2012).

We used AMOVA (Excoffier et al. 1992) analysis in Arlequin to test a priori predictions about various spatial configurations of genetic differentiation arising from barriers to dispersal. To test the hypothesis that divergence between Indian and Pacific populations explains the most molecular variance in COI, we tried two different spatial configurations. We first separated locations in western Indonesia from central Indonesia (following Crandall et al. 2008b), and then delineated Indian and Pacific populations along the Sunda Shelf. The contribution of the Halmahera Eddy to population structure was assessed by configurations whereby central Indonesia locations were grouped separately from Cenderwasih locations, and these groupings were contrasted against configurations that combined central Indonesia, and Cenderwasih locations as a single group. Similarly the distinctiveness of Cenderwasih from the Pacific was evaluated by comparing separate and combined groupings. Furthermore, Guam was allowed to group by itself and with other Pacific locations due to preliminary analyses pointing to genetic distinctiveness of this location. These varying spatial configurations were evaluated using AMOVA based on  $\Phi$ -statistics derived from Tamura-Nei distances between haplotypes and from haplotypes identified in Arlequin (as above). For AMOVAs based on haplotype identity, we also used  $F'$ -statistics (Meirmans and Van Tienderen 2004).

**GEOGRAPHIC AND BIOPHYSICALLY INFORMED PREDICTIONS OF GENE FLOW AMONG SAMPLE LOCATIONS.**—Three pair-wise distance metrics were calculated as proxies for gene flow (i.e., realized dispersal over many generations) among sampled populations. As a simple null model of gene flow, we calculated the Euclidean distance among all 38 sampled locations. Although this model of gene flow implies that dispersal occurs relative to straight-line routes only, it is a common dispersal distance proxy used in marine population genetic studies (Riginos and Liggins 2013). The second dispersal distance proxy used for all locations was the shortest over-water distance calculated with a least-cost path algorithm. The distance calculations are similar to the Euclidean distance, but the least-cost path was forced around all land boundaries. These two measures of geographic distance are expected to explain more of the variance in genetic structure when historical processes such as colonization dominate the signal from gene flow (Selkoe 2008).

Finally, we used a biophysical model of larval dispersal (Treml et al. 2012) to quantify the relative dispersal strength among sampled populations within the model domain (Fig. 1). This dispersal model includes coral reef habitat (Spalding et al. 2001),

oceanographic data describing sea surface currents for three years (ROMS, Wang et al. 2005), and several biological parameters describing the dispersal characteristics of *L. laevigata*: seasonal spawning periodicity, a 25-d maximum pelagic larval duration, a 1- to 2-d precompetency period, strong late-stage swimming/homing behavior, and a 30% d<sup>-1</sup> larval mortality (Yamaguchi 1973). The model outputs the probability that larvae released in one location survive and settle in every other recipient location, summarized as a 1002 × 1002 source-reef by destination-reef matrix. This dispersal probability matrix was converted to a migration matrix representing the proportion of settlers to every reef patch that came from all upstream larval sources. See Treml et al. (2012) for model details and sensitivity analysis. The migration matrix, *M*, was converted to “oceanographic dispersal distance” using  $\log(M^{-1})$  to transform the values to be the same rank-order as geographic distance (high proportion of settlers then have a short distance) required for many network-based algorithms. This inverse dispersal strength matrix was used as a proxy for dispersal distance, and is referred to as such throughout this paper. This oceanographic dispersal distance is expected to explain more of the variance in genetic structure when contemporary dispersal events dominate the signal relative to historical connections (White et al. 2010).

To identify the emergent geographic clustering of reef habitat (and sample locations) determined by the dispersal strengths represented in the migration matrix, we used a network-based leading eigenvector community detection algorithm (Newman 2006). This algorithm identifies the optimal clustering within a network by optimizing the network’s modularity, or simply maximizing the density of within-cluster connections while minimizing between-cluster connections. The original asymmetric migration matrix was converted to a symmetric matrix by taking the maximum dispersal strength between all pairs of reefs. *Linckia laevigata* sample sites were overlaid with the network clustering results thereby revealing the potential clustering of sample sites based on dispersal potential among all reefs. These spatial clusters were then used to aggregate sampling sites in the phylogeographic diffusion model described below.

