

Copyright is owned by the Author of the thesis. Permission is given for a copy to be downloaded by an individual for the purpose of research and private study only. The thesis may not be reproduced elsewhere without the permission of the Author.

***CLONING AND CHARACTERISATION OF TWO
SUBTILISIN-LIKE PROTEASE GENES FROM
NEOTYPHODIUM LOLII***

A Thesis presented in partial fulfilment of the
requirements for the degree of
Master of Science in Molecular Genetics
at Massey University, Palmerston North,
New Zealand

Michelle Kay McGill

2000

ABSTRACT

PCR amplification of *Neotyphodium lolii* genomic DNA with degenerate primers detected two different sequences with homology to subtilisin-like proteases. These two PCR products were used to screen a *N. lolii* Lp19 genomic library.

The *prt1* gene was isolated by screening the genomic library with the GH30 PCR product. This gene encodes a putative peptide of 434 amino acids that is most similar to subtilisin-like proteases from *Aspergillus* sp. The *prt1* gene contained a single intron, which was in a position conserved with other fungal genes. 3'RACE was used to determine the polyadenylation site for the *prt1* gene.

Repetitive DNA was a feature of both the 3' untranslated region (UTR) and sequences downstream of the *prt1* gene. Within the 3' UTR, a complex microsatellite was found extending over 50 base pairs. Downstream of the gene, a minisatellite locus of 360 base pairs in size was found, consisting of 40 copies of a 9 base pair AT-rich repeat.

Expression of *prt1* was examined in cultures with various types of carbon and nitrogen sources. Although no conclusive results could be drawn, the type of carbon and nitrogen available did have some effect on *prt1* expression. Repression of *prt1* expression was only observed in media supplemented with sucrose and glutamate.

A 500 bp fragment from the *prt1* promoter was introduced into the vector pFunGus to create a translational fusion with *gusA*. This vector, pMM9, was transformed into *Penicillium paxilli*. Although transformation frequencies were low, the transformants obtained appeared to be stable for hygromycin resistance. Expression of GUS was observed in seven out of twelve of the stable transformants. This showed that the promoter fragment in pMM9 was sufficient for expression of GUS in a heterologous system.

The *pri2* gene was isolated by screening a genomic library with the GH3 PCR product. Partial sequence has been obtained for the *pri2* gene. The *pri2* gene contains at least three introns, the first of which is conserved with *pri1*. From the sequence obtained, *pri2* encodes a peptide with strong similarity to subtilisin-like proteases from *Metarhizium anisopliae*, a fungal pathogen of insects.

ACKNOWLEDGEMENTS

Firstly, I would like to express my thanks to my supervisor, Professor Barry Scott, for his excellent supervision during my project. I have deeply appreciated your support and encouragement throughout my thesis. Thanks to Dr. Grant Hotter for his initial interest and work on this project, especially for the primer design and probe preparation. To Mike Christensen, thank you for our discussions on the endophytes and their interactions with their hosts. Your enthusiasm and knowledge about endophyte associations has been an inspiration. Thanks also to Dr. Andrew Griffiths for the endophyte genomic DNA used for Southern blotting in Figures 3.2 and 4.2, and to Dr. Peter Farley for the conversations on proteases and their regulation. Thanks also to Dr. Gretchen McCaffrey for her supervision while Barry was away on sabbatical and her encouragement throughout my degree.

To Carolyn Young, how do people in other labs cope without you? I'd probably still be library screening if it wasn't for your help. Thanks for all your help and advice for the practical work, and especially for proof-reading most of this thesis and helping me to put it all together. To Lisa McMillan, thanks for your help with the proofreading, for answering all my questions and for help learning all those new experimental procedures. To my fellow lab-members past and present, Christina, Austen, Emily, Xiuwen, Renae, Raj, Rohan, Mike and Shuguang, and the other members of MGU (Bek, Beccy, Seth, Janet, Paula and Jonathan) thanks for your help and support. To Bec, thankyou for your encouragement during the writing of this thesis, especially when I felt it would never be finished.

