Characterisation of an Interaction Involved in Viral Replication

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Abstract

Human rhinoviruses (HRVs) are a major cause of illness worldwide and, as members of Picornaviridae, are closely related to several other human and animal pathogens that exact a large medical and economic cost on society. Viral infections in general are particularly difficult to treat, as viruses co-opt many of the host’s own biochemical pathways, making disabling the virus without harming the host very difficult. Carefully targeted strategies are required and detailed structural information is useful, both to identify new drug targets, and to fully understand interactions. One particular protein expressed by picornaviruses is 3C protease, which is responsible for post-translational processing of the viral capsid. This protease has a cysteine as its active site nucleophile, a functionality not found in eukaryotic proteases. The unusual active site makes 3C an attractive target for pharmaceuticals. Drugs that block the proteolytic action of 3C are currently in clinical trials. In addition to its proteolytic activity, 3C protease also has another function, that of an RNA binding protein. This activity has been shown to be required during replication of the viral RNA genome. In this study, the structure of 3C protease from HRV14 is investigated using NMR and other biophysical techniques. The structural information gained from these studies is used, along with data on 3C protease RNA-binding activity acquired using solution-state NMR and SAXS data, to elucidate a structure of the 3C–RNA complex. In addition, the dynamics of the free protein and of the protein in the presence of a specific inhibitor are investigated by solution-state NMR, and the potential role of dynamics in the function of the protein is explored. Finally, potential allosteric interaction between the RNA-binding and proteolytic functions of 3C is postulated, and further interactions of 3C and the 3C–RNA complex are discussed. It is hoped that a more complete understanding of 3C and its interactions will lead to more effective treatments for picornaviral infections in the future.
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Glossary of Abbreviations

1D  one-dimensional
2D  two-dimensional
3D  three-dimensional
3C 3C protease in a previous study
3CI 3C\textsuperscript{pro} with inhibitor
3CD covalently linked 3C\textsuperscript{pro} and 3D\textsuperscript{pol}
3CIR 3CI with SLD
3D\textsuperscript{pol} 3D polymerase
BB1 \(\beta\)-barrel number 1
BB2 \(\beta\)-barrel number 2
BMRB Biological Magnetic Resonance Data Bank
BTM bis-Tris propane / MES
CD circular dichroism
CN1-3C 1st C- and N-labelled sample of 3C produced
CPMG Carr-Purcell-Meiboom-Gill
CSA chemical shift anisotropy
CSI chemical shift index
CSP chemical shift perturbation
ddH\textsubscript{2}O distilled deionised water
DLS dynamic Light Scattering
DTT dithiothreitol
HRV | human rhinovirus  
---|---  
HRV-14 | human rhinovirus isotype 14  
HRV-2 | human rhinovirus isotype 2  
HSQC | heteronuclear single quantum coherence  
IMAC | immobilised metal ion affinity chromatography  
IPTG | isopropyl-β-D-thiogalactopyranoside  
LB | Luria-Bertani broth  
MES | (2-(N-morpholino)ethanesulphonic acid  
NOE | nuclear overhauser effect  
N2-3C | 2nd N-labelled sample of 3C produced  
$R_1$ | longitudinal relaxation rate  
$R_{1p}$ | relaxation rate under a transverse spinlock  
$R_2$ | transverse relaxation rate  
RBD | RNA-binding domain  
RCI | random coil index  
RDC | residual dipolar coupling  
$R_{ex}$ | contribution to relaxation from slow exchange processes  
$R_{ex}^{CC}$ | $R_{ex}$ as determined from cross-correlation analysis  
$R_{ex}^{LS}$ | $R_{ex}$ as determined from Lipari-Szabo model-free analysis  
$S^2$ | order parameter  
$S_{ave}^2$ | average order parameter  
SDS-PAGE | sodium dodecyl sulphate polyacrylamide gel electrophoresis  
SLD | stemloop-d  
$T_1$ | characteristic longitudinal relaxation time  
$T_2$ | characteristic transverse relaxation time  
TROSY | transverse relaxation optimised spectroscopy
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