

Copyright is owned by the Author of the thesis. Permission is given for a copy to be downloaded by an individual for the purpose of research and private study only. The thesis may not be reproduced elsewhere without the permission of the Author.

Biosensors for fertility and pregnancy in cattle

A thesis presented
in partial fulfilment of the requirements
for the degree of
Doctor of Philosophy in Chemistry
at Massey University, Palmerston North,
New Zealand

Yu-ting Hsu

2013

Acknowledgements

First and foremost I would like to thank my supervisors Associate Professor Eric Ainscough and Associate Professor Ashton Partridge who were friendly and supportive throughout my study. Additionally, I would like to thank Professor Bernd Rehm and the Polybatics group for providing the polyhydroxyalkanoate granules and valuable information and advice towards the research and development of a surface plasmon resonance sensor.

I would like to acknowledge the MacDiarmid Institute for providing financial support.

I wish to thank Dr Wayne Campbell and Dr Krishanthi Jayasundera for the synthesised progesterone ligands. I would like to thank Associate Professor Len Blackwell and Dr Jenness Guthrie for advice and information on the lateral flow immunoassay development.

Finally, I wish to thank my family and Ross for supporting my efforts to pursue a PhD.

Abstract

This project is focused on progesterone sensing, using both surface plasmon resonance (SPR) and lateral flow immunoassay (LFIA) methods with a new progesterone (P4) sensing material to develop cost effective assays for progesterone sensing in bovine serum and milk samples.

P4-PEG-OVA was synthesised, characterised and used for P4 detection. The P4-PEG-OVA sensor surface showed an improvement in surface response compared with two shorter ligand 4TP-P4-OVA and 4TPH-P4-OVA in SPR studies.

An analysis method has been developed and modified for bovine serum and milk analyses. The results indicated the P4-PEG-OVA ligand allowed sensitive P4 detection in SPR sensing and allowed bovine P4 cycle profiling. The SPR analysed data was compatible with the ECLIA and ELISA independent analyses and the P4 cycle of each of the three bovine milk samples showed a very similar trend and the extraction level was also consistent.

The P4-PEG-OVA ligand was used to develop a LFIA sensor strip, and the inhibition assay for bovine serum and milk analyses established. The results indicated that, after appropriate sample pre-treatment, the bovine estrous cycle profile could be detected. The LFIA method can be a potentially quick, easy and cost effective semi-quantitative P4 analysis for serum and milk samples.

A new material, polyhydroxyalkanoate (PHA) granules has been investigated for the possibility of developing a new surface biosensor. From the surface studies, the results indicated that the 3GNZZPhaC beads have the potential to become an alternative binding material for SPR sensing due to its unique gold binding property. A flow cell was designed, constructed, and tested on 3GNZZPhaC beads prior the preliminary SPR investigations.

The ZZPhaC beads also showed the gold binding property and ZZPhaC beads were used for SPR studies. The results suggested a possible application for them as a new SPR binding material for antibody detection.

Contents

Chapter 1: Introduction	1
1.1 Dairy farming in New Zealand.....	1
1.2 Mammalian/bovine progesterone (P4) cycle	1
1.3 Biosensors	3
1.4 Surface Plasmon Resonance (SPR)	5
1.4.1 SPR-based applications	7
1.4.2 The Biacore X100 system	8
1.4.2.1 The microfluidic system	9
1.4.2.2 Biacore sensing chips	10
1.4.2.3 Surface immobilisation	11
1.5 The Lateral Flow Immunoassay (LFIA)	13
1.5.1 Structure of a lateral flow immunoassay strip	16
1.5.1.1 Backing card	17
1.5.1.2 Membrane materials	17
1.5.1.3 Sample pad	18
1.5.1.4 Absorbing pad	18
1.5.1.5 Conjugate pad	19
1.5.1.6 Labelling material	19
1.5.2 Capillary flow rate	19
1.6 Immunoassay	20
1.6.1 Immunoassay of steroids	20
1.6.2 Progesterone sensing	21
1.7 Progesterone sensing and the present study.....	23
References	24
Chapter 2: SPR based biosensors	27
2.1 Introduction	27
2.1.1 The evanescent wave	29

