

Copyright is owned by the Author of the thesis. Permission is given for a copy to be downloaded by an individual for the purpose of research and private study only. The thesis may not be reproduced elsewhere without the permission of the Author.

Dynamical Modelling of the effect of Insulin-like Growth Factor 1 on Human Cell Growth

A thesis presented in fulfilment of the requirements for the degree of

Master of Science
in Mathematics
at Massey University, Albany, New Zealand

Gemma Phillips

2013

Abstract

Insulin-like Growth Factor-1 (IGF-1) plays a vital role in human growth and development. Interactions with IGF-1 receptors and IGF-1 binding proteins (IGFBPs) regulate IGF-1 function. Boroujerdi et al. (1997) published a mathematical model describing dynamic regulation of IGF-1. We extended the Boroujerdi et al. (1997) model to evaluate the role of cyclic Gly-Pro (CGP) in dynamic regulation of IGF-1 function. Recent research from the Liggins Institute suggests that a metabolite of IGF-1, CGP, may have a role in regulating IGF-1 homeostasis, possibly through competitive binding to IGFBPs.

The goal of the research was to understand the kinetics of IGF-1, IGFBPs and CGP, along with their interactions with IGF-1 receptors. This goal and an understanding of how the kinetics mediate IGF-1 function was achieved through consideration of the nonlinear dynamics of the physiology using a modelling approach.

The resulting models were directly focused on three central theories. The first is that CGP can either inhibit, stimulate or maintain IGF-1 function based on the extent of receptor binding. The other theories are that CGP regulates IGF-1 through competitive binding to IGFBPs and that CGP does not directly interact with the IGF-1 receptors.

Four *in vitro* models were developed and fitted to experimental data. These included two implicit models which relied on two feedback terms in the equations. The second model was an alteration of the first to produce a reduction in cell number levels for high doses of CGP added to the system. The other two models were explicit models, the first of which could not express the IGF-1 dynamics well (it showed no CGP response). Although the models incorporated these theories, there are other mechanisms influencing the system which will have an effect on the data. Therefore the fourth model was introduced as a simplified version of the third. This was aimed at resembling cell culture situations more closely and was designed to have the receptor bound IGF-1 dependent on IGF-1 and CGP production rates.

The models can be used to predict cellular response in an *in vitro* situation, or as a basis for further research in this field.

Acknowledgements

I would like to thank my supervisors, Professor Graeme Wake and Dr Paul Shorten for their constant advice, encouragement and support throughout this project.

Prof. Wake and Dr. Shorten were funded to supervise this thesis via the Gravida National Centre for Growth and Development.

Thank you to Dr Jian Guan (Head Biologist at Liggins Institute) for her feedback, guidance and biological explanations.

In addition I would like to express my gratitude towards Mr Tony Pleasants and the Norman F Barry Foundation, Wake's Scientific Consulting and Dr Guan the head of Neuroscience from Liggins Institute (Ministry of Business and Employment, MIBIE) for funding this project.

Lastly I would like to express my appreciation to my family, Alistair Watt, and his family for their understanding, love and support.

