

Copyright is owned by the Author of the thesis. Permission is given for a copy to be downloaded by an individual for the purpose of research and private study only. The thesis may not be reproduced elsewhere without the permission of the Author.

THE β -LACTAMASES OF THE
MARINE GENUS PHOTOBACTERIUM

A THESIS PRESENTED IN PARTIAL FULFILMENT OF THE
REQUIREMENTS FOR THE DEGREE OF
MASTER OF SCIENCE IN MICROBIOLOGY
AT MASSEY UNIVERSITY

SUSAN ELIZABETH LAMB

1981

ABSTRACT

All naturally occurring isolates of all five species of the marine genus *Photobacterium* have β -lactamase activity. In this study, the β -lactamase from the laboratory strain *P. leiognathi* 206 is fully characterised. The enzyme is constitutive and is released from the cell by osmotic shocking techniques, suggesting a periplasmic location. The enzyme is maximally active at pH 6.2 and has over 90% maximum activity at pH 7.0. The hydrolytic activity of the β -lactamase is independent of zinc ion presence. On the basis of substrate profile and inhibition studies the β -lactamase is classified as a Richmond and Sykes Class II enzyme. Crypticity tests indicated that there is no outer membrane permeability barrier to β -lactam substrates.

Comparative substrate profiles performed with the β -lactamases from three strains of each species of *Photobacterium*, indicate that the enzymes from strains of *P. leiognathi*, *P. angustum* and *P. phosphoreum* are active only on penicillins, whereas those from *P. fischeri* and *P. logei* also hydrolyze cephalosporins. This division is in agreement with an imminent taxonomic change for the latter two species. Analytical iso-electric focusing of the β -lactamases from 45 *Photobacterium* strains resulted in pI values which were not necessarily species specific and there was little correlation between the pI of the β -lactamase and its substrate profile, excepting the enzymes from *P. angustum* and *P. logei*.

Although plasmid DNA is present in many *Photobacterium* strains, conjugative transfer of β -lactamase activity from six different *Photobacterium* donors to a restrictionless *Escherichia coli* mutant was not observed. All attempts to 'cure' the bacteria of β -lactamase activity with five different curing agents, were also unsuccessful. A chromosomal location for the βla^+ gene is postulated.

DEDICATION

I would like to dedicate this thesis to my parents, Dr and Mrs R.F. Lamb, without whose limitless financial support and unceasing encouragement this work would not have been possible.

ACKNOWLEDGEMENTS

I wish to express my gratitude to Massey University and the Department of Microbiology and Genetics for allowing me to carry out post-graduate studies.

I would especially like to thank Professor D.F. Bacon (Head of Department) for all efforts made on my behalf, for always being readily available whenever a problem needed discussing and for his kindly advice.

I am very indebted to my supervisor, Dr Kathy Smith, whose help, guidance, knowledge, advice, understanding and perseverance were of untold value.

Very special appreciation goes to Dr Eric Terzaghi for the voluntary expenditure of his valuable time and for his very helpful comments and advice.

Grateful thanks go to my sister Jan, for the selfless hours spent typing the first copy and to Mrs Veronica Lobb for her excellent typing of the final copy. Special thanks also to Audrey Larsen for her superb presentation of the figures throughout this work, to the Central Photographic Unit, Massey University, for the reproduction of photographs, and to Mr Don Patrick for the fine job of binding the thesis.

I would also like to particularly acknowledge and thank the following people: -

The staff of the Department of Microbiology and Genetics for technical assistance, friendship and interesting conversations which made the laboratory working hours much pleasanter. Special thanks in this respect, go to Bronwyn Dymock.

Dr P. Berquist and D. Lane, Department of Cell Biology, Auckland University, for kindly sending me the restrictionless *Escherichia coli* mutant.

Dr M. Matthew, Glaxo Laboratories, London, for the very generous gift of nitroacetin and for her useful working notes pertaining to iso-electric focusing.

Dr R. Jolly, Department Veterinary Pathology and Public Health, Massey University, for his generosity in allowing me to use the analytical iso-electric focusing equipment.

Drs M. Hardman and J. McIntosh, Department of Chemistry, Biochemistry and Biophysics, Massey University, for their advice and comments on biochemical problems encountered.

The staff of Massey University Library for obtaining interloan requests and for their help.