**EVALUATING GENETIC DIFFERENTIATION USING DISPERSAL PROXIES.**—We used a multiple regression on distance matrices (MRDM, Legendre et al. 1994) to evaluate the relative influence geographic distance and multiple discrete landscape factors that might contribute to population genetic structure; this methodology performs favorably compared to many other methods (Balkenhol et al. 2009). In a simple model where the linear relationship between geographic distance and genetic distance are evaluated, MRDM is equivalent to a Mantel test. Dispersal distance proxies included Euclidean distances and overwater distance, as well as the biophysical dispersal distances between populations. The biophysical distances represent a hypothesis of contemporary dispersal. Euclidean and overwater distances could represent simpler (null) models of contemporary dispersal, but they also might capture historical averages of dispersal (that is both recurrent gene flow and colonization). The predictive contributions of four putative barriers/divisions were also evaluated including (1) Western Sumatra, as found in Crandall et al.’s (2008) earlier survey of *L. laevigata*; (2) Sunda Shelf, a focal point of Pleistocene marine disjunctions due to the expansion of the Sunda Shelf land mass at low sea level stands; (3) Halmahera Eddy, a contemporary hydrodynamic barrier; and (4) Pacific east of Cenderwasih, which might

represent a location of habitat limitation. For barriers, dummy variables (0 vs 1) were coded with 0 for population pairs found on the same side of the putative barrier.  $\Phi_{ST}$ ,  $F_{ST}$ , and  $D_{est}$  values between pairs of populations were used as response variables. All variables were normalized and both forward and backward model selection were implemented as in Legendre et al. (1994). Models including Euclidean and overwater distances included all populations with sample size  $\geq 10$  (26 populations). Because biophysical distances were only available for some population pairs, the analyses involving these predictors was reduced to a more restricted subset of populations (20 populations). In addition, the decomposed matrices involving biophysical predictors were twice as large because the distances are asymmetric (distance from  $X$  to  $Y$  does not equal  $Y$  to  $X$ ). Permutated probabilities for all matrix regression models were evaluated with ImPerm (Wheeler 2010).

**BAYESIAN SELECTION OF IMPORTANT MIGRATION PARAMETERS.**—To estimate the historical gene flow required to explain the current distribution of mitochondrial genetic variation in *L. laevigata*, we modeled the phylogeographic history of COI as a discrete diffusion process following methods developed by Lemey et al. (2009). Using a coalescent perspective, we assigned a geographic location as a discrete character trait having one of 14 possible states (based on spatial clusters delineated by the biophysical model, see below) to each COI sequence, and reconstructed the most probable location of each ancestor back to the most recent common ancestor (MRCA) of the entire sample using BEAST 1.6.2 (Drummond and Rambaut 2007). Changes in location between ancestor and descendent nodes were modeled as a migration event, the probability of which was governed by a time-reversible matrix of migration rates among locations (similar to the GTR model used for models of molecular evolution). To reduce the degrees of freedom in the matrix, this method uses Bayesian stochastic search variable selection to only allow  $W$  of the rates to be non-zero, where the prior on  $W$  is a truncated Poisson distribution with mean  $\eta$  (see Lemey et al. 2009 for full details). In a Bayesian framework, the geographic location of the MRCA, as well the migration events required to explain most topologies can be estimated simultaneously with models of nuisance parameters such as those for molecular evolution, demography and tree topology.

Because our sequences came from 38 different localities, many with relatively small sample sizes, we assigned their geographic location as one of the  $k = 9$  spatial clusters resulting from the clustering algorithm on the migration matrix plus the following groups of sites based on geographic isolation: Fiji/Taveuni, Tonga, Phuket/Aceh, Seychelles, and South Africa, for a total of 14 clusters (Fig. 3). We used a TN93 model of molecular evolution and an uncorrelated relaxed clock (Drummond et al. 2006) to model COI sequence evolution. Because this species and most other Indo-Pacific species appear to have a history of demographic fluctuation over evolutionary time, we implemented a Bayesian skyline model of demography as well (Drummond et al. 2005), which relaxes assumptions of any particular demographic history. We set a fairly uninformative truncated Poisson prior for the number of allowable migration rates within the matrix with an offset of  $k - 1 = 13$  (the minimum number of rates required to connect all populations) and a mean of 10 (95% of the prior probability mass lies between 5 and 21 rates). All other priors were set to their defaults, and we elected not to use distance-informed priors on gene flow parameters, so that the model had no a priori information about the underlying geography. The

