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# YEAST METABOLISM IN FRESH AND FROZEN DOUGH

A thesis presented in partial fulfilment of the requirements for the degree of

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### **ABSTRACT**

Fresh bakery products have a very short shelf life, which limits the extent to which manufacturing can be centralised. Frozen doughs are relatively stable and can be manufactured in large volumes, distributed and baked on-demand at the point of sale or consumption. With appropriate formulation and processing a shelf life of several months can be achieved.

Shelf life is limited by a decline in proofing rate after thawing, which is attributed to a) the dough losing its ability to retain gas and b) insufficient gas production, i.e. yeast activity. The loss of shelf life is accelerated by delays between mixing and freezing, which allow yeast cells the chance to ferment carbohydrates.

This work examined the reasons for insufficient gas production after thawing frozen dough and the effect of pre-freezing fermentation on shelf life. Literature data on yeast metabolite dynamics in fermenting dough were incomplete. In particular there were few data on the accumulation of ethanol, a major fermentation end product which can be injurious to yeast.

Doughs were prepared in a domestic breadmaker using compressed yeast from a local manufacturer and analysed for glucose, fructose, sucrose, maltose and ethanol. Gas production after thawing declined within 48 hours of frozen storage. This was accelerated by 30 or 90 minutes of fermentation at 30°C prior to freezing.

Sucrose was rapidly hydrolysed and yeast consumed glucose in preference to fructose. Maltose was not consumed while other sugars remained. Ethanol, accumulated from consumption of glucose and fructose, was produced in approximately equal amounts to CO<sub>2</sub>, indicating that yeast cells metabolised reductively.

Glucose uptake in fermenting dough followed simple hyperbolic kinetics and fructose uptake was competitively inhibited by glucose. Mathematical modelling indicated that diffusion of sugars and ethanol in dough occurred quickly enough to eliminate solute gradients brought about by yeast metabolism.

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## LIST OF ABBREVIATIONS

DETA dielectric thermal analysis

DM dry matter

DMA dynamic mechanical analysis

DMTA dynamic mechanical thermal analysis

DSC differential scanning calorimetry

DTA differential thermal analysis

NMR nuclear magnetic resonance

SD standard deviation

SE standard error

TAM total available monosaccharides [mmol.(100g dough)<sup>-1</sup>]

# LIST OF SYMBOLS USED

a<sub>w</sub> water activity

A area  $[m^2]$ 

 $A_{dough}$  area of dough per yeast cell [mm<sup>2</sup>]

c solute concentration [mol.L<sup>-1</sup>]

D diffusion coefficient [m².s¹]

D<sub>e</sub> estimated diffusion coefficient for ethanol diffusing in dough aqueous phase

[cm<sup>2</sup>.s<sup>-1</sup>]

 $D_q$  estimated diffusion coefficient for glucose diffusing in dough aqueous phase

 $[cm^{2}.s^{-1}]$ 

E ethanol concentration [mol.L<sup>-1</sup>]

E <sub>i</sub>	initial ethanol concentration	[mol.L <sup>-1</sup> ]
$E_{max}$	maximum ethanol concentration at which sugar uptake occurs	s [mol.L <sup>-1</sup> ]
<b>F</b> <sub>i</sub>	initial fructose concentration	[mol.L <sup>-1</sup> ]
$G_i$	initial glucose concentration	[mol.L <sup>-1</sup> ]
$G_R$	glucose consumption rate (microsystem model)	[mol.L <sup>-1</sup> .min <sup>-1</sup> ]
i	space partition point number	-
j	time partition point number	-
J	diffusion flux	[mol.m <sup>-2</sup> .s <sup>-1</sup> ]
J(x)	diffusion flux at position x	[mol.m <sup>-2</sup> .s <sup>-1</sup> ]
k	Boltzmann constant	[N.m.K <sup>-1</sup> ]
$K_i$	competitive inhibition constant	[mol.L <sup>-1</sup> ]
K <sub>m</sub>	affinity constant	[mmol.L <sup>-1</sup> ]
L	radius of the sphere of aqueous phase in the microsystem mo	odel [m]
М	molar mass	[g.mol <sup>-1</sup> ]
n	number of samples or replicates	-
$Q_{\text{st}}$	net isoteric heat of adsorption	[kJ.mol <sup>-1</sup> ]
r	surface radius of curvature	[m]
r <sub>dough</sub>	radius of the hypothetical circle of dough allocated to each ye	ast cell [mm]
R	universal gas constant	[N.m.K <sup>-1</sup> .mol <sup>-1</sup> ]
S	sugar concentration	[mmol.L <sup>-1</sup> ]
$S_A$	concentration of substrate A	[mmol.L <sup>-1</sup> ]
Sı	concentration of inhibitor	[mmol.L <sup>-1</sup> ]
t	time	[min]

		xvi
<i>t</i> <sub>n</sub>	time co-ordinate number 'n'	-
$\Delta T_f$	freezing point depression	[°C]
Τ	absolute temperature	[K]
V	rate of sugar uptake	[mmol.L <sup>-1</sup> .h <sup>-1</sup> ]
V <sub>m</sub>	mean volume	[ml]
$V_M$	molar volume	[m <sup>3</sup> .mol <sup>-1</sup> ]
$V_{max}$	maximum specific rate of sugar uptake	[mmol.(g biomass) <sup>-1</sup> .h <sup>-1</sup> ]
$V_{G}$	glucose uptake rate	[mmol.(100g dough) <sup>-1</sup> .min <sup>-1</sup> ]
$V_F$	fructose uptake rate	[mmol.(100g dough) <sup>-1</sup> .min <sup>-1</sup> ]
$\mathbf{x}_{n}$	space co-ordinate number 'n'	-
X	biomass concentration	[g.L <sup>-1</sup> ]
$Y_{ef}$	molar yield of ethanol from fructose	-
$Y_{eg}$	molar yield of ethanol from glucose	-

# **GREEK SYMBOLS**

α	ethanol inhibition coefficient	-
γ	surface tension	[N.m <sup>-1</sup> ]
η	coefficient of viscosity	[N.s.m <sup>-2</sup> ]
π	3.14159	-
ρ	density	[g.m <sup>-3</sup> ]