TABLE OF CONTENTS

	PAGE
DEDICATION	iii
ACKNOWLEDGEMENTS	iv
TABLE OF CONTENTS	vii
LIST OF TABLES	x
LIST OF FIGURES	xiii
<u>SECTION A:</u> <u>INTRODUCTION</u>	1
1. The genus <i>Photobacterium</i>	1
1.1 The classification of <i>Photobacterium</i>	1
1.2 The ecology of <i>Photobacterium</i> strains	5
2. β -Lactamase	6
2.1 Definition of β -lactamase	6
2.2 Nomenclature	6
2.3 Classification of β -lactamase	10
2.4 Distribution of β -lactamases among bacterial genera	15
2.5 Physiology and expression of β -lactamases	16
(a) Location of β -lactamase in the bacterial cell	16
(b) The permeability barrier in Gram-negative bacteria	17
(c) Production of β -lactamases in the cell	18
2.6 Genetic determination of β -lactamases	20
2.7 The clinical importance of β -lactamases	22
3. Aims of this research	24

Cont'd..

TABLE OF CONTENTS CONTINUED

	PAGE
<u>SECTION B:</u> <u>MATERIALS</u>	25
1. Bacterial strains	25
2. Reagents and media	31
2.1 Reagents	31
2.2 Marine media	35
2.3 Terrestrial media	37
3. Antibiotics	38
<u>SECTION C:</u> <u>METHODS</u>	39
1. Cultivation and maintenance of bacterial strains	39
2. Crude enzyme preparations	42
2.1 Sonication	42
2.2 Osmotic shock treatment	42
3. Assay of β -lactamase activity	42
3.1 Standard iodometric assay	42
3.2 Detection of β -lactamase activity in solid media	47
4. Substrate profile	48
5. Inhibition studies	49
6. Induction of β -lactamase	50
7. Analytical iso-electric focusing	50
8. Slab gel Electrophoresis	52
9. Genetic determination of β -lactamase	53
9.1 Plasmid curing	53
9.2 Conjugation	54
9.3 Transformation	57

Cont'd.

TABLE OF CONTENTS CONTINUED

	PAGE
<u>SECTION D:</u> <u>RESULTS</u>	61
1. Factors affecting β -lactamase activity and assay	61
1.1 Factors affecting iodometric assay	61
(a) Effect of temperature	61
(b) Effect of pH	61
1.2 Reproducibility of the iodometric assay	64
1.3 Stability of the β -lactamase	66
(a) Effect of centrifugation temperature and resuspending media	66
(b) Effect of zinc ions on β -lactamase	68
2. Physiology and expression of β -lactamase	68
2.1 β -Lactamase production during bacterial growth	68
2.2 Inducibility of the β -lactamase from <i>P. leiognathi</i> 206	71
2.3 Cellular location of β -lactamase	82
(a) Test for extracellular enzyme	82
(b) Cell-bound β -lactamase	84
(i) Modifications to the osmotic shock method	84
(ii) β -Lactamase released by osmotic shock methods	85
(iii) Concentration of cells to increase β -lactamase yield	89
(c) Permeability barrier of the cell wall to β -lactams	89
3. Classification of β -lactamases	92
3.1 Substrate profile of the β -lactamase from <i>P. leiognathi</i> 206	94
3.2 Inhibition studies with the β -lactamase from <i>P. leiognathi</i> 206	94

Cont'd..

TABLE OF CONTENTS CONTINUED

	PAGE
<u>SECTION D:</u> <u>CONTINUED</u>	
4. The β -lactamases produced by other <i>Photobacterium</i> strains	98
4.1 Substrate profiles of 16 independent isolates of <i>Photobacterium</i>	98
4.2 Analytical iso-electric focusing of β -lactamases	100
(a) Enzyme preparations	101
(b) Staining of iso-electric focusing gels for β -lactamase activity	101
(c) Iso-electric focusing results	105
(d) Slab gel electrophoresis	108
5. The genetic determination of β -lactamases	110
5.1 Plasmid curing experiments	111
5.2 Transfer of βla^+ genes by conjugation	116
(a) Bacterial strains	116
(b) Selective media	117
(c) Conjugation results	117
5.3 Transformation of βla^+ genes	120
<u>SECTION E:</u> <u>DISCUSSION</u>	129
1. Factors affecting β -lactamase assay and activity	129
2. Production of β -lactamase during bacterial growth	132
3. Induction of β -lactamase	133
4. Cellular location of β -lactamase	135
5. The permeability barrier to β -lactams	136
6. Classification of the β -lactamase from <i>P. leiognathi</i> 206	137
7. Comparisons of the β -lactamases of <i>Photobacterium</i> strains	139
8. Genetic determination	143
<u>SECTION F:</u> <u>CONCLUSION</u>	145