List of Figures

1	Schematic diagram of the IGF-1 receptor.	3
2	Lab pictures of Steve Moon at Liggins Institute, washing the cells . . .	7
3	Mini-PROTEAN Tetra system used for western blotting	8
4	Model diagram reprinted from Boroujerdi et al. (1997)	11
5	Model simulations for subject one reproduced from Boroujerdi et al. (1997) (left) along side the replicated model simulations (right)	15
6	Bifurcation analysis showing the effect of IGF-1 production rate, R_a , on free plasma IGF-1, q_2 . (Boroujerdi et al. (1997) model)	17
7	Bifurcation analysis showing the effect of IGFBP (50kDa) production rate, $R_{a,1}$, on the amount of free IGFBPs (50kDa), q_1 . (Boroujerdi et al. (1997) model).	17
8	Bifurcation analysis showing the effect of IGFBP-3 (150kDa) production rate, $R_{a,2}$, on the amount of free IGFBP-3 (150kDa), q_4 . (Boroujerdi et al. (1997) model).	18
9	Bifurcation analysis showing the effect of IGF-1 production rate, R_a , on free plasma IGF-1, q_2 . (Boroujerdi et al. (1997) model, $-k_{02}q_2$ term included)	21
10	Model diagram reprinted from Mizuno et al. (2001)	23
11	Reproduced Mizuno et al (2001) time series plots (left) alongside the replicated results (right)	24
12	<i>In vivo</i> model diagram	27
13	Feedback term, F, in the implicit <i>in vivo</i> model	29
14	Feedback term, G, in the implicit <i>in vivo</i> model	29
15	Bifurcation analysis showing q_4 vs a_1 (fixed low a_2 value).	33
16	Bifurcation analysis showing q_4 vs a_1 (fixed medium a_2 value).	33
17	Bifurcation analysis showing q_4 vs a_1 (fixed high a_2 value).	34
18	Bifurcation analysis showing q_4 vs a_a (fixed low a_1 value).	34
19	Bifurcation analysis showing q_4 vs a_2 (fixed medium a_1 value).	35
20	Bifurcation analysis showing q_4 vs a_2 (fixed high a_1 value).	35
21	Bifurcation analysis showing q_4 vs negative a_1	36
22	Enlarged view of the Hopf bifurcation when looking at q_4 vs negative a_1	37
23	Bifurcation analysis of q_4 vs negative a_2	38
24	Enlarged view of the Hopf bifurcation when looking at q_4 vs negative a_2	38
25	Bifurcation analysis showing q_1 against $R_{a,1}$ (<i>In Vivo</i> Model)	39

26	Bifurcation analysis showing free q_2 against $R_{a,2}$ (<i>In Vivo</i> Model).	39
27	Bifurcation analysis showing q_5 against $R_{a,5}$ (<i>In Vivo</i> Model).	40
28	Bifurcation analysis showing q_6 against $R_{a,6}$ (<i>In Vivo</i> Model).	40
29	Diagram of the first <i>in vitro</i> model (implicit)	43
30	Equation one fitted to IGF-1 only data ($R^2=0.69$)	44
31	Equation two fitted to IGF-1 only data ($R^2=0.71$)	45
32	Equation three fitted to IGF-1 only data ($R^2=0.82$)	45
33	<i>In vitro</i> model one predictions for IGF-1 only treatments (left) and IGF-1 only treatment data	51
34	<i>In vitro</i> model one predictions for CGP only treatments (left) and CGP only treatment data	51
35	<i>In vitro</i> model one predictions for combination one treatments (left) and combination one treatment data	52
36	<i>In vitro</i> model one predictions for combination two treatments (left) and combination two treatment data	52
37	<i>In vitro</i> model two predictions for IGF-1 only treatments (left) and IGF-1 only treatment data (right)	55
38	<i>In vitro</i> model two predictions for CGP only treatments (left) and CGP-1 only treatment data (right)	55
39	<i>In vitro</i> model two predictions for combination one treatments (left) and combination one treatment data (right)	56
40	<i>In vitro</i> model two predictions for combination two treatments (left) and combination two treatment data (right)	56
41	<i>In vitro</i> model three (explicit) diagram	59
42	Graph of BSA influence over IGF-1 binding with and without CGP. Data provided by Dr Jian Guan (Liggins Institute). The red line is BSA treatment with CGP (equimolar with IGF-1) and the blue line is BSA treatment without CGP.	60
43	<i>In vitro</i> model three predictions for IGF-1 only treatments (left) and IGF-1 only treatment data (right).	64
44	<i>In vitro</i> model three predictions for CGP only treatments (left) and CGP only treatment data (right).	64
45	<i>In vitro</i> model three predictions for combination one treatments (left) and combination one treatment data (right).	65
46	<i>In vitro</i> model three predictions for combination two treatments (left) and combination two treatment data (right).	65

