Copyright is owned by the Author of the thesis. Permission is given for a copy to be downloaded by an individual for the purpose of research and private study only. The thesis may not be reproduced elsewhere without the permission of the Author.

Stimuli Sensitive Polysaccharide Based Hydrogels as Colon Targeted Drug Delivery Vehicles.

A thesis submitted in partial fulfilment of the requirements for the degree of

Doctor of Philosophy in Chemistry

Massey University, Turitea Campus, Palmerston North, New Zealand

> Iman Kavianinia 2014



ABSTRACT

Stimuli Sensitive Polysaccharide Based Hydrogels as Colon Targeted Drug Delivery Vehicles.

By Iman Kavianinia

Supervised by Professor David R.K. Harding ^a, Associate Professor Paul G. Plieger ^a and Professor Nadia G. Kandile ^b

^a Institute of Fundamental Sciences, College of Sciences,
 Massey University, Palmerston North.
 ^b Department of Chemistry, Faculty for Women, Ain Shams University, Heliopolis,
 Cairo, Egypt

Administering drugs orally is by far the most widely used route of administration that will help eliminate the pain caused by injection, psychological barriers associated with multiple daily injections and possible infection from injection sites. However, it is important for oral drug administration to overcome several different obstacles during the delivery through the gastrointestinal tract. The barriers can be morphological barriers and physiological factors such as a wide range of pH and enzymatic activities. The lower water content and fluid mobility of the colon, which leads to longer retention times and also lower proteolytic activity of colon compared to other areas of the gastrointestinal tract, make the colon an ideal site for both systemic and local delivery of drugs. Therefore aggressive research efforts have recently focused on development of new strategies for delivering drugs to the colon.

As a drug delivery systems, hydrogels have received increasing attention due to their outstanding merits. Among the various hydrogels, including natural, synthetic and natural/synthetic hybrid hydrogels, chitosan has attracted significant attention in a broad

range of pharmaceutical and biomedical applications. Chitosan is a hydrophilic polyelectrolyte heteropolysaccharide composed of randomly (1→4)-linked 2acetamido-2-deoxy-β-D-glucopyranose 2-amino-2-deoxy-β-D-glucopyranose and linked by $(1\rightarrow 4)$ - β -glycosidic bonds. Unlike most known bioadhesive polymers, chitosan displays unique pharmaceutical and biomedical applications due to the large number of hydroxy and amino groups on the backbone of chitosan. These functional groups can be readily modified. This study was commenced with the aim of engineering a carrier with high enough physicochemical stability to reach the colon and to be able to protect a drug from various obstacles throughout the gastrointestinal tract. In this study, a new generation of chitosan derivatives was developed. Furthermore, their viability was investigated for potential applications as drug carriers to the colon. Chitosan based films with improved physical properties from introducing a cyclic imide moiety into the chitosan matrices was developed and characterised. Mechanical, thermal and chemical analyses of these films show that the heterocyclic imide linkage imparts excellent thermal, mechanical and chemical stability to the chitosan film. Additionally, spray dried chitosan microspheres with improved mechanical stability were examined for the controlled drug release of bovine serum albumin as a model protein drug. Additionally, a novel generation of amphoteric crosslinked chitosan derivatives was designed to be pH sensitive and bacterially degradable. Tabletted carriers were designed to protect the drug from the harsh acidic environment of the stomach and the rigorous enzymic activity of the small intestine and deliver the drug to the colon. Tabletted formulation forms of these novel amphoteric derivatives of chitosan showed the excellent potential formulations as colon specific drug delivery vehicles.

ACKNOWLEDGEMENTS

The study described in this thesis was carried out at the Department of Chemistry and at the Institute of Fundamental Science (IFS), Massey University, New Zealand.

I would like to express my sincere appreciation to my chief supervisor, Professor David R.K. Harding, for accepting me as a student and providing all kinds of support throughout my graduate years. His enthusiasm, passion and encouragement will definitely influence my career. His valuable advice, from professional to personal levels, will never be forgotten. I am very grateful to have had the opportunity to learn under his guidance.

I would like to thank my co-supervisors Associate Professor Paul G. Plieger and Professor Nadia G Kandile for all their guidance throughout the length of this project. I cannot thank you enough for all your unlimited support and encouragement, and for all the productive discussions.

I would also like to thank Mr. David Lun for all the technical support and assistance with the initial training on the HPLC, MALDI and ESI-MS. I would also like to pass my gratitude to Dr Pat Edwards for his help when using the NMR and advice for structural analysis. I also wish to thank Dr Jason Hindmarsh, Institute of Food, Nutrition & Human Health, Massey University, for his assistance and providing access to the solid-state NMR spectrometer. I am also very thankful to Professor Geoff Jameson, for helping me carry out the X-ray analysis and for his voluble assistance in interpreting the results. I would like to thank my office mate Dr. Olekile Tibe, for his friendship, discussions, cooperation and support.

