

Copyright is owned by the Author of the thesis. Permission is given for a copy to be downloaded by an individual for the purpose of research and private study only. The thesis may not be reproduced elsewhere without the permission of the Author.

MOLECULAR ANALYSIS
OF
THE LACTOSE METABOLISING GENES
FROM
LACTOCOCCUS LACTIS

*A thesis presented in partial fulfilment of the
requirement for the degree of
PhD in Biotechnology
at Massey University
New Zealand.*

SIRINDA YUNCHALARD
1993

ABSTRACT

Two lactococcal strains possessing different lactose metabolism systems were chosen for the molecular analysis of lactose metabolising genes from *Lactococcus lactis*. These were *Lactococcus lactis* ssp. *cremoris* strain H2 and *Lactococcus lactis* ssp. *lactis* strain ATCC 7962. An attempt was made to sequence a previously cloned 4.4 kb *EcoRI* fragment of pDI21 reported to encode D-tagatose 1,6-bisphosphate aldolase. A comparison of sequence data generated from this fragment with DNA sequences in the GenEMBL data base revealed that the clones provided for this study were not lactococcal DNA but were chromosomal DNA of *E. coli* which encoded genes involved in purine biosynthesis. This part of the research programme was therefore abandoned.

The aim of the second part of this study programme was to clone and characterize the β -galactosidase gene from plasmid pDI3, an uncharacterized plasmid of *Lactococcus lactis* ssp. *lactis* strain ATCC 7962. This was of interest as most lactococcal bacterial strains metabolize lactose by way of the Lac-PEP:PTS system and possess high phospho- β -galactosidase activity, whereas strain 7962 metabolizes lactose by way of the lactose permease system and possesses high β -galactosidase activity and low phospho- β -galactosidase activity. Previous plasmid curing experiments indicated that the β -galactosidase activity of strain 7962 is associated with pDI3. DNA hybridization work between pDI3 and a previously cloned DNA fragment containing the β -galactosidase gene of another Gram-positive genus, *Clostridium acetobutylicum* (*cbgA*) showed that pDI3 contains DNA sequence that is to some extent homologous to β -galactosidase sequence.

Initial experiments were carried out to confirm the involvement of pDI3 on strain 7962's β -galactosidase activity. Strain ATCC 7962 wildtype containing four plasmids and the strain cured of three other plasmids (i.e. derivative strain of 7962 containing only pDI3) exhibited a Lac⁺ phenotype, while the strain cured of all four plasmids exhibited a Lac⁻ phenotype.

As part of the mapping strategy and the attempt to clone the β -galactosidase gene from strain 7962, various fragments of pDI3 were cloned. Hybridization experiments using the cloned pDI3 fragments as DNA probes were also carried out to confirm the arrangement of pDI3 fragments. A physical map of pDI3 was constructed using the restriction enzymes *Bam*HI, *Pst*I, and *Sal*I. The size of pDI3 was confirmed to be 70 kb and it may contain some small repeat sequences. Some *Eco*RI restriction sites contained in pDI3 were also determined.

Several approaches were used to localize and identify the β -galactosidase gene on pDI3. Southern hybridizations were first carried out using the *cbgA* gene. The *cbgA* gene showed weak homology to a 4.3 kb *Eco*RI doublet from pDI3. Two redundant oligonucleotide probes were designed from the highly conserved domain of deduced amino acid sequences of the available β -galactosidase sequences from other closely related Gram-positive bacteria as well as some deduced amino acid sequences derived from β -galactosidase sequences of Gram-negative bacteria. The 4.3 kb *Eco*RI doublet and a 4.3 kb *Hind*III doublet exhibited weak homology to these probes.

On the basis of these results one of the 4.3 kb *Eco*RI fragments was subsequently cloned and transformed into a *Lac*⁻ *E. coli* strain, MC1022. The cloned 4.3 kb *Eco*RI fragment (the 4.3a fragment) was shown to cross hybridize to one of the oligonucleotide probes, but did not show β -galactosidase activity. The 4.3a kb *Eco*RI fragment was also cloned into pBR322 and transformed into another *Lac*⁻ *E. coli* host (JM109) and no β -galactosidase activity was detected.