LIST OF TABLES

TABLE		PAGE
I	The distinguishing properties of <i>Photobacterium</i> species	4
II	β -Lactam structures	
a	Penicillins	7
b	Cephalosporins	8
III	Classes of β -lactamases	11
IV	<i>Photobacterium</i> strains	26-30
V	β -Lactamase activity present at different temperatures of assay	62
VI	Reproducibility of the assay with different levels of β -lactamase activity	65
VII	Effect of temperature of centrifugation of bacterial cells and resuspension of the cells in different media, on cell-associated β -lactamase activity	67
VIII	β -lactamase activity in the presence of zinc ions	69
IX	Effect of growth-inhibitory β -lactam concentrations on the growth of <i>P. leiognathi</i> 206 cells	81
X	Extracellular β -lactamase from <i>P. leiognathi</i> 206	83
XI	Release of β -lactamase during osmotic shock and comparison of 'shocking' solutions	87

LIST OF TABLES CONTINUED

TABLE		PAGE
XII	β -Lactamase activity release from <i>P. leiognathi</i> 206 by osmotic shock methods	88
a	β -Lactamase activity released from <i>P. leiognathi</i> 206 cells	88
b	β -Lactamase activity and viable count of <i>P. leiognathi</i> 206 cells following each stage of the osmotic shock procedure	83
c	Effect of concentrating <i>P. leiognathi</i> 206 cells during the osmotic shock procedure on enzyme yields	90
XIII	The β -lactamase crypticity factors of several independent isolates of <i>Photobacterium</i> species	93
XIV	Substrate profile of the β -lactamase from <i>P. leiognathi</i> 206	95
XV	Inhibition of the β -lactamase from <i>P. leiognathi</i> 206 by four compounds	97
XVI	Substrate profiles of the β -lactamases from independent isolates of <i>Photobacterium</i> species	99
XVII	Levels of β -lactamase activity present in crude enzyme preparations	102
XVIII	Hydrolysis of the chromogenic cephalosporin, nitrocefin, by the β -lactamases from <i>Photobacterium</i>	104
XIX	Observations with the trial agar overlay staining technique after iso-electric focusing	106

LIST OF TABLES CONTINUED

TABLE		PAGE
XX	Growth of <i>P. leiognathi</i> 206 in the presence of curing agents	113
XXI	Plate detection of <i>P. leiognathi</i> 206 β -lactamase activity following curing treatments	114
XXII	Mitomycin C-treated <i>P. leiognathi</i> 206 selectively plated for wild-type β -lactamase activity levels	115
XXIII	Single cell resistances to β -lactams used in selective plates	
	a Benzylpenicillin	118
	b Ampicillin	119
XXIV	Results of conjugation experiments	
	a Cross of donor <i>P. leiognathi</i> 206 and recipient strain A43	121
	b Cross of donor <i>P. leiognathi</i> 206 and recipient <i>E. coli</i> PB1395	122
XXV	Characteristics of 10 independent exconjugant clones	
	a From the cross of strain 534 and A43	123
	b From the cross of strain 534 and <i>E. coli</i> PB1395	124
XXVI	Results of further conjugant experiments	
	a Crosses with <i>Photobacterium</i> donors and recipient strain P_9Sm^r	125
	b Crosses with <i>Photobacterium</i> donors and recipient <i>E. coli</i> PB1395	126

LIST OF FIGURES

FIGURE		PAGE
1	β -Lactamase hydrolysis of β -lactams	
a	β -Lactamase action on a penicillin compound	9
b	β -lactamase action on a cephalosporin compound	9
2	β -Lactamase groupings proposed by Sykes and Matthew (1976)	14
3	Growth curves of <i>Photobacterium</i> strains	
a	Spectronic 20 OD at 525 nm versus dry weight of cells	40
b	Klett optical density versus mg dry weight of cells	41
4	Optical density at 490 nm versus titration volume of sodium thiosulphate	46
5	Effect of pH on the activity of β -lactamase from <i>P. leiognathi</i> 206	63
6	β -Lactamase production during bacterial growth	70
7	Growth of <i>P. leiognathi</i> 206 in the presence of β -lactams	
a	Benzympenicillin	73
b	Ampicillin	74
c	Methicillin	75
d	Cephaloridine	76
8	β -Lactamase production by <i>P. leiognathi</i> 206 in the presence of β -lactams	
a	Benzympenicillin	77
b	Ampicillin	78
c	Methicillin	79
d	Cephaloridine	80

LIST OF FIGURES CONTINUED

FIGURE		PAGE
9	Effect of 1mM EDTA on viability of <i>P. leiognathi</i> 206 cells	86
10	Sample photograph of iso-electrically focused β -lactamases, stained with the agar overlay gel method	107
11	The iso-electric points of β -lactamases produced by strains of <i>Photobacterium</i>	109