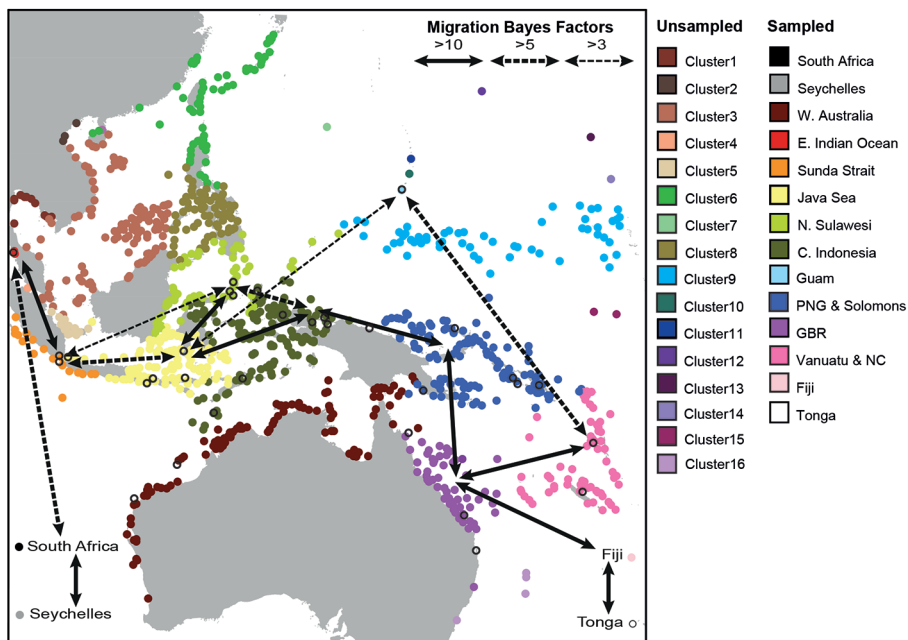


Figure 3. Best supported migration routes (Bayes Factor  $> 3$ ) among modular population clusters delineated by the biophysical model. Optimal clusters were identified using the network modularity algorithm, and are shown in different colors. Each colored point represents the geographic centroid of reef patches used in the biophysical model. Open circles show genetic sample locations, as in Figure 1.

model was run four times for at least 50 million steps of Markov chain Monte Carlo (MCMC), and convergence was checked in Tracer 1.5. We then combined the logfiles and treefiles after trimming off an appropriate amount of burn in, and constructed a maximum clade credibility tree, from which we extracted the ancestral location probabilities. We established the significance of migration rate parameters if they were supported by a Bayes Factor of 3, which also corresponded with the migration rate parameter being required to explain  $>50\%$  of sampled topologies.

Because MRCA location probability might be biased toward spatial clusters with a large sample size, we evaluated the prior distribution for root location by randomly swapping the location state among sequences during the MCMC chain such that sample size from each spatial cluster remained the same, but the sequences assigned to each location becomes random. This randomization, performed with the tip state operator in BEAST 1.7.5, as described by Edwards et al. (2011), removed location information from the dataset, allowing us to observe the prior expectation for root location.

Given that single-locus inference is still common for Indo-Pacific species (see discussion in Bowen et al. 2014), the genealogical approaches used here make much more effective use of the high information content in the mitochondrial locus than do estimates of genetic structure. Moreover,  $F_{ST}$  methods as well as more explicit population genetic models such as Migrate (Beerli and Felsenstein 2001) make an implicit assumption of genetic equilibrium (i.e., haplotypes have maintained the observed distribution for a long time about  $\frac{1}{2} N_e$  generations) that is violated by the dynamic changes in the marine habitats of the Coral Triangle over the past hundreds

of millennia. The spatial diffusion approach is not completely parameterized as a population genetic model (although  $\Theta$  is still estimated as part of the skyline model) so it cannot make estimates for the amount of gene flow among populations as is done in Migrate and IMa. However, the absence of population genetic parameters allows a more flexible model that can reconstruct the most likely location of the mitochondrial common ancestor, together with the avenues of colonization and gene flow among multiple populations without prior knowledge of population history (unlike IMa2, Hey and Nielsen 2007; see also Bloomquist et al. 2010). As with all coalescent methods, the addition of sequence data from nuclear loci will eventually allow us broaden our inference to the demographic history of the species, rather than just that of the maternal lineage.

## RESULTS

**STUDY LOCATIONS AND COI SEQUENCING.**—We obtained mtDNA sequences from a total of 791 *L. laevigata* individuals including 274 new sequences that greatly expanded the geographic scope of population sampling, especially from the western Pacific (Table 1, Fig. 1). To avoid using too much missing data by nucleotide position (<5%), we trimmed our alignment to 392 bp in length, containing a total of 110 substitution sites, and this portion of COI was used for subsequent analysis. This resulted in 209 unique haplotypes. New sequences have been deposited in Genbank (Accession Numbers KF834572–KF834833) and a complete Fasta-formatted file of the 791 individuals is deposited in <http://www.datadryad.org>.