Based on the pDI3 physical map constructed, the 4.3a kb *Eco*RI fragment was shown to overlap a 13.6 kb *Sal*I fragment. This fragment was cloned into pBR322 and the plasmid designated pSY303, and this was transformed into *E. coli* JM109. Introduction of pSY303 into *E. coli* JM109 gave a *Lac*⁺ phenotype. *Lac*⁺ phenotypes were also found for other *Lac*⁻ *E. coli* host strains including *E. coli* PB2959. It was found that pSY303 expressed β -galactosidase constitutively, however the addition of 0.1% (w/v) lactose into the medium gave a higher level of expression. An inducer for β -galactosidase, IPTG, was not required for expression. Glucose had no repression effect on β -galactosidase.

Some instability of the Lac⁺ phenotype was observed. Transformants that were initially Lac⁺ manifested phenotypic segregation into Lac⁺ and Lac⁻ colonies. Both were found to retain the intact pSY303 plasmid and there was no difference in pSY303 DNA isolated from representative Lac⁺ colonies and from representative Lac⁻ colonies. Some stable Lac⁺ colonies were observed that became dark blue. Analysis of the plasmid from these colonies showed that pSY303 had undergone a deletion. The generation of these deletion derivatives may be a consequence of small repeat sequences.

In conclusion, a physical map of pDI3 has been constructed and the β -galactosidase gene from pDI3 of *Lactococcus lactis* ssp. *lactis* strain ATCC 7962 was cloned and was found to express constitutively in *E. coli*, and at a higher level in the presence of lactose.

ACKNOWLEDGEMENTS

I wish to thank my supervisors Prof. Barry Scott, Dr Graham Manderson and Dr Chris Pillidge for their guidance throughout this study. My sincere thanks are extended to Barry for accepting me as part of his territory "Scott Base" and providing valuable advice, supervision, and instructions throughout the end of my study. Special appreciation is also extended to Graham for his encouragement and guidance especially in the preparation of this thesis. Special thanks also go to Chris for discussion of experimental work and the review of literature.

Thanks to everyone at Scott Base for the encouragement and humor provided during the day to day routines of research work, Richard, Mike, Yasuo and in particular Dianne, Carolyn and last but not least to Austen.

I gratefully acknowledge the considerable support given by senior staff at the New Zealand Dairy Research Institute and for access to the Laboratory facilities at the Institute. The support and the access to laboratory facilities given by the Biotechnology Department and the Microbiology and Genetics Department are appreciated. Thanks also go to Terri and Karen for figure preparation of this thesis.

This research was funded in part by the Ministry of Foreign Affairs and Trade for which gratitude is expressed. I am particularly grateful to Prof. Ian Watson for arranging further financial support from Massey University towards the end of the programme.

I extend a special thanks to a number of close friends throughout or in part during the course of my study: Rem, Mrs J. Kessell, and Peter together with his family. I particularly thank the late Mr. and Mrs. Riseborough who wished me well and encouraged me to persevere with my study since my arrival to New Zealand.

Finally I wish to thank my families, friends and Department of Biotechnology, Khon Kaen University, Thailand, for their love, patience, and support during the considerable distance and time that we were required to be apart.

TABLES OF CONTENTS

	Page
ABSTRACT	i
ACKNOWLEDGEMENT	iv
TABLE OF CONTENTS	v
LIST OF FIGURES	x
LIST OF TABLES	xv
ABBREVIATIONS	xviii
CHAPTER 1: LITERATURE REVIEW	1
1.1 General Introduction	1
1.2 Lactic starters	2
1.3 Characteristics of lactococci	5
1.4 Taxonomy of lactococci	6
1.4.1 Classification	6
1.4.2 Nomenclature	8
1.5 Genetics of lactococci	11
1.5.1 Genetic material arrangement	11
1.5.2 Plasmid biology	11
1.5.3 Functional properties of lactococcal plasmids	13
1.5.3.1 Lactose Metabolism	14
1.5.3.2 Proteolytic Activity	15
1.5.3.3 Citrate Utilization	18
1.5.4 Genetic transfer systems	19
1.5.4.1 Transduction	19
1.5.4.2 Conjugation	21
1.5.4.3 Transposition	24
1.5.4.4 Protoplast Transformation and Transfection	26
1.5.4.5 Protoplast fusion	27
1.5.4.6 Electroporation	28
1.6 Lactococcal gene cloning	30
1.6.1 Host-Vector systems	31
1.6.1.1 Lactococci host strains	31