To my parents Pat and Teresa, I could never, ever have achieved this without both of you. Thankyou so much for all your love and support, both financial and emotional, especially during the times I would have given up without your help. To my sister Roslyn, thanks for always being there for me, and thanks also to my brothers Steven and Desmond. Special thanks must go to my flatmates for their encouragement while I have been writing. Todd, Andrew, Larnie and Shan, thanks for being great friends. You made sure life at the flat was fun and that I didn't stress out too much. Thanks for the many laughs and songs, although I'm not sure if I want to remember all of the lyrics! Thanks also to Htar for her friendship, support and encouragement.

TABLE OF CONTENTS

ABSTRACT.....	i
ACKNOWLEDGEMENTS	iii
TABLE OF CONTENTS.....	iv
LIST OF TABLES.....	xiv
LIST OF FIGURES	xv
CHAPTER ONE INTRODUCTION.....	1
1.1 FUNGAL ENDOPHYTES OF GRASSES.....	1
1.1.1 Taxonomy and evolution of <i>Neotyphodium</i> endophytes.....	4
1.2 THE MUTUALISTIC SYMBIOSIS BETWEEN GRASS AND ENDOPHYTE.....	6
1.2.1 Effects on the plant	6
1.2.1.1 Resistance to herbivores and plant pathogens	6
1.2.1.2 Effects of endophyte infection on plant growth and development	8
1.2.1.3 Improved drought tolerance	8
1.2.2 Effects on the endophyte	9
1.2.2.1 Seed dissemination of endophytes.....	9
1.2.2.2 Fungal nutrition and effects on host metabolism.....	10
1.2.2.3 Protection from the external environment.....	11
1.3 SPECIFICITY OF ENDOPHYTE-HOST INTERACTIONS	12

1.3.1	Host specificity of endophytes.....	12
1.3.2	Host resistance to endophyte infection	12
1.3.3	The physical interface between the endophyte and its host	13
1.4	FUNGAL EXTRACELLULAR PROTEINASES	14
1.4.1	Protein localisation and post-translational modifications of extracellular enzymes.....	14
1.5	SUBTILISIN-LIKE PROTEASES	15
1.5.1	Roles of proteinases in fungal nutrition and pathogenesis.....	15
1.5.1.1	<i>Roles of fungal subtilisin-like proteases in pathogenesis.....</i>	15
1.5.1.2	<i>Regulation of subtilisin-like proteases</i>	16
1.5.1.3	<i>Roles for subtilisin-like proteases in fungal nutrition.....</i>	17
1.5.2	Endophyte subtilisin-like proteases	18
1.6	AIMS OF THIS PROJECT	19
 CHAPTER TWO MATERIALS AND METHODS.....		20
2.1	FUNGAL AND BACTERIAL STRAINS, λ CLONES AND PLASMIDS..	20
2.2	GROWTH OF CULTURES.....	20
2.3	MEDIA.....	20
2.3.1	CD salts medium.....	20
2.3.2	LB medium	24

2.3.3	Potato Dextrose Agar (PD agar) and Potato Dextrose Broth (PD broth)	24
2.3.4	Regeneration medium	24
2.3.5	SOC medium	24
2.3.6	Top Agarose	24
2.3.7	Media supplements	25
2.4	BUFFERS AND SOLUTIONS	25
2.4.1	Commonly used stock solutions	25
2.4.2	DNase (RNase free)	25
2.4.3	DNase I buffer	26
2.4.4	Electrophoresis solutions.....	26
2.4.4.1	<i>SDS loading dye</i>	26
2.4.4.2	<i>1 x TAE electrophoresis buffer</i>	26
2.4.4.3	<i>1 x TBE electrophoresis buffer</i>	26
2.4.5	GUS analysis solutions.....	26
2.4.5.1	<i>GUS extraction buffer</i>	26
2.4.5.2	<i>GUS assay buffer</i>	27
2.4.6	Lysozyme	27
2.4.7	Protoplasting solutions.....	27
2.4.7.1	<i>OM buffer</i>	27
2.4.7.2	<i>PEG buffer</i>	27
2.4.7.3	<i>ST buffer</i>	27
2.4.7.4	<i>STC buffer</i>	27
2.4.8	RNase (DNase free)	27
2.4.9	SM buffer.....	27
2.4.10	Southern blot solutions.....	28
2.4.10.1	<i>10 x Denhardt's hybridisation buffer</i>	28
2.4.10.2	<i>Solution 1</i>	28
2.4.10.3	<i>Solution 2</i>	28