I would like to show my gratitude and sincerest thankfulness to Dr. Nick Cave and his research group members, Dr Magda Dunowska and Mrs Gaya Gopakumar, Institute of Veterinary, Animal & Biomedical Sciences.

I thank all the members of IFS for all the friendship and for making my days more cheerful and also for the funding me to attend conferences in Spain and Australia.

Finally, I wish to extend a big thank you to my family for their continuous support throughout project, you are my sources of strength and inspiration.

TABLE OF CONTENTS

Abstra	act		i
Ackno	wledge	ments	.iii
Table	of Cont	ents	v
List of	f Tables		xiv
List of	f Figures	S	ΧV
List of	f Abbrev	viations and Symbolsxx	kiii
		ations	
2150 0	- Poolite		
Chap	oter Oı	ne: Introduction	
1.0 H	ydrogel		1
1.1 Cl	assifica	tions of hydrogels	2
1.1.1		ding to source	
		Natural hydrogels	
		Synthetic hydrogels	
	1.1.1.3	Hybrid hydrogels	
1.1.2	Accord	ding to the method of preparation	4
	1.1.2.1	Homopolymers	
	1.1.2.2	Copolymers	
	1.1.2.3	Interpenetrating polymer network hydrogels	
1.1.3	Accord	ling to ionic charge	4
	1.1.3.1	Non-ionic hydrogels	
	1.1.3.2	Ionic hydrogels	
		Ampholytic Hydrogels	
1.1.4	Accord	ding to the biodegradability	5
	1.1.4.1	Biodegradable hydrogels	
	1.1.4.2	Non-biodegradable hydrogels	
		Ampholytic Hydrogels	
1.1.5	Accord	ding to method of crosslinking	6
	1.1.5.1	Physically crosslinked hydrogels	
		Chemically crosslinked hydrogels	
1.1.6	Accord	ding to physical properties	7
	1.1.6.1	Conventional hydrogels	
	1.1.6.2	Stimuli responsive hydrogels "Smart hydrogels"	

1.2 St	imuli responsive hydrogel classification	8
1.2.1	Chemical responsive hydrogels	8
	1.2.1.1 pH responsive hydrogels	
	1.2.1.2 Glucose responsive hydrogels	
1.2.2	Physical responsive hydrogels	10
	1.2.2.1 Pressure responsive hydrogels	
	1.2.2.2 Temperature responsive hydrogels	
	1.2.2.3 Ultrasound responsive hydrogels	
	1.2.2.3 Field responsive hydrogels	
	1.2.2.3 Light responsive hydrogels	
1.2.3	Biochemically responsive hydrogels	12
	1.2.3.1 Antigen responsive hydrogels	
	1.2.3.2 Enzyme responsive hydrogels	
	1.1.3.3 Ultrasound responsive hydrogels	
	1.1.3.3 Field responsive hydrogels	
	1.1.3.3 Light responsive hydrogels	
1.3 Bi	iomedical applications of stimuli-responsive hydrogels	13
1.3.1	Applications of hydrogels in tissue engineering	13
1.3.2	Applications of hydrogels in wound healing	14
1.3.3	Applications of hydrogels in drug delivery	15
	1.3.3.1 Ocular drug delivery	
	1.3.3.2 Rectal drug delivery	
	1.3.3.3 Subcutaneous delivery	
	1.3.3.4 Transdermal delivery	
	1.3.3.5 Oral drug delivery	
1.4 Si	te specific drug delivery	20
1.4 H	uman gastrointestinal physiology	20
1.5.1	Anatomy and physiology of the stomach	21
1.5.2	Anatomy and physiology of the small intestine	22
1.5.3	Anatomy and physiology of the large intestine	22
1.6 C	olon specific drug delivery	24
1.6.1	Anatomy and physiology of colon	25
1.6.2	Factors affecting in the design of colon-specific drug delivery system	25
	1.6.2.1 pH of the colon	
	1.6.2.2 Transit time to colon	
	1.6.2.3 Colonic micro flora and their enzymes	

1.6.3	Strateg	gies for targeting drugs to the colon	. 26
	1.6.3.1	Covalent linkage of drug with a carrier	
		1.6.3.1.1. Prodrug approaches	
		1.6.3.1.1.1 Azo bond conjugate	
		1.6.3.1.1.2 Glycoside conjugation	
		1.6.3.1.1.3 Glucoronide conjugates	
		1.6.3.1.1.4 Amino acid conjugation	
	1.6.3.2	Approaches to intact molecule delivery to the colon	
		1.6.3.2.1 Bioadhesive systems	
		1.6.3.2.2 Pressure controlled systems	
		1.6.3.2.3 Time dependent delivery	
		1.6.3.2.4 pH dependent approach	
		1.6.3.2.5 Microbially triggered drug delivery to the colon	
1.7 Po	lysacch	aride-based colon targeted drug delivery systems	. 32
1.7.1	Chitin		. 35
1.7.2	Chitos	an	. 36
	1.7.2.1	Production of chitosan	
	1.7.2.2	Structure of chitosan	
	1.7.2.3	Properties of Chitosan	
	1.7.2.4	Physiochemical characteristic of chitosan	
		1.7.2.4.1 Degree of deacetylation	
		1.7.2.4.2 Molecular weight	
	1.7.2.5	Biological properties of chitosan	
	1.7.2.6	Derivatives of chitosan	
		1.7.2.6.1 Physical modification	
		1.7.2.6.2 Chemical modification	
1.8 Cl	nitosan	based colon drug delivery systems	. 47
1.8.1	Micros	spheres	. 47
1.8.2	Nanop	articles	. 48
1.8.3	Beads		. 49
1.8.4	Tablet	s	. 49
1.9 Tł	esis ob	jective	. 50
		in knowledge that this research aims to fill	
1.11 T	hesis st	ructure	.51
1.12 R	Referenc	es	. 53