**GENETIC DIVERSITY AND DIFFERENTIATION.**—Thirty equally parsimonious haplotype networks were recovered (one network chosen at random is shown in Fig. 2). The haplotype network highlights the high diversity of haplotypes and presence of many unique or private haplotypes. Consistent with previous mtDNA sequencing surveys (i.e., Williams 2000, Crandall et al. 2008b, Kochzius et al. 2009), there were two large emergent clusters. The haplotypes of sampled locations in the West and Central Pacific were largely restricted to one cluster (i.e., Vanuatu, New Caledonia, Tonga and parts of the Solomons, Fiji, and Papua New Guinea; top cluster, Fig. 2.) but not entirely (e.g., the sample from Taveuni in Fiji comprised individuals from both clusters). In contrast, locations sampled in the Indian Ocean had haplotypes that fell within the other cluster (bottom cluster, Fig. 2) or were intermediary to both clusters. Many haplotypes were shared across many regions, and especially within the Central Indo-Pacific locations, haplotypes from both clusters were common.

Individual populations varied in observed mtDNA diversity (Table 1) with haplotype diversity ranging from 0.42 to 0.80 in populations with  $n > 10$ . Several populations showed deviations from neutral equilibrium conditions especially as evaluated by Fu's (1997)  $F_s$  statistic. Among the 26 populations where 10 or more individuals were sampled, there was significant genetic differentiation among many population pairs regardless of the statistic used (Online Table S1). Non-metric multidimensional scaling (NMDS) based on  $F_{ST}$  values with Tamura-Nei distances among haplotypes recovered relationships among populations that roughly approximated geography (Fig. 3; non-metric stress = 0.045). Dimension 1 of the NMDS was very strongly correlated with longitude ( $R^2 = 0.81$ ,  $P < 1 \times 10^{-9}$ ), while there was no correlation of NMDS dimension 2 to latitude ( $R^2 = 0.01$ ,  $P = 0.58$ ).



All population groupings in hierarchical AMOVAs resulted in significant  $\Phi_{CT}$  and  $F_{CT}$  values, indicative of substantial regional population structure. The population grouping that consistently returned the highest  $\Phi_{CT}$  and  $F'_{CT}$  values was a two-regional grouping whereby Pacific populations (all populations east of Cenderwasih Bay) were delineated from the remaining populations including those from Cenderwasih, the central Indo-Pacific, and the Indian Ocean populations ( $\Phi_{CT} \leq 0.282$ ,  $P < 0.001$ ;  $F'_{CT} \leq 0.637$ ,  $P < 0.038$ ). For all groupings,  $\Phi_{SC}$  values were also significantly greater than zero ( $P < 0.001$ ) and  $F'_{SC}$  were marginally significant ( $P \leq 0.06$ ), indicative of genetic differentiation among populations within regions (Online Table S2).

**DISPERSAL DISTANCE PROXIES.**—The bivariate correlation between dispersal distance matrices, evaluated with the simple Mantel Test, revealed significant relationships for 20 localities within the Coral Triangle model domain. For the 190 site-pairs, the correlations between all distance-based dispersal proxies were high. The correlation coefficient for Euclidean distance and overwater distance was  $r_M = 0.996$  (10,000 permutations,  $P < 0.0001$ ). For Euclidean distance and mean dispersal distance,  $r_M = 0.896$  (10,000 permutations,  $P < 0.0001$ ). For overwater distance and mean dispersal distance,  $r_M = 0.902$  (10,000 permutations,  $P < 0.0001$ ).

The leading eigenvector community structure algorithm revealed 25 groups across the Coral Triangle (highlighted by different colors in Fig. 3), with strong consistency among dispersal distance metrics, thresholds, and community detection algorithms (not shown). Nine of these groups contained sampling sites for which we had more than 10 samples.