2.4.10.4	Solution 3.....	28
2.4.10.5	20 x SSC.....	28
2.4.10.6	2 x SSC.....	28
2.4.11	STE (100/10/1) buffer.....	28
2.4.12	STET buffer.....	29
2.4.13	TE (10/0.1) buffer.....	29
2.4.14	10 x TNE.....	29
2.4.15	Tris-Equilibrated Phenol.....	29
2.5	DNA ISOLATION.....	29
2.5.1	Isolation of DNA Fragments from Nusieve™ and Seaplaque™ Agarose Gels.....	29
2.5.2	Plasmid Isolation by the Rapid Boiling Method.....	30
2.5.3	Plasmid Isolation using the Quantum Plasmid Preparation Kit (BIO-RAD).....	30
2.5.4	Plasmid isolation using QIAGEN® Kit (Qiagen).....	30
2.5.5	Purification of phage λ DNA.....	30
2.5.6	Purification of PCR products.....	31
2.6	PURIFICATION OF DNA BY PHENOL/CHLOROFORM EXTRACTION.....	31
2.7	PRECIPITATION OF DNA WITH ETHANOL OR ISOPROPANOL.....	32
2.8	DNA QUANTIFICATION.....	32
2.8.1	Fluorometric Quantitation of DNA.....	32
2.8.2	Minigel Method for Determination of DNA concentration.....	32

2.9	RESTRICTION ENDONUCLEASE DIGESTION OF DNA	33
2.10	AGAROSE GEL ELECTROPHORESIS OF DNA	33
2.10.1	Staining and photographing gels.....	33
2.10.2	Sizing DNA fragments	34
2.11	SUBCLONING.....	34
2.11.1	DNA ligations.....	34
2.11.1.1	<i>CAP treatment of Vector DNA</i>	34
2.11.1.2	<i>SAP Treatment of Vectors</i>	34
2.11.1.3	<i>Ligations</i>	35
2.11.1.4	<i>Shotgun cloning of λ DNA fragments</i>	35
2.11.2	Transformation of <i>E. coli</i> by electroporation	35
2.11.2.1	<i>Preparation of Electro-Competent E. coli cells</i>	35
2.11.2.2	<i>Electroporation</i>	36
2.11.3	Screening of transformants.....	36
2.12	SOUTHERN BLOTTING AND HYBRIDISATION	37
2.12.1	Southern Blotting	37
2.12.2	Preparation of Labelled Probe with High Prime DNA Labelling Kit	37
2.12.3	DNA Hybridisation.....	38
2.12.4	Autoradiography	38
2.12.5	Stripping filters and blots for re-use	38
2.13	LIBRARY SCREENING BY PLAQUE HYBRIDISATION.....	38
2.13.1	Plating phage λ	39

2.13.2	Filter lifts	39
2.13.3	Hybridisation of phage λ DNA to [α - ³² P]dCTP-labelled DNA probes	39
2.14	DNA AMPLIFICATION BY THE POLYMERASE CHAIN REACTION (PCR).....	40
2.14.1	Primers.....	40
2.14.2	General PCR reaction.....	40
2.15	DNA SEQUENCING.....	42
2.15.1	Automated DNA sequencing	42
2.15.2	Sequence analysis	42
2.16	TECHNIQUE FOR WORKING WITH RNA	42
2.17	ISOLATION OF RNA FROM FUNGAL CULTURES.....	42
2.17.1	DNase I treatment of RNA	43
2.18	RT-PCR ANALYSIS OF RNA	43
2.18.1	Synthesis of cDNA using random primers.....	43
2.18.2	RT-PCR using cDNA.....	44
2.19	3' RACE ANALYSIS	44
2.19.1	Preparation of cDNA using oligo dT primer	44
2.19.2	Amplification using gene specific primers.....	44
2.20	PREPARATION OF FUNGAL PROTOPLASTS	44