Cnap	oter Iwo: Preparation and characterization of	crossiinkea
Chite	osan based films with excellent physiochemical proper	ties
2.0 In	troduction	77
2.1 M	aterials and Methods	78
2.1.1	Materials	78
2.1.2	Preparation and crosslinking of the polymeric films	79
2.2. C	haracterization of the crosslinked films	79
2.2.1	Fourier transform infrared (FTIR) spectroscopy	79
2.2.2	Nuclear magnetic resonance (NMR) spectroscopy	79
2.2.3	Determination of the degree of substitution	80
2.2.4	Swelling studies	80
2.2.5	Film thickness	81
2.2.6	CHN elemental analysis	81
2.2.7	Mechanical test	82
2.2.8	Thermal gravimetric analysis (TGA) and differential thermal	
	gravimetric analysis (DTG)	82
2.2.9	In vitro biodegradation study	82
2.2.10	Contact angle analysis	82
2.3 Re	esults and discussion	77
2.3.1	The degree of substitution	82
2.3.2	Fourier Transform Infrared (FTIR) Spectroscopy	85
2.3.3	Mechanical testing	86
2.3.4	Swelling studies	87
2.3.5	Solid state 13C NMR analysis	89
2.3.6	In vitro biodegradability	90
2.3.7	Thermal gravimetric analysis (TGA) and differential thermal	
	gravimetric analysis (DTG)	91
2.3.8	Water Contact Angle Measurements	93
2.4 Co	onclusion	94
2.5 Re	eferences	95

Chap	oter Three: In vitro evaluation of spray-dried chite	osan
micr	ospheres cross-linked with pyromellitic dianhydride for	oral
color	n-specific delivery of protein drugs	
3.0 In	troduction	99
3.1 M	aterials and Methods	. 100
3.1.1	Materials	. 100
3.1.2	Preparation of the spray-dried chitosan microspheres	. 100
3.1.3	Crosslinking of the spray-dried chitosan microspheres	. 101
3.2. C	Characterization	. 101
3.3. D	etermination of the swelling behavior of the microparticles	. 102
3.4. B	SA loading and release experiments	. 102
3.4.1	Protein encapsulation efficiency and loading capacity	. 102
3.4.2	In vitro drug-release study	. 103
3.5. H	IPLC protein analysis	. 103
3.6. S	tatistics	. 104
3.7 R	esults and discussion	. 104
3.7.1	Preparation of the spray-dried chitosan microspheres	. 104
3.7.2	Swelling studies	. 108
3.7.3	Microsphere encapsulation efficiency (EE) and loading capacity (LC) study	109
3.7.4	In vitro release study	. 111
3.8 C	onclusion	. 114
3.9 R	eferences	. 115
Chap	oter Four: Synthesis and characterization of a novel generatio	n of
ampl	hoteric pH sensitive hydrogels	
4.0 In	troduction	. 119
4.1 M	aterials and Methods	. 120
4.1.1	Materials	. 120
4.1.2	Preparation of chitosan films	. 121
4.1.3	Preparation of amic acid derivatives	. 121
4.1.4	Preparation of crosslinked chitosan films	. 121
4.2. E	nzyme inhibitory effect	. 121
4.2.1	Trypsin inhibition study	. 122