47	<i>In vitro</i> model four (explicit simplified model) diagram	67
48	<i>In vitro</i> model four predictions for IGF-1 only treatments (left) and IGF-1 only treatment data (right).	70
49	<i>In vitro</i> model four predictions for CGP only treatments (left) and CGP only treatment data (right).	70
50	<i>In vitro</i> model four predictions for combination one treatments (left) and combination one treatment data (right).	71
51	<i>In vitro</i> model four predictions for combination two treatments (left) and combination two treatment data (right).	71
52	<i>In vitro</i> model four (simplified explicit) diagram with antibody treatment	72
53	Model testing results for IGF-1 and CGP treatments with no antibodies data plot (left) next to model predicted values (right), $R^2=0.6890$. . .	75
54	Model testing results for IGF-1 and CGP treatments with antibodies data plot (left) next to model predicted values (right), $R^2=0.7815$. . .	75
55	Model simulations reproduced from Boroujerdi et al. (1997) for subjects 2-4.	86
56	Replicated model simulations for subjects 2-4.	87
57	Bifurcation analysis for \mathbf{q} variables against IGF-1 production rate, R_a in $\text{nmol min}^{-1} \text{L}^{-1}$ (Boroujerdi et al. (1997) model)	88
58	Bifurcation analysis for \mathbf{q} variables against IGFBP (50kDa) production rate, $R_{a,1}$ in $\text{nmol min}^{-1} \text{L}^{-1}$ (Boroujerdi et al. (1997) model)	89
59	Bifurcation analysis for \mathbf{q} variables against IGFBP-3 (150kDa) production rate, $R_{a,2}$ in $\text{nmol min}^{-1} \text{L}^{-1}$ (Boroujerdi et al. (1997) model) . . .	90
60	Bifurcation analysis for \mathbf{q} variables against IGF-1 production rate, R_a in $\text{nmol min}^{-1} \text{L}^{-1}$ (Boroujerdi et al. (1997) model with $-k_{02}q_2$ term) . . .	91
61	Bifurcation analysis for \mathbf{q} variables against IGFBP production rate, $R_{a,1}$ in $\text{nmol min}^{-1} \text{L}^{-1}$ (<i>In Vivo</i> Model)	92
62	Bifurcation analysis for \mathbf{q} variables against IGF-1 production rate, $R_{a,2}$ in $\text{nmol min}^{-1} \text{L}^{-1}$ (<i>In Vivo</i> Model)	93
63	Bifurcation analysis for \mathbf{q} variables against receptor production rate, $R_{a,5}$ in $\text{nmol min}^{-1} \text{L}^{-1}$ (<i>In Vivo</i> Model)	94
64	Bifurcation analysis for \mathbf{q} variables against CGP production/infusion rate, $R_{a,6}$ in $\text{nmol min}^{-1} \text{L}^{-1}$ (<i>In Vivo</i> Model)	95
65	Example of a stable spiral equilibrium, before the Hopf bifurcation point at $R_a=1 \text{ nmol min}^{-1}$ (negative complex eigenvalue)	100

66	Example of a Hopf bifurcation at the bifurcation point with zero amplitude at $R_a=5 \text{ nmol min}^{-1}$ (purely imaginary eigenvalues)	100
67	Example of an unstable spiral equilibrium, after Hopf bifurcation point at $R_a=10 \text{ nmol min}^{-1}$ (positive complex eigenvalue). The amplitude increases until it reaches a constant amplitude (reaches the limit cycle). .	101
68	Example of a Hopf bifurcation diagram	101
69	Example of a local minimum compared to a global Minimum	105