**DISTANCE AS A PREDICTOR OF GENE FLOW AND DIFFERENTIATION.**—Using the multiple regression on distance matrices (MRDM) approach to evaluate predictors of genetic structure showed strong positive relationships between both Euclidean and overwater distances and either  $\Phi_{ST}$  or  $D_{est}$  ( $R_M^2$  ranged from 0.46 to 0.68, for the 26 population comparisons), consistent with an isolation by distance pattern. The relationship between  $F_{ST}$  and distance was substantively lower ( $R_M^2 < 0.03$ ). Due to the collinearity of Euclidean distance and overwater distance ( $R_M^2 = 0.99$  for 26 localities) and overwater distance and dispersal distance ( $R_M^2 = 0.90$  for the 20 localities) optimal full models retained only one of these distances. Both forward and backward model selection converged on the same linear model for  $\Phi_{ST}$  or  $D_{est}$ . For  $\Phi_{ST}$ , the best model contained both Euclidean distance and a barrier to the east of Cenderwasih Bay (Fig. 4:  $R^2 = 0.69$ ,  $P < 0.001$ ), and for  $D_{est}$ , the best model contained overwater distance, the barrier to the east of Cenderwasih Bay, and the West Sumatra delineation ( $R^2 = 0.50$ ,  $P < 0.001$ ). For  $F_{ST}$ , the best model contained only Euclidean distance but did not explain much variance ( $R^2 = 0.03$ ,  $P = 0.014$ ). With the more restricted set of 20 populations for which we had mean dispersal distance predictions, overwater distance was better predictor of genetic structure than any of the other predictors for  $\Phi_{ST}$  and  $D_{est}$  ( $\Phi_{ST}: R_{OW}^2 = 0.48$ ,  $R_{Euc}^2 = 0.46$ ,  $R_{BP}^2 = 0.39$ ,  $D_{est}: R_{OW}^2 = 0.30$ ,  $R_{Euc}^2 = 0.30$ ,  $R_{BP}^2 = 0.21$ ) and  $F_{ST}$  was not well predicted regardless of distance metric ( $R^2 < 0.12$ ).

**BAYESIAN SELECTION OF IMPORTANT MIGRATION PARAMETERS.**—Four independent replicate BEAST runs converged to the same likelihood distribution after the removal of approximately 10–30 million burn-in steps from each run. The combined logfiles each contained about 107 million total steps, with high effective sample size (ESS > 200) values further indicating convergence. The analysis highlighted 14

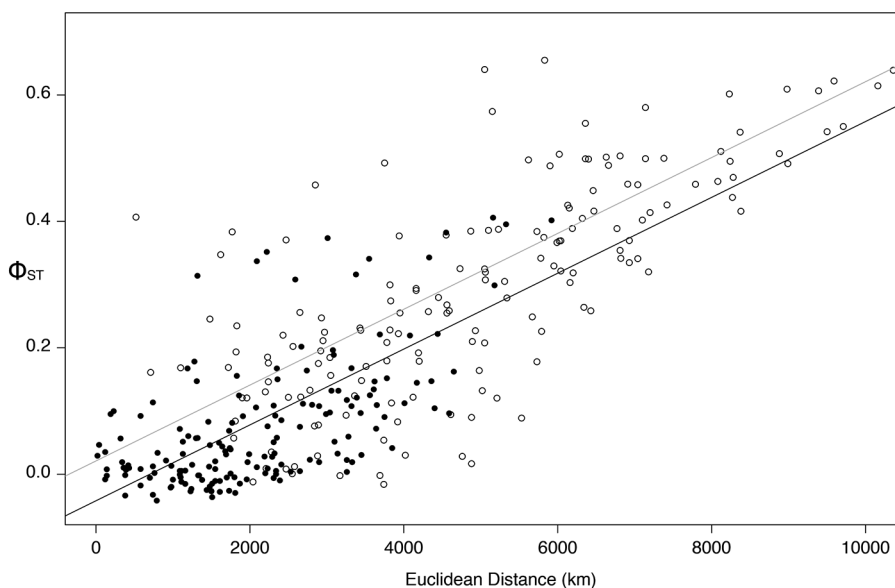


Figure 4. Best linear model of genetic differentiation. Pairwise  $\Phi_{ST}$  values by Euclidean distance and showing the effect of the division between Indian and Pacific oceans east of Cenderwasih ( $R^2 = 0.69$ ,  $P < 0.001$ ). Grey points represent population pairs including both Indian and Pacific ocean populations whereas black points represent population pairs within either ocean. Gray and black lines represent the regression lines for between and within ocean comparisons.

migration rates as having a Bayes Factor (BF) of  $>3$  (posterior odds of  $> 3:1$ ). Most of these (11/14) were between adjacent spatial clusters, even though no prior information on location was given in the analysis (Fig. 3). Regions in the central portion of the Coral Triangle had multiple connections among them. All population clusters were connected to the network by at least one significant migration parameter with the exception of Western Australia (here comprising only 7 samples from Ningaloo Reef). The best-supported migration parameter to this region came from the GBR with a BF of 2.54. Randomization of tip locations yielded a migration matrix that was also apparently random, with only five of the 22 well-supported migration parameters occurring between neighboring spatial clusters.