2.21	TRANSFORMATION OF <i>PENICILLIUM PAXILLI</i>	45
2.21.1	Transformation of <i>P. paxilli</i>	45
2.21.2	Single spore purification of transformants	45
2.22	β -GLUCURONIDASE ASSAY.....	45
 CHAPTER 3 ANALYSIS OF THE LP19 <i>PRT1</i> GENE		47
3.1	SOUTHERN BLOTTING OF ENDOPHYTE DNA	47
3.2	LIBRARY SCREENING AND MAPPING	47
3.2.1	Library screening	47
3.2.2	Mapping of <i>prt1</i> positive clones	52
3.2.2.1	Restriction digestion of positive λ clones.....	52
3.2.2.2	Restriction mapping of λ MM30.2 and λ MM30.4	55
3.2.3	Shot gun cloning of fragments from λ MM30.4.....	55
3.3	SEQUENCE ANALYSIS	65
3.3.1	Construction of a drop out vector from pMM2	65
3.3.2	Sequencing	65
3.3.3	Analysis of the <i>prt1</i> promoter region.....	65
3.3.4	Proposed sites for transcription and translation initiation for <i>prt1</i>	71
3.3.5	Analysis of the <i>prt1</i> coding region.....	71
3.3.6	Analysis of the 3' UTR of <i>prt1</i>	73
3.3.6.1	Determination of the polyadenylation site by 3' RACE	73

3.3.6.2	<i>Composition of the 3' UTR of prt1</i>	73
3.3.7	Repetitive DNA downstream of the <i>prt1</i> gene	76
3.3.8	Proposed peptide sequence of Prt1	76
3.4	DEVELOPMENT OF A <i>PRT1</i> -GUS EXPRESSION VECTOR	79
3.4.1	Amplification of <i>prt1</i> promoter fragment	79
3.4.2	Subcloning of <i>prt1</i> promoter fragment into pFunGus	79
3.4.3	Subcloning of a <i>hph</i> selectable marker into pMM8.....	84
3.4.4	Sequencing of promoter fragment in pMM9.....	84
3.5	TRANSFORMATION OF PMM9 INTO <i>PENICILLIUM PAXILLI</i>	84
3.5.1	Preparation and transformation of <i>P. paxilli</i> protoplasts.....	84
3.5.2	GUS qualitative assays of transformants	86
3.6	EXPRESSION OF THE <i>PRT1</i> GENE IN RESPONSE TO NUTRIENTS ...	86
 CHAPTER FOUR ANALYSIS OF THE LP19 <i>PRT1</i> GENE		94
4.1	SOUTHERN BLOTTING OF ENDOPHYTE DNA	94
4.2	LIBRARY SCREENING AND MAPPING	94
4.2.1	Library screening	94
4.2.2	Mapping of positive clones	99
4.2.2.1	<i>Restriction digestion of positive λ clones</i>	99
4.2.2.2	<i>Restriction mapping of λMM3.3</i>	99
4.2.2.3	<i>Subcloning</i>	104

4.3 SEQUENCING.....	104
4.3.1 Sequencing of pMM7.....	104
4.3.2 Analysis of the <i>pri2</i> coding region.....	104
4.3.3 Features of the Prt2 peptide.....	109
4.3.3.1 <i>Comparison of prt2 to other fungal subtilisin-like proteases</i>	109
CHAPTER 5 DISCUSSION	115
5.1 SUBTILISIN-LIKE PROTEASES IN ENDOPHYTES.....	115
5.2 SUBTILISIN-LIKE PROTEASES IN MULTI-GENE FAMILIES	116
5.3 FEATURES OF THE <i>PRT1</i> GENE.....	118
5.3.1 Coding region of the <i>pri1</i> gene.....	118
5.3.2 Regulation of <i>pri1</i>	118
5.3.3 The 3'UTR and repetitive DNA downstream of <i>pri1</i>	120
5.4 FEATURES OF THE <i>PRT2</i> GENE.....	121
5.5 FUTURE WORK.....	122
APPENDIX.....	124
A1.0 PLASMID MAPS	124
A1.1 pAN7-1	124

A1.2	pCB1004.....	125
A1.3	pFunGus	126
A1.4	pGEM-T Easy	127
A1.5	pUC vectors	128
A1.6	λ GEM [®] -12.....	129
A2.0	PROBE INFORMATION	130
A2.1	Amplification of protease genes using degenerate primers.....	130
A2.2	Restriction map of pGH30 and GH30 sequence.....	131
A2.3	Restriction map of pGH3 and GH3 sequence.....	133
A2.4	Comparison of GH30 with <i>prt1</i> and GH3 with <i>prt2</i>	135
A3.0	SEQUENCE DATA.....	137
A3.1	<i>prt1</i> contig.....	137
A3.2	<i>prt2</i> contig.....	147
A3.3	Comparison of pMM9 promoter sequence with <i>prt1</i> promoter.....	151
REFERENCES.....		152