2.2	Development of SPR assays	30
2.3	Progesterone (P4) derivatives	33
2.3.1	Synthesis of P4-PEG-OVA	34
2.3.1.1	A brief overview of the synthesis of P4-PEG-COOH	34
2.3.1.2	Conjugation of P4S(CH ₂) ₂ PEG-COOH to ovalbumin (OVA) to give P4-PEG-OVA	35
2.3.1.3	Immobilisation of ligands onto the CM5 chip	36
2.3.2	Binding performance with monoclonal rat Anti-progesterone (Sigma P1922)	39
2.3.2.1	4TP-P4-OVA	40
2.3.2.2	4TPH-P4-OVA	41
2.3.2.3	P4-PEG-OVA	42
2.3.2.4	Comparison of binding performance between P4-linkers	43
2.3.3	Binding performance with monoclonal mouse Anti-P4 (Serotech)	44
2.3.3.1	Binding performance stability on the 4TP-P4-OVA sensor	44
2.3.3.2	Comments about mouse and rat Anti-P4 binding to a 4TP-P4-OVA surface	46
2.3.3.3	Comments about mouse and rat Anti-P4 binding to a 4TPH-P4-OVA surface	47
2.3.3.4	Comments about mouse and rat Anti-P4 binding to a P4-PEG-OVA surface	48
2.3.3.5	Comparison of binding performance between P4 linkers	49
2.3.4	Inhibition assay development with mouse Anti-P4 (Serotech)	50
2.3.4.1	Assay development with 4TP-P4-OVA	51
2.3.4.2	Assay development with 4TPH-P4-OVA	52
2.3.4.3	Assay development with P4-PEG-OVA	53
2.4	Determination of P4 in bovine serum samples	56
2.4.1	Non-specific binding of P4 to corticosteroid binding globulin	56
2.4.2	Free P4 analysis	58

2.4.3	Preparation of serum samples: solvent extraction	59
2.4.4	Determination of P4 in bovine serum samples	61
2.5	Determination of P4 in bovine milk samples	64
2.5.1	Bovine milk 1	65
2.5.2	Bovine milk 2	67
2.5.3	Bovine milk 3	68
2.6	Conclusions	70
	References	71
 Chapter 3: Lateral flow immunoassay for progesterone sensing		75
3.1	Introduction	75
3.2	LFIA system	77
3.2.1	Processing methodologies	79
3.3	Development of the P4-PEG-OVA sensor strip	81
3.3.1	Antibody titration	81
3.3.2	Conjugation of Anti-P4 to gold nanoparticles	83
3.3.3	Development of the P4-PEG-OVA strip	83
3.3.4	The P4 binding curve	84
3.3.5	The P4 standard curves	86
3.3.5.1	Concentration scouting	86
3.3.5.2	The P4 standard curve established with the P4-PEG-OVA test strips	87
3.4	Determination of P4 in bovine serum using LFIA	88
3.4.1	Preliminary free P4 analysis	88
3.4.2	Bovine serum samples analysis	91
3.4.2.1	P4 standard curves with low [P4] serum samples	91
3.4.2.2	Serum analysis for cow 1	93
3.5	Bovine milk progesterone	99
3.5.1	Independent analysis	101
3.5.2	P4 standard curves for milk analysis	103

3.5.3	Milk sample preparation	104
3.5.4	Comparison of the P4 LFIA analysis to the independent ELISA analysis of milk samples after protein and fat removal	107
3.6	Conclusions	115
	References	116

Chapter 4: Toward an alternative sensing material for SPR sensing:

	Polyhydroxyalkanoate granules	119
4.1	Introduction to polyhydroxyalkanoates	119
4.2	Polyhydroxyalkanoate granules	120
4.2.1	Formation of polyester granules	122
4.2.2	Multifunctional inorganic binding beads	123
4.3	Applications of PHA granules	126
4.4	Surface applications	127
4.4.1	Dip coating	127
4.4.2	Spin coating	128
4.5	Flow cell system	129
4.6	Microscopy studies	134
4.6.1	Scanning electron microscopy studies	134
4.6.1.1	3GNZZPhaC immobilisation on the Au surface	134
4.6.1.2	SEM of ZZPhaC immobilisation on the Au surface	136
4.6.2	Preliminary atomic force microscopy (AFM) studies	138
4.7	The selection of ZZPhaC beads as an alternative SPR sensing material	141
4.7.1	Preliminary binding test	142
4.7.1.1	Outside immobilisation of ZZPhaC beads	142
4.7.1.2	Binding test using rat Anti-P4 on a ZZPhaC surface	143
4.7.1.3	ZZPhaC immobilisation (in Biacore X100)	150
4.7.1.4	Binding performance testing with rat Anti-P4 on the Au-ZZPhaC surface	153
4.8	Preliminary LFIA testing	158