	Page
1.6.1.2 Lactococcal vectors	34
1.6.2 Lactococcal gene cloning and studies	37
1.7 Carbohydrate metabolism in lactic acid bacteria	41
1.7.1 Lactose	41
1.7.2 Lactose utilization pathways in microorganisms	41
1.7.3 Lactose metabolism in lactococci	42
1.7.4 Lactose transport system	42
1.7.5 Biochemistry of lactose utilization using the PEP:PTS system	44
1.7.6 Biochemistry of lactose utilization using the lactose permease system	45
1.7.7 Lactose hydrolysing enzymes in lactococci	46
1.7.8 Molecular analysis of lactose hydrolysing genes of the Lac PEP:PTS system of lactic acid bacteria	47
1.7.9 Molecular analysis of Lactose permease/ β -galactosidase system	49
1.8 Background and aims of the study	54
1.8.1 The Research Programme:	61
CHAPTER 2: MATERIALS AND METHODS	62
2.1 Bacterial Strains and Plasmids	62
2.2 Preparation of Stock Solutions and Culture Media	62
2.3 Buffers and Solution	68
2.4 Growth and Maintenance of Bacterial Strains	70
2.5 Isolation of lactococcal DNA	71
2.5.1 Small scale preparations of plasmid DNA from lactococci	71
2.5.2 Large scale plasmid DNA preparation from lactococci	71
2.5.3 Isolation of total genomic DNA from lactococci	73
2.6 Isolation of <i>E. coli</i> DNA	73

	Page
2.6.1 Small Scale Isolation of <i>E. coli</i> plasmid DNA by Rapid boiling method	73
2.6.2 Small scale isolation of <i>E. coli</i> plasmid DNA by Rapid alkaline extraction method	74
2.6.3 Large scale isolation of <i>E. coli</i> plasmid DNA	75
2.6.4. Isolation of genomic DNA from <i>E. coli</i> .	75
2.7 Purification of DNA	76
2.7.1 Ultracentrifugation in a CsCl-buoyant density gradient	76
2.7.2 Extraction with phenol/chloroform solution	77
2.7.3 Precipitation of DNA	78
2.7.4 RNaseA treatment	78
2.7.5 Column chromatographic method	79
2.7.6 Micro-dialysis through millipore membrane	79
2.8 Determination of DNA Purity and DNA Quantitation	80
2.9 Horizontal Agarose Gel Electrophoresis of DNA	81
2.10 Determination of Molecular Weights of DNA	82
2.11 Recovery of DNA fragments from agarose Gel	82
2.12 Enzymatic Manipulation of DNA	83
2.12.1 DNA digestion with restriction endonuclease	83
2.12.2 Dephosphorylation of vector DNA	84
2.12.3 DNA ligation	84
2.12.4 Labelling DNA by using random primer	85
2.13 DNA Transfer	85
2.14 DNA-DNA Hybridization to Southern Blots	86
2.15 Hybridization of Synthetic Oligonucleotide Probe to DNA Bound to Nylon Membrane.	87
2.15.1 Labelling 5 ' End by the Forward Reaction	87
2.15.2 DNA transfer to hybridize with [γ - ³² P]-labelled oligonucleotide probe	88
2.15.3 Hybridization of DNA immobilized on nylon membrane with [γ - ³² P]ATP labelled oligonucleotide probe.	89