4.2.2	Chymotrypsin inhibition study	122
4.3 Re	esults and discussion	122
4.3.1	Preparation of chitosan amic acid films	122
4.3.2	Fourier transform infrared (FTIR) spectroscopy	125
4.3.3	Solid state 13C NMR analysis	126
4.3.4	Mechanical test	127
4.3.5	Thermal gravimetric analysis (TGA) and differential thermal	
	gravimetric analysis (DTG)	128
4.3.6	Swelling studies	129
4.3.7	Water contact angle measurements	130
4.3.8	In vitro biodegradability	131
4.3.9	Enzyme inhibitory effect	132
4.4 Co	onclusion	133
4.5 Re	eferences	134
Chap	oter Five: Development of a pH sensitive carrier system b	ased on a
nove	l water soluble chitosan and alginate for colon targe	
novel deliv	l water soluble chitosan and alginate for colon targe	eted drug
novel delive 5.0 In	l water soluble chitosan and alginate for colon targe ery troduction	eted drug
novel delive 5.0 In 5.1 Ex	l water soluble chitosan and alginate for colon targe ery troduction	eted drug137138
novel delive 5.0 In 5.1 Ex	l water soluble chitosan and alginate for colon targe ery troduction	137 138
novel delive 5.0 In 5.1 Ex 5.1.1	l water soluble chitosan and alginate for colon targe ery troduction	137 138 138
novel delive 5.0 In 5.1 Ex 5.1.1 5.1.2 5.1.3	l water soluble chitosan and alginate for colon targe ery troduction	137138138138
novel delive 5.0 In 5.1 Ex 5.1.1 5.1.2 5.1.3	water soluble chitosan and alginate for colon targetery troduction xperimental Materials Preparation of Cts-TMAC amic acid (CTAA) Preparation of CTAA amic acid (CTAA) film	
novel delive 5.0 In 5.1 Ex 5.1.1 5.1.2 5.1.3 5.2. C	water soluble chitosan and alginate for colon targetery troduction xperimental Materials Preparation of Cts-TMAC amic acid (CTAA) Preparation of CTAA amic acid (CTAA) film Characterization of the CTAA amic acid (CTAA) film	137138138138139
novel delive 5.0 In 5.1 Ex 5.1.1 5.1.2 5.1.3 5.2. C 5.2.1	water soluble chitosan and alginate for colon targetery troduction xperimental Materials Preparation of Cts-TMAC amic acid (CTAA) Preparation of CTAA amic acid (CTAA) film Characterization of the CTAA amic acid (CTAA) film NMR spectroscopy	
novel delive 5.0 In 5.1 Ex 5.1.1 5.1.2 5.1.3 5.2. C 5.2.1 5.2.2	I water soluble chitosan and alginate for colon targetery troduction xperimental Materials Preparation of Cts-TMAC amic acid (CTAA) Preparation of CTAA amic acid (CTAA) film tharacterization of the CTAA amic acid (CTAA) film NMR spectroscopy X-ray diffraction (XRD)	
novel delive 5.0 In 5.1 Ex 5.1.1 5.1.2 5.1.3 5.2. C 5.2.1 5.2.2 5.2.3 5.2.4	I water soluble chitosan and alginate for colon targetery troduction	
novel delive 5.0 In 5.1 Ex 5.1.1 5.1.2 5.1.3 5.2. C 5.2.1 5.2.2 5.2.3 5.2.4 5.3. Pr	I water soluble chitosan and alginate for colon targetery troduction Aperimental Materials Preparation of Cts-TMAC amic acid (CTAA) Preparation of CTAA amic acid (CTAA) film Characterization of the CTAA amic acid (CTAA) film NMR spectroscopy X-ray diffraction (XRD) Scanning electron microscope Solubility test	
novel delive 5.0 In 5.1 Ex 5.1.1 5.1.2 5.1.3 5.2. C 5.2.1 5.2.2 5.2.3 5.2.4 5.3. Pr 5.4. Second state of the second state of t	I water soluble chitosan and alginate for colon targetery troduction	
novel delive 5.0 In 5.1 Ex 5.1.1 5.1.2 5.1.3 5.2. C 5.2.1 5.2.2 5.2.3 5.2.4 5.3. Pr 5.4. So 5.5. R	I water soluble chitosan and alginate for colon targetery troduction	

5.8. C	ytotoxicity test	142
5.9 Re	sults and discussion	143
5.9.1	Preparation of CTAA	143
5.9.2	Characterization of the chitosan derivative	143
5.9.3	Thermal gravimetric analysis (TGA) and differential thermal	
	gravimetric analysis (DTG)	148
5.9.4	Enzyme inhibitory effect of CTAA	149
5.9.5	Solubility test	150
5.9.6	Swelling studies of CTAA/alginate films	150
5.9.7	In vitro drug release study	152
	5.9.7.1 Effect of pH of media on release profile of 5-FU from CTAA/alginate films	
	5.9.7.2 Effect of enzyme on release profile of 5-FU from CTAA/alginate films	
5.9.8	Cytotoxicity studies	156
5.10 (Conclusion	156
5.11 F	References	157
	specific delivery system using novel amphoteric chitosan	based
6.0 In	ix tablet.	
6.1 M		160
	ix tablet.	
6.1.1	ix tablet.	161
6.1.1 6.1.2	ix tablet. troductionaterials and Methods	161 161
6.1.2	ix tablet. troduction aterials and Methods Materials	161 161 161
6.1.2 6.2. C	ix tablet. troduction aterials and Methods Materials Preparation of Amic acid derivatives	161 161 161 162
6.1.2 6.2. C	ix tablet. troduction aterials and Methods Materials Preparation of Amic acid derivatives haracterization of the hydrogel	161 161 161 162 162
6.1.2 6.2. C 6.3. P	ix tablet. troduction aterials and Methods Materials Preparation of Amic acid derivatives haracterization of the hydrogel reparation and evaluation of tablets	161 161 162 162 162
6.1.2 6.2. C 6.3. P 6.3.1	ix tablet. troduction aterials and Methods Materials Preparation of Amic acid derivatives haracterization of the hydrogel reparation and evaluation of tablets Preparation of compression-coated tablets	161 161 162 162 162
6.1.2 6.2. C 6.3. P 6.3.1 6.3.2	ix tablet. troduction. aterials and Methods. Materials. Preparation of Amic acid derivatives. haracterization of the hydrogel. reparation and evaluation of tablets. Preparation of compression-coated tablets. Tablet crushing strength	161 161 162 162 162 163
6.1.2 6.2. C 6.3. P 6.3.1 6.3.2 6.3.3	ix tablet. troduction	161 161 162 162 162 163
6.1.2 6.2. C 6.3. P 6.3.1 6.3.2 6.3.3 6.3.4 6.3.5	ix tablet. troduction aterials and Methods Materials Preparation of Amic acid derivatives haracterization of the hydrogel reparation and evaluation of tablets Preparation of compression-coated tablets Tablet crushing strength Swelling behaviour of tablets. Tablet erosion study	161 161 162 162 162 163 163
6.1.2 6.2. C 6.3. P 6.3.1 6.3.2 6.3.3 6.3.4 6.3.5 6.4. In	ix tablet. troduction aterials and Methods Materials Preparation of Amic acid derivatives haracterization of the hydrogel reparation and evaluation of tablets Preparation of compression-coated tablets Tablet crushing strength Swelling behaviour of tablets. Tablet erosion study Enzymatic degradation study of tablets	161 161 162 162 162 163 163 164