4.9 Conclusions160

References161

Chapter 5: Conclusions and future work165

Appendix.....168

List of figures, tables and reaction schemes

Chapter 1

Figure 1.1	A bovine estrous cycle can be monitored by measuring the P4 concentration in both serum and milk samples. The duration of a typical bovine estrous cycle is approximately 21 days.	2
Figure 1.2	A typical biosensor comprises a capture agent unit which consists of a biological element such as an enzyme, immunoagents, or microorganisms.	3
Figure 1.3	A modern SPR sensing system based on the Kretschmann configuration.	6
Figure 1.4	Sensorgram showing the steps of an analysis cycle. The progesterone standard was injected at 30 $\mu\text{L}/\text{min}$ for 120 s and this was followed by a regeneration step of injection of NaOH solution (10 mM) for 30 s which regenerates the surface. The analyte flows through flow cell one (FC1) and flow cell two (FC2) sequentially; the response was measured by subtracting the reference response of FC1 from the binding response of FC2.	7
Figure 1.5	The Biacore X100 system. To study the interaction between the capture agent and the analyte, the sensor chip requires surface immobilization with the capture agent which is then inserted into the sensor chip holder. The continuous flow micro-fluid channel enables analyte flow at the chosen condition and enables real time observation of the capture agent and the analyte.	8
Figure 1.6	The microfluidic system and flow cell of a Biacore X100. Two flow cells, FC1 and FC2 are formed over one sensor surface when the sensor chip is docked in the instrument, and the flow cells can be used in series or individually. The automatic in-line reference substitution was done by using FC2 minus FC1 from same sample injection.	9
Figure 1.7	Each sensor chip consists of a gold coated surface with a different matrix. The CM5 chip is the most versatile and this has a carboxymethylated dextran matrix as a coupling layer.	11

Figure 1.8	The sandwich format also known as positive assay. The assay usually consists of nanoparticles such as gold in either a sample solution or conjugate pad to give signals when it binds to the test and control lines. The sandwich assay gives a signal in the presence of sample/analyte, and the signal is increased as the concentration of target sample/analyte is increased.	14
Figure 1.9	The competitive format also known as negative assay. The analyte specific antibody is usually conjugated with nanoparticles to give signals when it binds to the test line. Anti-analyte antibody or analyte-protein conjugate is commonly used as the test line. As the concentration of the target sample/analyte increases, the analyte binds to the test line. This results in the occupancy of the binding sites of the test line decreases. Hence, the competitive assay gives decreasing signals as the concentration of the target sample/analyte increased. Further details are given in section 1.3.1.	15
Figure 1.10	Schematic diagram of a lateral flow immunoassay strip. The strip consists of a sample pad, conjugate release pad, a membrane where the test and control lines are sprayed and a absorbent pad.	16
Figure 1.11	Microtiter plate for assay development	17
Figure 1.12	Strip tests using a series of standard solutions, gold nanoparticles are commonly used as a indicator and are pink coloured. The image was obtained using a photoscanner then subjected to quantitative analysis. The intensity of the test line is inverse proportional to the concentration of the target analyte.	17
Figure 1.13	Structure of nitrocellulose ester and protein dipoles. Nitrocellulose membranes bind to protein electrostatically through interaction between the dipole of the nitro ester and the dipole of the peptide bonds of the protein.	18
Table 1.1	To study an interaction, it is important to select the most suitable sensor chip for individual studies. There is a range of sensor chips available commercially for specific requirements.	10