	Page
2.16 Transformation of plasmid DNA	89
2.16.1 CaCl ₂ transformation in <i>E. coli</i>	90
2.16.2 Electroporation in <i>E. coli</i>	90
2.16.3 Transformation of lactococci by electroporation	91
2.17 β-galactosidase enzyme assay	92
2.18 DNA Sequencing	93
2.18.1 The ExonucleaseIII unidirectional deletion procedure	93
2.18.2 Sizing of the Exonuclease III deletion subclones	94
2.18.3 Preparation of double-stranded DNA template	95
2.18.4 Alkaline denaturation of double-stranded DNA template	95
2.18.5 The annealing and labelling reaction	95
2.18.6 Gel electrophoresis in DNA sequencing	95
2.18.6.1 Preparation and assembly of glass plates	96
2.18.6.2 Preparation of gel solutions and sequencing gel	96
2.18.6.3 DNA sequencing gel electrophoresis	96
2.18.7 Assembly and analysis of sequence data	97
CHAPTER 3: SEQUENCING OF A 4.4 kb <i>Eco</i>RI FRAGMENT THAT ENCODES D-TAGATOSE 1,6-BISPHOSPHATE ALDOLASE.	98
3.1 Introduction	98
3.2 Results	98
3.3 Discussion	99
3.4 Conclusion	110
CHAPTER 4: MOLECULAR CLONING AND CHARACTERIZATION OF THE β-GALACTOSIDASE GENE FROM <i>Lactococcus lactis</i> ssp. <i>lactis</i> STRAIN ATCC 7962	111
4.1 Introduction	111
4.2 Confirmation that pDI3 encodes β-galactosidase	111

	Page
4.3 Construction of a physical map of pDI3	114
4.3.1 Restriction enzyme digestion of pDI3	114
4.3.2 Molecular cloning of pDI3 fragments	118
4.3.3 DNA hybridizations to pDI3 digests for the construction of the physical map of pDI3	124
4.3.4 Data assembly for the construction of a physical map of pDI3	135
4.4 Detection of the β -galactosidase gene of pDI3	145
4.4.1 Localization of the β -galactosidase gene of pDI3 using a heterologous probe.	145
4.4.2 Localization of β -galactosidase gene of pDI3 using an oligonucleotides probe.	146
4.5. Molecular cloning of the 4.3 kb <i>EcoRI</i> doublet of pDI3	150
4.6 Hybridization of a β -galactosidase oligonucleotide probe to the cloned 4.3 kb <i>EcoRI</i> fragments of pDI3	150
4.7 Molecular cloning of the other possible β -galactosidase fragments from pDI3	155
4.8 Expression of <i>Lactococcus lactis</i> β -galactosidase in <i>E. coli</i>	161
4.9 The instability of pSY303 in <i>E. coli</i>	168
4.10 Restriction endonuclease map of the 13.6 kb <i>SalI</i> insert of pSY303	170
4.11 Discussion	177
4.12 Conclusion	186
 CHAPTER 5: SUMMARY AND CONCLUSIONS	 187
 BIBLIOGRAPHY	 192
APPENDIX I	238
APPENDIX II	239

LIST OF FIGURES

FIGURE		Page
1	Biochemical pathways of lactose transport and lactose catabolic trait in lactococci.	43
2	Restriction endonuclease map and the locations of <i>Lac</i> and <i>Prt</i> regions of pDI21 of <i>Lactococcus lactis</i> ssp. <i>cremoris</i> strain H2.	55
3	Pathways of lactose transport and lactose utilization of <i>Lactococcus lactis</i> ssp. <i>lactis</i> strain ATCC 7962.	57
4	Restriction enzyme map of the 4.4 kb <i>EcoRI</i> of pBH501 according to Hodge (1989).	100
5	The 401-36a contig demonstrating the gelassembly of the DNA templates from both strands using the Gelassemble and Bigpicture programme of the GCG package, University of Wisconsin.	101
6	The consensus sequence of the 401-36a contig illustrating the internal <i>SphI</i> and <i>BspMII</i> restriction sites.	102
7	The alignment between the 401-36a contig and the <i>Eco purM</i> gene using the GAP programme of the GCG package, University of Wisconsin.	103