6.5.2	BSA determination	165
6.6. R	esults and discussion	165
6.6.1	Preparation of chitosan amic acid derivatives	166
6.6.2	Characterization	168
6.6.3	Enzyme inhibitory effect of CBAA	176
6.6.4	Swelling and erosion behaviour tablets	176
	6.6.4.1 Cts-CBAA (75:25%)	
	6.6.4.2 Cts-COAA (75:25%)	
6.6.5	Enzymatic degradation of tablet	185
6.6.6	In vitro drug release study	185
	6.6.6.1 Controlled colon specific delivery system of 5-ASA using Cts:CBAA table	et
	6.6.6.1.1 Effect of coating polymer ratio	
	6.6.6.1.2 Effect of pH and enzyme	
	6.6.6.2 Colon specific delivery of protein therapeutics using Cts:COAA tablet	
	6.6.6.2.1 Effect of coating polymer ratio	
.	6.6.6.2.2 Effect of pH and enzyme	40.4
6.6.7	Cytotoxicity studies	
6.7. C	onclusion	195
6.8 Re	eferences	195
Chap	oter Seven: Development and evaluation of a novel colon	targeting
drug	delivery system for the treatment of Tritrichomona	as foetus
intest	tinal infection in cats	
7.0 In	troduction	199
7.1 M	aterials and Methods	200
7.1.1	Materials	200
7.1.2	Preparation of Cts-PMDA amic acid (CPAA)	200
7.1.3	Swelling behaviour of the tablets	200
7.1.4	Tablets erosion study	202
7.1.5	In vitro drug release study	202
7.1.6	HPLC analysis	202
7.2. R	esults and discussion	203
7.2.1	Preparation of CPAA	203
7.2.2	Characterization	
	7.2.2.1 Fourier transform infrared (FTIR) spectroscopy	

	7.2.2.2	Nuclear magnetic resonance (NMR) spectroscopy	
	7.2.2.3	Scanning electron microscopy (SEM)	
	7.2.2.4	Powder X-ray diffraction study	
	7.2.2.5	Thermogravimetric analysis (TGA)	
	7.2.2.6	Enzyme inhibitory effect of CPAA	
	7.2.2.7	Swelling and erosion behaviour of Cts:CPAA tablets	
	7.2.2.8	Enzymatic degradation study	
	7.2.2.9	Swelling and erosion behaviour of Cts:CPAA tablets	
7.2.3	In vitro	o drug release study	203
	7.2.3.1	Effect of coating polymer ratio	
	7.2.3.2	Effect of pH and enzyme	
7.2.4	Cytoto	oxicity studies	203
7.3. C	onclusio	on	216
7.4 Re	eference	es	216
Chap	ter Ei	ght: Summary and possible future directions	
7.0 Su	mmary	of results	219
7.1 Re	ecomme	endations for future studies	223
Appei	ndices		225

LIST OF TABLES

Number	Page
Table 1-1 Example of various environmentally stimulated hydrogel systems used for drug deliver	y 19
Table 1-2 Anatomical and physiological features of the gastrointestinal tract.	23
Table 1-3 Commonly used pH responsive coating polymers in oral drug delivery.	31
Table 1-4 Polysaccharide based colon targeted delivery systems	33
Table 1-5 Relationships between Cts biological properties and its degree of deacetylation (DD)	
and molecular weight (Mw)	43
Table 1-6 Examples of various crosslinked chitosan	45-46
Table 2-1 Puncture strength (PS) and elongation at break (% E) of the chitosan and crosslinked	
chitosan films.	87
Table 4-1 The elemental analyses results and the substitution degree of chitosan and crosslinked	
chitosan	124
Table 4-2 Puncture strength (PS) of the chitosan and crosslinked chitosan films.	127
Table 5-1 Solubility test results for Cts and CTAA.	150
Table 7-1 Effect of β -glucosidase enzyme on the Cts:CPAA (75:25%) polymer degradation in	
pH 5.5	212
Table 7-2 Effect of different coating weight ratio (%) on in-vitro release of ronidazole	213