The probability distribution for the location of the most common recent ancestor shows a fairly flat surface across the Coral Triangle, but these probabilities were generally higher than for peripheral populations (Online Fig. 2A). However, randomization of tip locations showed that the prior expectation for the location of the root was highly correlated with sample size (Online Fig. 2A,B;  $R^2 = 0.995$ ).

## DISCUSSION

Most phylogeographic studies of Indo-Pacific species to date have relied on estimates of mitochondrial genetic structure to then make inferences about gene flow (e.g., Crandall et al. 2008b, Ackiss et al. 2013, Raynal et al. 2014, and see Keyse et al. 2014 for a review of the geographic and genetic scope of 108 such studies). Because

most such studies assume that patterns of gene flow have been held at a static equilibrium over a long period of time, it is impossible to determine from them whether this gene flow (or lack thereof) is historical or contemporary (but inferences are often made about the latter). However, when we invert our view to consider explicit models of gene flow (IBD and spatial diffusion) rather than genetic structure, we are able to see that the data contain a good deal of spatial information, even in a species with relatively good planktonic dispersal potential and low genetic structure. The fact that spatial distances explain the data better than do oceanographic distances from a biophysical model indicates that historical colonizations and subsequent gene flow events are more strongly reflected in the mitochondrial genome of *L. laevigata* than is contemporary gene flow. These results complement and extend upon earlier observations that genetic data sets from this diverse region are often haunted by the “ghosts of dispersal past” (Benzie 1999).

These novel insights into the population structure and evolution of *L. laevigata* result directly from population sampling that includes localities from the periphery of the Indian and Pacific oceans as well as from the Coral Triangle. The vast species ranges of many Indo-Pacific marine animals, such as that of *L. laevigata*, make comprehensive population genetic and phylogeographic studies difficult. Previous studies of Indo-Pacific taxa have either sampled in the Pacific and Indian oceans with limited sampling in the Coral Triangle (e.g., Lavery et al 1996, Benzie 1999, Williams and Benzie 1998), or extensively within the Coral Triangle without a broader context (e.g., Barber et al 2006, DeBoer et al. 2008, Crandall et al 2008b), albeit with some notable exceptions (e.g., Crandall 2008a, Vogler et al. 2012, 2013). Here, we are able to bring together one of the most spatially comprehensive surveys of genetic variation for any single Indo-Pacific species to date, resulting in the broad geographic context necessary for strong inference. Although our inference is confined to the history of a single locus, the extensive geographic scope of sampled populations combined with state-of-the-field analyses (reviewed in Liggins et al. 2013) as well as results from a biophysical model allow us to understand the phylogeography of this species at greater resolution than ever before.

**SIGNAL FROM THE SEASCAPE.**—The first line of evidence for a strong geographic signal in this mitochondrial data set is provided by the result from non-metric multidimensional scaling of  $\Phi_{ST}$ , which shows an extremely strong correlation with longitude ( $R^2 = 0.81$ , Fig. 5). This NMDS result is reminiscent of classic results from human population genetics (Cavalli-Sforza et al. 1994, Novembre et al. 2008), in which the first two dimensions of variation in the genetic data show a good fit to geography. However, this analysis is based on population genetic distances from a single information-rich locus, rather than PCA scores among individual genotypes (although the latter can be a special case of the former, Wang et al. 2010). Those studies demonstrated that a species (e.g., humans) that has experienced recent expansions and/or high equilibrium gene flow among neighboring populations (and thus has relatively few truly genetically distinct populations) will contain a good deal of spatial information in the two best explanatory dimensions of a multivariate analysis. Although it may be difficult to differentiate between historical and contemporary processes with this non-parametric method, our NMDS approach demonstrates that it is more fruitful to think about Indo-Pacific species in terms of gradational differentiation reflecting historical or contemporary gene flow rather than in terms of