Chapter 2

Figure 2.1	Under total internal reflection, all the incoming light is reflected within the prism.	28
Figure 2.2	Schematic of a boundary-value problem to describe the propagation of a surface plasmon polariton.	28
Figure 2.3	Evanescent wave generated under TIR.	29
Figure 2.4	At θ_{SPR} , the reflected light intensity decreases, and this difference is measured in SPR. As the molecule interacts with the immobilised molecule on the surface, the change in refractive index results in a shift in θ_{SPR} .	30
Figure 2.5	The activation steps of the carboxylic acid group of CM5 with EDC and NHS.	32
Figure 2.6	4TP-P4-OVA	33
Figure 2.7	4TPH-P4-OVA	33
Figure 2.8	P4-PEG-OVA	33
Figure 2.9	Buffer conditions for a successful immobilisation of ligands onto a CM5 sensor surface. The pH of the buffer has to be higher than the pK of the dextran layer to generate a negatively charged surface for the positively charged ligand to bind to the surface.	36
Figure 2.10	Sensorgram of flow cell two (FC2) showing the process of activation of the CM5 sensor surface, immobilisation of the P4-PEG-OVA onto a CM5 surface and the deactivation/cleaning of the sensor surface. The P4-PEG-OVA solution was injected after the surface was activated by the EDS/NHS solution. The sensor surface was regenerated/deactivated after the injection of ethanolamine-HCl solution.	38

Figure 2.11	Sensorgram of flow cell one (FC1) showing the process of activation of the CM5 sensor surface, immobilisation of the OVA ligand on the CM5 and the deactivation/cleaning of the sensor surface. The OVA solution was injected in short pulses after the CM5 surface was activated by the EDS/NHS solution and this was followed by an injection of ethanolamine-HCl to regenerate the surface.	38
Figure 2.12	Binding curve of 4TP-P4-OVA with rat monoclonal Anti-P4.	40
Figure 2.13	Binding curve of 4TPH-P4-OVA with rat monoclonal Anti-P4.	41
Figure 2.14	Binding curve of Anti-P4 on a P4-PEG-OVA sensor chip.	42
Figure 2.15	Comparison of binding curves for P4 derivatives.	43
Figure 2.16	Mouse Anti-P4 surface performance varied on two different days (Batch 1 and Batch 2) with the same sensor surface (4TP-P4-OVA), but it is a significantly more sensitive antibody compared with rat Anti-P4.	44
Figure 2.17	Binding performance of mouse Anti-P4 with Run 1 and Run 2 performed 6 months apart. The surface response varied over time, but still gave a relative linear response.	45
Figure 2.18	Binding curve of 4TP-P4-OVA with mouse Anti-P4.	46
Figure 2.19	Binding curve of 4TPH-P4-OVA with mouse Anti-P4.	47
Figure 2.20	Binding curve of P4-PEG-OVA with mouse Anti-P4.	48
Figure 2.21	Mouse Anti-P4 binding curves of 4TP-P4-OVA, 4TPH-P4-OVA, P4-PEG-OVA conjugates.	49

Figure 2.22	4TP-P4-OVA standard curve for P4 concentration extrapolation has a working range of 0.27-2.38 ng/mL.	51
Figure 2.23	The 4TPH-P4-OVA standard curve for P4. The working range has been calculated to be 0.384-0.742 ng/mL.	52
Figure 2.24	A P4 standard curve scouting on a P4-PEG-OVA sensor surface.	53
Figure 2.25	A P4-PEG-OVA standard curve for P4 concentration extrapolation has a working range of 0.294 to 1.942 ng/mL.	54
Figure 2.26	Mouse Anti-P4 standard curves of 4TP-P4-OVA, 4TPH-P4-OVA, and P4-PEG-OVA conjugates.	55
Figure 2.27	Bovine serum binding curve on a P4-PEG-OVA sensor surface.	57
Figure 2.28	The SPR analysed P4 concentration in serum (free P4) and the ECLIA analysed P4 concentrations in serum samples were overlapped to monitor the P4 concentrations over 2 cycles.	58
Figure 2.29	The surface responses of blank extraction, hexane extraction and ethyl acetate extraction of serum samples. The surface response is inverse proportional to the concentration of P4 in the sample. Hence, the ethyl acetate extracted sample presented the highest response compared with the blank (buffer) and hexane extracted samples.	59
Figure 2.30	Surface response for a hexane extracted serum sample and a THF extracted serum sample.	60
Figure 2.31	The P4 concentration of top (black) and bottom (red) fraction of the extracted and filtered serum samples.	61
Figure 2.32	The combined P4 concentration from extracted serum samples.	62