LIST OF FIGURES

Number	Page
Figure 1-1 Example of hydrogels for biomedical and pharmaceutical applications a) Hydrogel	
loaded with a model drug and b) poly (2-hydroxyethylmethacrylate) soft contact lens	1
Figure 1-2 Molecular structures of typical polymers used for natural and synthetic hydrogel	
preparation	3
Figure 1-3 Formula of some typical natural and synthetic biodegradable polymers	6
Figure 1-4 Stimuli responsive swelling of hydrogels	8
Figure 1-5 Structures of some pH-sensitive polymers.	9
Figure 1-6 Synthesis of the complex between a phenylboronic acid complex and glucose in	
aqueous solution.	9
Figure 1-7 Chemical formulas of polymers that form or are part of thermoresponsive hydrogels	11
Figure 1-8 Structure of some polymers commonly used in tissue engineering	14
Figure 1-9 Structure of some polymers commonly used in wound healing	15
Figure 1-10 Drug level in blood with a) traditional drug administration and b) controlled	
drug delivery	16
Figure 1-11 Anatomy of gastrointestinal tract.	21
Figure 1-12 Pathway of colonic reduction of sulfasalazine.	27
Figure 1-13 Dexamethasone-2-β-D-glucoside prodrug.	28
Figure 1-14 Dexamethasone β-D-glucuronide prodrug.	28
Figure 1-15 Structure of 5-aminosalicyl-glycine prodrug.	29
Figure 1-16 Chemical manufacturing processes for chitin	36
Figure 1-17 Chemical manufacturing processes for chitosan	37
Figure 1-18 Structure of chitin, chitosan and cellulose	38
Figure 1-19 Deactylation mechanism of chitin	41
Figure 1-20 Chitosan active sites	44
Figure 2-1 Representation of crosslinked chitosan based films.	84
Figure 2-2 FTIR spectra of chitosan, and dianhydride-crosslinked chitosan.	85
Figure 2-3 Swelling behaviour of chitosan films at different pH.	88
Figure 2-4 13C DP-MAS spectra of chitosan and dianhydride-crosslinked chitosan	89
Figure 2-5 Results of the degradability of the chitosan and crosslinked chitosan in a LYZ solution.	91
Figure 2-6 a) TGA and b) DTG thermograms of chitosan and crosslinked chitosan	92
Figure 2-7 Appearances of water drops on Cts and crosslinked Cts film surfaces	94
Figure 3-1 Schematic representation of a Cts-PMDA microsphere.	104
Figure 3-2 FTIR spectra of Cts and Cts-PMDA microspheres.	105

Number	Page
Figure 3-3 ¹³ C DP-MAS spectra of chitosan-PMDA.	106
Figure 3-4 SEM images and size distribution of chitosan and Cts-PMDA microparticles	107
Figure 3-5 Swelling behaviour of Cts-PMDA microspheres in a) SGF and SIF	
b) simulated gastrointestinal tract (2 h in pH 1.2, 6 h in pH 7.4, 12 h in pH 7)	108
Figure 3-6 The influence of BSA initial concentration on a) encapsulation efficiency and	
b) loading capacity of Cts-PMDA microspheres	110
Figure 3-7 Effect of pH on cumulative release of BSA from Cts and Cts-PMDA microspheres	112
Figure 3-8 The BSA release profile in simulated gastrointestinal fluid for 2 h, followed	
by 6 h in simulated intestinal fluid and then 12 h in simulated colonic fluid	110
Figure 4-1 Schematic representation of the different possible reactions between the amino	
group of chitosan and dianhydride derivatives	123
Figure 4-2 FTIR spectra of chitosan, and amic acid derivatives of chitosan	125
Figure 4-3 ¹³ C DP-MAS spectra of chitosan and amic crosslinked chitosan.	126
Figure 4-4 TGA and b) DTG thermograms of chitosan and crosslinked chitosan	128
Figure 4-5 Swelling behaviour of chitosan films at different pH.	130
Figure 4-6 Visualisation of water contact angles on chitosan and crosslinked chitosan film	
surfaces	131
Figure 4-7 Results of the degradability of the chitosan and amic acid derivatives of chitosan	
in a lysozyme.	132
Figure 5-1 Reaction schemes and FTIR spectra of crosslinked chitosan with TMAC a) at 130 °C	
b) at room temperature.	144
Figure 5-2 The (a) 1H NMR (500 MHz) and (b) 13C DP-MAS spectrum of CTAA	145
Figure 5-3 1H-13C HSQC of CTAA (700 MHz)	146
Figure 5-4 2D-XRD of (a) Cts and (b) CTAA.	147
Figure 5-5 SEM images of a) Cts (at 10 μm and 50 μm) and b) CTAA (at10 μm and 50 μm).	148
Figure 5-6 TGA and DTG thermograms of Cts and CTAA	149
Figure 5-7 Swelling behaviour of CTAA/alginate films at SIF and SGF.	151
Figure 5-8 Effect of pH on the release profile of 5-FU from CTAA/alginate films	152
Figure 5-9 Effect of enzyme on the release profile of 5-FU from CTAA/alginate films	154
Figure 5-10 Percent viability of cells incubated with tested compounds as compared to control cells.	155
Figure 6-1 Generalized reaction mechanism of chitosan with dianhydride derivatives	166
Figure 6-2 Schematic representation CBAA and COAA	167
Figure 6-3 FTIR spectra of a) Cts, b) COAA and c) CBAA	169
Figure 6-4 13C DP-MAS spectra of (a) Cts, (b) CBAA and c) COAA	170
Figure 6-5 SEM image of (a) Cts, (b) CBAA and c) COAA.	172
Figure 6-6 2D-XRD of (a) Cts and (b) CBAA.	173
Figure 6-7 2D-XRD of (a) Cts and (b) COAA	
Figure 6-8 TGA and DTG thermograms of Cts and CBAA	175
Figure 6-9 TGA and DTG thermograms of Cts and COAA.	175