LIST OF TABLES

Table 1.1	Origin of asexual endophyte taxonomic groups.....	5
Table 2.1	Fungal and bacterial strains, λ clones and plasmids.....	21
Table 2.2	Primer sequences.....	41
Table 3.1	Genomic DNA fragments homologous to the pGH30 insert.....	49
Table 3.2	Sizes of <i>Hind</i> III fragments homologous to the pGH30 insert.....	51
Table 3.3	Restriction fragments generated by digestion of <i>prt1</i> positive λ clones.....	54
Table 3.4	Fragment sizes of λ MM30.2 restriction digests.....	57
Table 3.5	Fragment sizes of λ MM30.4 restriction digests.....	59
Table 3.6	Codon bias table for the <i>prt1</i> gene.....	72
Table 3.7	Transformation of <i>Penicillium paxilli</i>	85
Table 4.1	Genomic DNA fragments hybridising to the pGH3 insert	96
Table 4.2	Sizes of endophyte genomic <i>Hind</i> III fragments homologous to the pGH3 insert.....	98
Table 4.3	Restriction fragments of λ clones hybridising to the pGH3 insert.....	101
Table 4.4	Fragment sizes of λ MM3.3 restriction digests.....	103

LIST OF FIGURES

Figure 1.1	Life cycles of sexual and asexual grass endophytes	2
Figure 3.1	Southern analysis of the <i>prt1</i> gene	48
Figure 3.2	Southern analysis of endophyte genomic DNA homologous to the pGH30 insert.....	50
Figure 3.3	Southern analysis of <i>prt1</i> positive λ clones	53
Figure 3.4	Restriction mapping of λ MM30.2 and fragments homologous to pGH30.....	56
Figure 3.5	Restriction mapping of λ MM30.4 and fragments homologous to pGH30.....	58
Figure 3.6	Overall organisation of λ clones and sequencing strategy for the fragment homologous to the pGH30 insert.....	60
Figure 3.7	Subcloning of fragments from λ MM 30.4.....	61
Figure 3.8	Nucleotide sequence of the <i>prt1</i> gene	66
Figure 3.9	Amplification of fragments during 3'RACE	74
Figure 3.10	Sequencing of 3'RACE results	75
Figure 3.11	Minisatellite DNA downstream of <i>prt1</i>	77
Figure 3.12	Proposed peptide sequence of Prt1, a subtilisin-like protease	78
Figure 3.13	Alignment of Prt1 with other fungal subtilisin-like protease peptide sequences	80
Figure 3.14	Development of the <i>prt1</i> expression vector pMM9	83
Figure 3.15	GUS qualitative assay for <i>Penicillium paxilli</i> transformed with pMM9..	87
Figure 3.16	Gene expression in Lp19 cultures grown in CD salts only	88

Figure 3.17	Gene expression in Lp19 cultures grown in CD salts supplemented with glutamate	89
Figure 3.18	Gene expression in Lp19 cultures grown in CD salts supplemented with sucrose.....	90
Figure 3.19	Gene expression in Lp19 cultures grown in CD salts supplemented with sucrose and nitrate	91
Figure 3.20	Gene expression in Lp19 cultures grown in CD salts supplemented with sucrose and glutamate	92
Figure 3.21	Gene expression in Lp19 cultures grown in CD salts supplemented with mannitol and glutamate	93
Figure 4.1	Southern blot analysis of the <i>prt2</i> gene	95
Figure 4.2	Southern blot analysis of endophyte genomic DNA homologous to the pGH3 insert	97
Figure 4.3	Southern analysis of positive λ clones and homologous fragments to the pGH30 insert.....	100
Figure 4.4	Southern analysis of λ MM3.3.....	102
Figure 4.5	Subcloning and sequencing of the <i>prt2</i> gene	105
Figure 4.6	Partial sequence of the <i>prt2</i> gene	106
Figure 4.7	Alignment of Prt2 with peptide sequences of other fungal subtilisin-like proteases.....	110
Figure 4.8	Alignment of At1, Prt1 and Prt2 subtilisin-like proteases.....	113