List of figures (continued)	Page
8 A Restriction enzyme map of the 4.4 kb <i>EcoRI</i> fragment of pBH501	105
8 B Restriction enzyme map of the 4.4 kb <i>EcoRI</i> fragment illustrating the location of the 401-36a contig and the <i>E.coli purM</i> gene and <i>purN</i> gene.	105
9 β -galactosidase activity of <i>Lactococcus lactis</i> ssp. <i>lactis</i> wildtype and plasmid cured derivatives.	112
10 Plasmid profiles of <i>Lactococcus lactis</i> ssp. <i>lactis</i> strain ATCC 7962 wildtype and plasmid cured derivatives.	113
11 Restriction enzyme digests of pDI3.	115
12 Single and double restriction enzyme digests of pDI3.	116
13 Restriction enzyme analysis of cloned fragments of pDI3.	121
14 Hybridization of the 1.7 kb <i>PstI</i> fragment from pSY201 to pDI3 digests.	125
15 Hybridization of the 4.2 kb <i>PstI</i> fragment from pSY202 to pDI3 digests.	128
16 Hybridization of the 6.7 kb <i>PstI</i> fragment from pSY203 to pDI3 digests.	129

List of figures (continued)	Page
17 Hybridization of the 8.3 kb <i>Pst</i> I fragment from pSY204 to pDI3 digests.	130
18 Hybridization of the 2.5 kb <i>Sal</i> I fragment from pDI3 to pDI3 digests.	132
19 Hybridization of the 5.5 kb <i>Sal</i> I fragment from pDI3 to pDI3 digests.	133
20 Hybridization of the 13.6 kb <i>Sal</i> I fragment from pDI3 to pDI3 digests.	134
21 Hybridization of the 15.9 kb <i>Sal</i> I fragment from pDI3 to pDI3 digests.	136
22 Hybridization of the 32.2 kb <i>Sal</i> I fragment from pDI3 to pDI3 digests.	137
23 Restriction endonuclease map of pDI3 isolated from <i>Lactococcus lactis</i> ssp. <i>lactis</i> strain NZDRI 5191 demonstrating restriction sites of restriction enzymes: <i>Bam</i> HI, <i>Pst</i> I, and <i>Sal</i> I.	141
24 A physical map of pDI3 isolates from <i>Lactococcus lactis</i> ssp. <i>lactis</i> strain NZDRI 5191 incorporating <i>Bam</i> HI, <i>Pst</i> I, and <i>Sal</i> I sites.	142
25 Hybridization of the 5.2 kb <i>Eco</i> RI fragment from pKH179 to pDI3 digests.	147

List of figures (continued)	Page
26 The design of the oligonucleotide probe, CP20.	148
27 Hybridization of the 20-mer oligonucleotide probe, CP20, to pDI3 digests.	149
28 The design of the oligonucleotide probe, BS13	151
29 Hybridization of the oligonucleotide probe, BS13 to cloned fragment of pDI3 and to β -galactosidase containing fragments from <i>Clostridium acetobutylicum</i> and <i>Streptococcus thermophilus</i> .	153
30 Hybridization of the 4.3 kb <i>EcoRI</i> fragment from PN1338 (pSY115) to pDI3 digests.	154
31 Hybridization of the 13.6 kb <i>SalI</i> from pDI3 to various digests of pDI3.	156
32 Hybridization of the 13.6 kb <i>SalI</i> from pDI3 to various digests of pDI3 and an <i>EcoRI</i> digest of pSY105.	159
33 Restriction enzyme comparison of pSY105 and pSY115.	160
34 β -galactosidase expression of the <i>E. coli</i> PN1357.	162
35 β -galactosidase expression of <i>E. coli</i> PB2959 containing pSY303 (<i>E. coli</i> PN1394).	165