List of Tables

1	The four types of treatments used for parameter calibration	9
2	The two types of treatments used for validation	9
3	Variable definitions reproduced from Boroujerdi et al. (1997)	10
4	Parameter definitions reproduced from Boroujerdi et al. (1997)	11
5	Data table to find initial variable values from Boroujerdi et al., 1997). . .	13
6	Subject one initial values from Boroujerdi et al. (1997)	14
7	Subject one parameter values from Boroujerdi et al. (1997)	14
8	Glossary of <i>in vivo</i> variables from Mizuno et al. (2001)	22
9	Glossary of <i>in vivo</i> parameters from Mizuno et al. (2001)	22
10	Glossary of the new <i>in vivo</i> model variables and parameters	26
11	<i>In vivo</i> parameter and initial variable values estimated from the Boro- jerdi et al. (1997) article	30
12	Final parameter values estimated from the Boroujerdi et al. (1997) model and a guess and check method which produced steady state values as close as possible to the estimates.	31
13	Steady state values found using estimated parameters from Boroujerdi et al. (1997) model and then a guess and check method involving XPPaut. .	32
14	Glossary of the implicit <i>in vitro</i> model one and two parameters	42
15	Glossary of the implicit <i>in vitro</i> model one and two variables	43
16	Equation one variance covariance matrix (A=3830.48 and B=39152.89)	46
17	Equation two variance covariance matrix (A=71142.26 and B=0.17)	46
18	Equation three variance covariance matrix(A=59596.03 and B=2.22 and C=28751.95)	46
19	Parameter and initial values which are assumed for the <i>in vitro</i> model one (Implicit) based on Tables 11-13	48
20	Estimated parameter and initial values before being fit to data for the <i>in vitro</i> model one (Implicit).	48
21	Type of data used for fitting parameters (molar mass of IGF is approx 7649g/mol)	49
22	Final parameter values for <i>in vitro</i> model one which are assumed (top) and parameter values after the equations were fit to data (bottom). . .	50
23	Initial values of <i>in vitro</i> model one (implicit) which were assumed (top) and initial values after being fit to data (bottom).	50

24	Final parameter values for <i>in vitro</i> model two (implicit) with quadratic q_6 term added, which were assumed (top) and parameter values which were fit to data (bottom).	54
25	Initial values for <i>in vitro</i> model two (implicit) with quadratic q_6 term added, which were assumed (top) and initial values which were fit to data (bottom).	54
26	Glossary of the explicit <i>in vitro</i> model three and four variables	58
27	Glossary of the explicit <i>in vitro</i> model three and four parameters	59
28	Assumed parameter and initial values for the <i>in vitro</i> model three (Explicit)	62
29	Parameter and initial value estimates for the third <i>in vitro</i> model (explicit)	62
30	Parameter values for the third <i>in vitro</i> model (explicit) which were assumed (top) and parameter values which have been fit to data (bottom).	63
31	Initial values for the third <i>in vitro</i> model three (explicit) after being fit to data.	63
32	Parameter values for the fourth <i>in vitro</i> model (explicit) which were assumed (top) and parameter values after being fit to data (bottom) . .	69
33	Initial values for the fourth <i>in vitro</i> model (explicit) which were assumed (top) and initial values after being fit to data (bottom).	69
34	Type of data used for testing <i>in vitro</i> model four	73
35	Subject two parameter and initial values reproduced from Boroujerdi et al. (1997)	83
36	Subject three parameter and initial values reproduced from Boroujerdi et al. (1997)	84
37	Subject four parameter and initial values reproduced from Boroujerdi et al. (1997)	85

Abbreviations

BSA	Bovine serum albumin
CGP	Cyclo-glycyl-proline or Cyclic Gly-Pro
DE	Differential equation
DKP	Diketopiperazine
EDTA	Ethylenediaminetetraacetic acid
GPE	Glycine-proline-glutamate
IGF-1	Insulin-like Growth Factor 1
IGF-1R	Insulin-like Growth Factor 1 Receptor
IGFBP	Insulin-like Growth Factor Binding Proteins
kDa	Kilo Dalton (Dalton is a unit of mass based on molecular mass)
nM	$\text{nmol } L^{-1}$
ODE	Ordinary Differential Equation
PBS	Phosphate Buffered Solution
PDE	Partial Differential Equation
RK4	Runge Kutta fourth order method
SDS	Sodium Dodecyl Sulfate
SSE	Sum of Squares of Errors
WST-1	Water Soluble Tetrazolium Salt-1