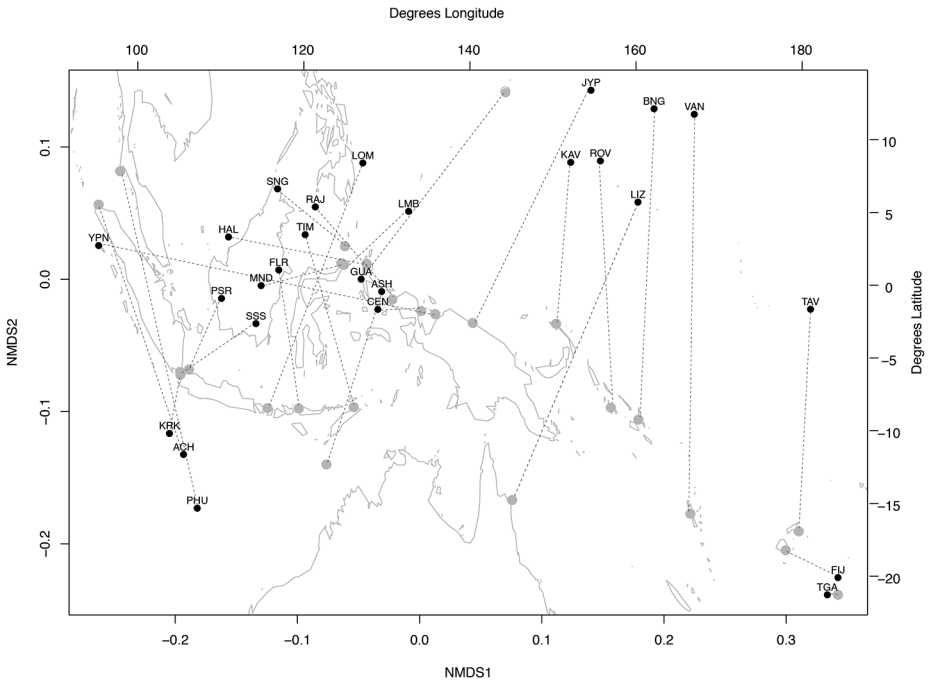


Figure 5. NMDS plots for  $\Phi_{ST}$  values from populations with sample size  $> 10$ . See Table 1 for abbreviations. The first NMDS dimension has a strong correlation with longitude ( $R^2 = 0.81$ ,  $P < 1 \times 10^{-9}$ ), so a map of geography has been underlaid for reference, with the geographical positions of each sample noted.

distinctly structured populations (see also Patterson et al. 2006; and Novembre and Stephens 2008 for in-depth discussion of underlying theory).

A second line of evidence for geographic signal in this data set is found in strong and significant MRDM correlations of population genetic distances ( $\Phi_{ST}$  or  $D_{est}$ ) with three different proxies for dispersal distance (Fig. 4). This isolation-by-distance result is consistent with a stepping-stone model of gene flow or colonization among neighboring populations. Interestingly, the two geographic distance proxies were consistently better than the modeled larval dispersal distance for explaining the contemporary pattern of genetic distances in mtDNA. This result appears to run counter to that of White et al. (2010) who found that larval dispersal distance from a biophysical model provided a much stronger explanation than geographic distance for patterns of genetic structure in microsatellites. The stronger correlation of genetic distance to geographic distance as compared to oceanographic distance in our dataset likely arises from a mismatch between the timescale over which mtDNA integrates (1000–100,000 yrs) and the timescale of contemporary larval dispersal among the geographic clusters (approximately 1–100 yrs). The genetic distances calculated from our mitochondrial data set are therefore probably capturing information about historical gene flow and colonization events rather than contemporary gene flow. Thus, mtDNA provides a valuable historical contrast to multi-locus genotyping (notably microsatellites and SNPs), which probably provide a closer fit to contemporary processes (Selkoe and Toonen 2006).

A final line of evidence comes from our phylogeographic diffusion model. By directly reconstructing the mitochondrial history in a spatial context, we are able to consider the role of colonization history and gene flow in the phylogeographic distribution of haplotypes, while simultaneously integrating over the uncertainty in the genealogy. Although we provided no prior information to the diffusion model about the relative spatial locations of each geographic cluster, it selected gene flow parameters that make intuitive sense: gene flow occurs for the most part among clusters that are geographic neighbors (Fig. 3). For example, our model found the Great Barrier Reef to be the most likely source for colonization of Western Australia (although this result did not rise above our threshold for significance), consistent with previous findings in another sea star (Vogler et al. 2013).

By starting with known sample locations and using ancestral state reconstruction to trace these locations back to the root over all possible genealogies, we were also able to arrive at a posterior probability distribution for the location of the MRCA (Online Fig. 2). While it is tempting to interpret from this distribution that the birthplace of the *L. laevigata* mitochondrial matriline was somewhere in the Coral Triangle, it does not depart significantly from a prior expectation based solely on sample size. We therefore can conclude that there is not much information about root location in this data set.