List of figures (continued)	Page
36 Hybridization of the 13.6 kb <i>SalI</i> fragment from pDI3 to the β -galactosidase positive <i>E. coli</i> strains.	167
37 Hybridization of the 13.6 kb <i>SalI</i> insert of PN1357 to various digests of pDI3.	169
38 Single and double restriction enzyme digests of pSY303.	173
39 Restriction endonuclease map of the 13.6 kb <i>SalI</i> fragment containing the β -galactosidase gene region of pDI3 from <i>Lactococcus lactis</i> subsp. <i>lactis</i> strain NZDRI 5191 indicating the internal <i>BglII</i> , <i>EcoRI</i> , and <i>HindIII</i> sites.	176

LIST OF TABLES

TABLE		Page
1	Metabolic pathways in lactic acid bacteria.	7
2	Biochemical and chemical characteristics differentiating species and subspecies of the genus <i>Lactococcus</i>	9
3	Current <i>Lactococcus</i> species designations and their former streptococcal species names	10
4	Plasmids associated with lactose and/or protein hydrolysing activities	16
5	Lactic streptococci strains which have used or have potential to use as host in gene cloning experiments	33
6	Summary of some of the lactococcal genes cloned in both lactococcal hosts and heterologous hosts.	38
7	List of cloned phospho- β -galactosidase genes in lactic acid bacteria.	48
8	List of cloned β -galactosidase genes in lactic acid bacteria and some other bacteria	51
9	Comparison of some physical properties of the β -galactosidase enzymes encoded by the sequenced β -galactosidase genes	53
10	The specific activities of β -galactosidase in strains of <i>Lactococcus lactis</i> ssp. <i>lactis</i>	58

List of tables (continued)	Page
11 Specific activities of phospho- β -galactosidase and β -galactosidase of strains of <i>Lactococcus lactis</i> ssp. <i>lactis</i> ATCC 7962.	60
12 Bacterial strains and plasmids used in this study	63
13 Lactococcal plasmid preparation protocol	72
14 Fragments sizes generated by single and double digestions of pDI3 with <i>Bam</i> HI, <i>Eco</i> RI, <i>Pst</i> I, and <i>Sal</i> I	117
15 Predicted restriction sites in <i>Eco</i> RI, <i>Pst</i> I, and <i>Sal</i> I fragments of pDI3 as determined by examining the double digest of each	119
16 Restriction enzymes sites indentified in the cloned pDI3 fragments used to construct a physical map of pDI3	123
17 Summary of hybridization results used to construct a physical map of pDI3	126
18 Summary of hybridization results between pDI3 digests and the 13.6 kb <i>Sal</i> I fragment of pDI3	157
19 Level of β -galactosidase activity of <i>Lactococcus lactis</i> ssp. <i>lactis</i> strain 7962 in premeabilized <i>E. coli</i> cells	163

List of tables (continued)

Page

20	Summary of hybridization results between pDI3 digests and the 13.6 kb <i>SalI</i> fragment from pSY303	171
21	Fragment profiles generated from single and double digestion of pSY303 with various restriction enzymes	174

ABBREVIATIONS

kb	kilobases
bp	basepairs
kDa	kilodaltons
MDa	megadaltons
nm	nanometres
g	gravity
rpm	revolution per minute
cfu	colony forming units
Ap	ampicillin
Cm	chloramphenicol
Em	erythromycin
Tc	Tetracycline
HEPES	N-[2-hydroxyethyl]piperazine-N'-[2-ethanesulphonic acid]
IPTG	Isopropyl thiogalactopyranoside
X-gal	5-bromo-4-chloro-3-indoyl galactopyranoside
ONPG	o-nitro-phenyl- β -D-galactoside
ONP	o-nitro-phenyl
ORF	open reading frame
MW	molecular weight
PCR	polymerase chain reaction
PTS	phosphotransferase system
PS	permease system
Lac-PEP:PTS	lactose phosphoenolpyruvate dependent phosphotransferase system
P- β -gal	phospho- β -galactosidase
β -gal	β -galactosidase
strain 7962	a plasmid cured derivative of <i>Lactococcus lactis</i> ssp. <i>lactis</i> strain ATCC 7962 containing only one plasmid, pDI3. (is also referred to as strain NZDRI 5191)
SDS	sodium dodecyl sulphate
OD	optical density
A	absorbance