Contents

1	Introduction	1
1.1	Objective of this Study	1
1.2	Thesis Outline	2
2	Biological Background:	2
2.1	Introduction	2
2.2	Data Collection Methods	5
2.2.1	<i>In vivo</i> data collection methods	5
2.2.2	<i>In vitro</i> data collection methods	6
2.3	Types of Data Collected	8
3	Previous Research:	10
3.1	Introduction	10
3.2	Boroujerdi et al. (1997) Model	10
3.3	Boroujerdi et al. (1997) Equations	12
3.4	Results from the Boroujerdi et al. (1997) Model	13
3.5	Extended Analysis of the Boroujerdi et al. (1997) Model	16
3.5.1	Bifurcation Analysis	16
3.6	Altered Boroujerdi et al. (1997) Model	19
3.6.1	Altered Boroujerdi et al. (1997) model equations	19
3.6.2	Bifurcation analysis of the altered Boroujerdi et al. (1997) model	21
3.7	Mizuno et al. (2001) Model	22
3.8	Mizuno et al. (2001) Equations	23
3.9	Results from the Mizuno et al. (2001) Model	24
3.10	Discussion	25
4	<i>In Vivo</i> Model:	26
4.1	<i>In Vivo</i> Model Introduction	26
4.2	<i>In Vivo</i> Model Equations	27
4.3	<i>In Vivo</i> Model Results	30
4.3.1	Bifurcation analysis of a_1 and a_2	32
4.3.2	Bifurcation analysis of \mathbf{q} variables	39
4.4	<i>In Vivo</i> Model Discussion	41

5	<i>In Vitro</i> Model One and Two (Implicit):	42
5.1	<i>In Vitro</i> Model One Introduction	42
5.2	<i>In Vitro</i> Model One Equations	44
5.3	<i>In Vitro</i> Model One Parameter Estimates	47
5.4	<i>In Vitro</i> Model one Results	49
5.5	<i>In Vitro</i> Model Two Introduction	53
5.6	<i>In Vitro</i> Model Two Equations	53
5.7	<i>In Vitro</i> Model Two Results	54
5.8	Discussion of <i>In Vitro</i> Model One and Two (Implicit)	57
6	<i>In Vitro</i> Model Three and Four (Explicit):	58
6.1	<i>In Vitro</i> Model Three Introduction	58
6.2	<i>In Vitro</i> Model Three Equations	61
6.3	<i>In Vitro</i> Model Three Parameter Estimates	62
6.4	<i>In Vitro</i> Model Three Results	63
6.5	<i>In Vitro</i> Model Four Introduction	67
6.6	<i>In Vitro</i> Model Four Equations	68
6.7	<i>In Vitro</i> Model Four Results	69
6.8	Testing of <i>In Vitro</i> Model Four Against Independent Data Set	72
6.9	Discussion of <i>In Vitro</i> Model Three and Four (Explicit)	76
7	Conclusion	78
	References	81
	Appendix A Subject 2-4 Parameter Values Boroujerdi et al. (1997):	83
A.1	Subject 2	83
A.2	Subject 3	84
A.3	Subject 4	85
	Appendix B Graphs:	86
B.1	Comparison of Boroujerdi et al. (1997) Simulations	86
B.2	Bifurcation Diagrams	88
B.2.1	Boroujerdi et al. (1997) Original Model Bifurcation Diagrams	88
B.2.2	Boroujerdi et al. (1997) Model ($-k_{02}q_2$ added) Bifurcation Diagrams	91
B.2.3	<i>In Vivo</i> Model Bifurcation Diagrams	92

Appendix C Basic Mathematical Background of Methods used in this

Research:	96
C.1 Introduction	96
C.2 Ordinary Differential Equations	96
C.3 Stability of the system	98
C.4 Explanation of Bifurcations	99
C.5 Law of Mass Action and Mass Action Kinetics	102
C.6 Nonlinear Least Squares	104
C.7 Variance-Covariance Matrix	108