Number
Figure 6-10 Photographs of radial and axial swelling behaviour of Cts-CBAA (75:25 %)
tablets in various media
Figure 6-11 Swelling behaviour of Cts-CBAA tablets in various media
Figure 6-12 Photograph of the radial and axial swelling behaviour of tablets in a) healthy
b) an IBD simulated gastrointestinal tract condition
Figure 6-13 Swelling behaviour of the Cts-CBAA tablets in a) healthy b) an IBD
simulated gastrointestinal tract condition
Figure 6-14 Percentages remaining of Cts-CBAA matrix tablets
Figure 6-15 Swelling behaviour of Cts-COAA tablets in various media
Figure 6-16 Swelling behaviour of Cts-COAA tablets in a simulated gastrointestinal tract
Figure 6-17 Photograph of radial and axial swelling behaviour of tablets in a) pH 1.2
b) 7.4 and c) simulated gastrointestinal tract pH protocol
Figure 6-18 Percentages remaining of Cts-CBAA matrix tablets
Figure 6-19 Effect of coating ratio on cumulative release of 5-ASA from matrix tablets
Figure 6-20 Effect of enzyme on cumulative release of 5-ASA from the selected tablet
(Cts:CBAA 75:25%)
Figure 6-21 Effect of coating ratio on cumulative release of BSA from matrix tablets
Figure 6-22 Effect of enzyme on cumulative release of BSA from selected tablet
(Cts:COAA 75:25%)193
Figure 6-23 Percent viability of cells incubated with tested compounds as compared to control cells194
Figure 7-1 Reaction illustration and FTIR spectra of crosslinked chitosan with PMDA a) at 130 °C
b) at room temperature
Figure 7-2 13C DP-MAS spectra of (a) Cts and (b) CPAA
Figure 7-3 SEM image of (a) Cts and (b) CPAA.
Figure 7-4 2D-XRD of (a) Cts, (b) CPAA
Figure 7-5 TGA and DTG thermograms of Cts and CPAA
Figure 7-6 Photographs of the radial and axial swelling behaviour of the Cts:CPAA (75:25%)
tablet in different pH media208
Figure 7-7 Swelling behaviour of Cts:CPAA (75:25%) tablets in a) pH 1.5, b) pH 6.5 and
c) pH 5.5209
Figure 7-8 Photographs of radial and axial swelling behaviour of tablets in a simulated GIT of a cat 210
Figure 7-9 Swelling behaviour of Cts:CPAA (75:25 %) tablets in a simulated gastrointestinal tract210
Figure 7-10 Percentages remaining of Cts:CPAA (75:25 %) matrix tablets a) 24 h in pH 1.2, 6.5,5.5
b) in a simulated gastrointestinal tract (30 min in pH 1.5 followed by 2 h in pH 6.5 then
24 h in pH 5.5)211
Figure 7-11 Effect of enzyme on cumulative release of ronidazole from the Cts:CPAA (75:25 %)214
Figure 7-12 Percent viability of cells incubated with tested compounds as compared to control cells215

LIST OF ABBREVIATIONS

AAm Acrylamide

5-ASA5-aminosalicylic acidBSABovine serum albumin

BAPNA N-α-benzoyl-l-arginine p-nitroanilide

BTEE N-benzoyl-L-tyrosine ethyl ester solution

BTDA Benzophenone-3,3',4,4'-tetracarboxylic dianhydride

CF Carboxyfluorescein

Cts Chitosan

CST Critical solution temperature

CFU Colony forming unit

CFAA Amic acid derivative chitosan crosslinked with BTDA
CFAA Amic acid derivative chitosan crosslinked with FDA
CNAA Amic acid derivative chitosan crosslinked with NTDA
COAA Amic acid derivative chitosan crosslinked with ODPA
CPAA Amic acid derivative chitosan crosslinked with PMDA