**POPULATION STRUCTURE OF *L. LAEVIGATA* IN A BROADER GEOGRAPHIC CONTEXT.**—Extensive sampling of populations of *L. laevigata* across the majority of its Indo-Pacific range revealed the presence of pronounced genetic structure, but in ways that are materially different from previous studies. Early studies of *L. laevigata* (Williams and Benzie 1996, 1997, 1998) are frequently cited as classic examples of divergence among populations of Pacific and Indian ocean marine species across the Indo-Malay-Philippine Archipelago. However, the present study shows a more nuanced picture. First, although the haplotype network delineates two large clusters of haplotypes, these are not highly divergent, regionally distinct clades. Enhanced sample size in our study has filled in previously missing haplotypes. The frequency of each cluster follows a longitudinal cline, resulting in a distinctive pattern of isolation-by-distance (Fig. 4). The multiple-regression approach implemented in MRDM considers the effects of putative barriers to dispersal simultaneously with those of isolation by distance (i.e., historical or contemporary stepping-stone gene flow). Out of four possible barriers considered in our MRDM analyses, we found that the putative barrier to the east of Cenderwasih Bay was the only barrier that, when considered together with the effects of geographic distance, provided a consistently good fit to the genetic distances. This result was also captured by our AMOVA analyses, which showed that a simple partition segregating populations to the west (Cenderwasih Bay plus Central Indonesia and Indian Ocean localities) and east (western and central Pacific localities) was a better descriptor of geographic differentiation than were partitions based on lines of disjunction associated with western Sumatra, the Sunda Shelf, the Halmahera current, or any combination of the four (Online Table 2).

Our recovery of only a single potential barrier to gene flow is distinctly different from previous work, which suggested barriers in the Sunda Strait and to the west of Cenderwasih Bay (Crandall et al. 2008b, Kochzius et al. 2009). It reflects the change in perspective provided by a larger study area and explicit consideration of the effects of stepping-stone gene flow (Meirmans 2012). While our other AMOVA partitions

were significant, and our NMDS plot shows some clustering of Indian Ocean, Central Indonesian, Papua + Great Barrier Reef, and South Pacific localities, these distinctions may be more an artifact of sampling design than of any discrete barrier to gene flow.

The proximal explanation for the significance of the genetic disjunction to the east of Cenderwasih Bay is a change in relative frequency of the Indian and Pacific clusters, with the Pacific cluster becoming dominant to the east of this barrier (see fig. 4a in Crandall et al. 2008). In a way then, the disjunction is an artifact of phylogenetic distance between the two major clusters. However, it has been shown that phylogeographic breaks such as this often come to rest in regions of low contemporary migration (Barton and Hewitt 1985). Therefore we suggest that this particular disjunction could ultimately be due to the lack of reef habitat – moving east from Cenderwasih Bay there is almost 700 km of coastline with very sparse and minimal reef habitat (Spalding et al. 2001). Whereas gene flow likely occurs occasionally across Northern Papua, there are few stepping-stone populations to facilitate the exchange of migrants across generations. The stomatopod *Haptosquilla pulchella* Miers, 1880 shows a sharp genetic discontinuity between Cenderwasih Bay and populations in Papua New Guinea, suggesting that this pattern occurs in other Indo-Pacific taxa, but in general this region is surprisingly unknown as few other studies have included samples from Cenderwasih Bay and the western Pacific (Keyse et al. 2014, and see Liu et al. 2012).

## CONCLUSIONS

The high dispersal capacity and wide range of many shallow reef Indo-Pacific marine organisms necessitate both dense and comprehensive sampling to provide the fullest phylogeographic context for each species. The present mitochondrial data set provides an example of the benefits of a spatially broadened perspective. Previously inferred barriers to gene flow turn out to be artifacts of an isolation-by-distance signature that was invisible at a smaller scale. The fact that this spatial signal is better explained by geographic distance than by oceanographic distances travelled by larvae indicates that the temporal resolution in the current dataset is low: our view of the present is obscured by the ghosts of dispersal past (sensu Benzie 1999). Future phylogeographic studies should strive to broaden their genetic perspective as well as their geographic perspective to increase temporal resolution. Nevertheless, when we consider models of gene flow rather than genetic structure we realize the importance of genetic exchange among neighboring demes as the primary reason for connectivity across a marine species' range (Crandall et al. 2012).

Detailed and broad genetic surveys such as this one usually have been beyond the scope of an individual study, or any single research group. As with most fields of science, increased collaboration and equitable sharing of data and expertise can provide a way forward (see Barber et al. 2014). We hope that the present study, which brings together data from numerous different laboratories and research efforts, will herald even greater collaborative endeavours for this region.



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