CTAA Amic acid derivative chitosan crosslinked with TMAC

DD Degree of deacetylation

DMF Dimethylformamide

DD Degree of deacetylation

DTG Differential thermal gravimetric analysis

Da Dalton

EC Ethylcellulose
Ea Electron affinity

EVAC Ethylenevinyl acetate copolymer

EE Encapsulation efficiency

ppm Parts per million

EVAC Ethylene dimethacrylate

FDA 4,4'-(Hexafluoroisopropylidene) diphthalic anhydride

5-FU 5-Fluorouracil

FTIR Fourier transform infrared spectroscopy

Gastrointestinal system

GIT Gastrointestinal tract

GA Glutaraldehyde

g Gram

HCl Hydrochloric acid

HPLC High-performance liquid chromatography

HEMA HydroxyethylmethacrylateHPMA HydroxypropylmethacrylateIBD Inflammatory bowel disease

LCST Lower critical solution temperature

LC Loading capacity

LD Lethal doseLYZ Lysozyme

MPEG Methoxyl poly(ethylene glycol)

Mw Molecular weight

MAS Magic angle spinning

MHzMegahertzMPaMegapascal

NTDA 1, 4, 5, 8-Naphthalenetetracarboxylic dianhydride

NaCS Sodium cellulose sulfate

NMR Nuclear magnetic resonance spectroscopy

ODPA 4,4'-Oxydiphthalic dianhydride

PBS Phosphate buffered saline

PAA Poly (acrylic acid)

PMAAc Poly (methacrylic acid)

PDMAEMA Poly (N,N'-dimethylaminoethyl methacrylate)

PL Poly (lysine)

PHEMA Poly (2-Hydroxyethyl methacrylate)

PEG Poly (ethylene glycol)

PNIPAm Poly (*N*-isopropylacrylamide)

PEO Polyethylene oxide

PS Puncture strength

PEGMA Polyethyleneneglycol methacrylate

PNPAm Poly (N-n-propylacrylamide)

PDEAM Poly (N,N-diethylacrylamide)

PVA Polyvinylalcohol

pH Measure of acidity and basicity in solution

PPS Sodium polyphosphate

PMDA Pyromellitic dianhydride

RDZ Ronidazole

RPM Revolutions per minute
SCF Simulated colonic fluid

SEM Scanning electron microscopy

Swelling percentage

SGF Simulated gastric fluids

SIF Simulate gastric fluids

T. foetus Tritrichomonas foetus

TGA Thermal gravimetric analysis
TMAC Trimellitic anhydride chloride

UC Ulcerative colitis

UCST Upper critical solution temperature

XRD X-ray diffraction

LIST OF PUBLICATIONS

- 1. **Kavianinia**, **I.**; Plieger, P. G.; Kandile, N. G.; Harding, D. R., In Vitro Evaluation of Spray-Dried Chitosan Microspheres Crosslinked with Pyromellitic Dianhydride for Oral Colon-Specific Delivery of Protein Drugs. *Article first published online:* 13 Feb **2013**.
- 2. **Kavianinia, I.;** Plieger, P. G.; Kandile, N. G.; Harding, D. R., Preparation and characterization of crosslinked chitosan based films with excellent physiochemical propertie. *International Journal of Biological Macromolecules*. Manuscript under revision.
- 3. **Kavianinia, I.;** Plieger, P. G.; Kandile, N. G.; Harding, D. R., Fixed-bed column studies on a modified chitosan hydrogel for detoxification of aqueous solutions from copper (II). *Carbohydrate Polymers.* **2012**, 90 (2), 875–886.
- 4. **Kavianinia, I.**; Plieger, P. G.; Kandile, N. G.; Harding, D. R., New hydrogels based on symmetrical aromatic anhydrides: Synthesis, characterization and metal ion adsorption evaluation, *Carbohydrate Polymers*. **2012**, 87 (1), 881–893.

Papers to be submitted

- 1. Development of a pH sensitive carrier system based on a novel water soluble chitosan and alginate for colon targeted drug delivery. *Under preparation*
- 2. Development and evaluation of a novel colon targeting drug delivery system for the treatment of Tritrichomonas foetus intestinal infection in cats. *Under preparation*
- 3. Synthesis and characterization of a novel generation of amphoteric pH sensitive hydrogels. *Under preparation*

- 4. Formulation and evaluation of a novel pH and enzyme controlled colon-specific delivery system of 5-ASA using amphoteric chitosan based matrix tablet. *Under preparation*.
- 5. Preparation and characterization of an amphoteric chitosan based matrix table chitosan based matrix tablet for oral colon-specific drug delivery of protein therapeutics. *Under preparation*.