Figure 2.33	The independent analysis results overlapped with the SPR analysis results. The plot indicated that the SPR P4 level and the independent analysed P4 level have the same trend.	63
Figure 2.34	[P4] in bovine milk 1 analysed by SPR.	65
Figure 2.35	Overlapped spectra of SPR analysed milk 1 [P4] and ELISA analysed milk 1 [P4].	66
Figure 2.36	[P4] in bovine milk 2 analysed by SPR.	67
Figure 2.37	Spectra of SPR analysed milk 2 [P4] and ELISA analysed milk 2 [P4].	67
Figure 2.38	[P4] in bovine milk 3 analysed by SPR.	68
Figure 2.39	Spectra of SPR analysed milk 3 [P4] and ELISA analysed milk 3 [P4].	69
Table 2.1		55
Scheme 2.1	The synthesis of the P4-PEG derivative (6) was carried out following the scheme above.	34
Scheme 2.2	Steps for the conjugation of P4-PEG-COOH to OVA to form P4-PEG-OVA.	35
 Chapter 3		
Figure 3.1	Development of a one step assay with Anti-P4-Au conjugates. P4-PEG-OVA and rat anti IgG served as test and control lines.	78
Figure 3.2	The image of the strips were obtained by using a photoscanner. The intensity of control and test lines on the test strips were read and quantified using the strip reader system.	79

Figure 3.3	A typical shape of a dose-response curve where the response depends on the dose of the drug (drug concentration).	80
Figure 3.4	UV/visible spectra of an antibody titration, where the λ_{\max} of the blank Au solution was at 523.5 nm.	82
Figure 3.5	P4-PEG-OVA strips with varied Anti-P4-Au concentrations (strips 1 to 12, 0.1 $\mu\text{g}/\text{mL}$ to 1.2 $\mu\text{g}/\text{mL}$). As the concentration of Anti-P4-Au increased from A1 to A12 the intensity of both test and control lines increased.	84
Figure 3.6	A P4-PEG-OVA binding curve indicated 0.1~0.3 $\mu\text{g}/\text{mL}$ of Anti-P4-Au should be sufficient for assay development.	84
Figure 3.7	A P4 standard curve scouting to establish the concentrations for P4 standard solutions.	86
Figure 3.8	The P4 standard curve obtained from the P4-PEG-OVA test strips. The standard curve was calibrated with triplicate measurements. The error bars represented the standard deviation of the measurements.	87
Figure 3.9	Intensity of control serum samples after protein separation using a centrifugal filter. The results indicated the concentrations of the free P4 present in serum samples may be too low for the sensing strip to detect.	88
Figure 3.10	P4 concentrations of cow 3 in spun down serum samples.	89
Figure 3.11	The serum samples from cow 3 were tested and the results compared with the ECLIA analysis.	90
Figure 3.12	P4 standard curves to test the matrix effect from the serum sample as well as the sensitivity of the assay by using different sets of P4 standards.	91
Figure 3.13	Serum with no standards fitted within the working range of the P4 standard curve.	92

Figure 3.14	Comparison between a normalized P4 standard curve and a normalized P4 standard curve with spiked low [P4] serum sample.	93
Figure 3.15	P4 standards were spiked with a serum sample which contained a low concentration of P4.	94
Figure 3.16	LFIA analysis of serum 1, which showed a distinct P4 cycle.	94
Figure 3.17	ECLIA analysis of serum from cow 1, which was used as a standard to validate the assays. The serum samples showed one complete P4 cycle with the two ends of the plot having high P4 concentration and low P4 concentrations in between, and this followed the normal trend for a bovine P4 cycle.	95
Figure 3.18	Overlapped plots of ECLIA analysis and LFIA analysis of serum 1.	96
Figure 3.19	Serum samples from cow 1 repeated for P4 concentration 6 months later. The pro-estrus and metestrus profile from LFIA analysis matched the profile from ECLIA analysis.	97
Figure 3.20	Overlapped LFIA P4 cycle and ECLIA P4 cycle for Serum 2.	98
Figure 3.21	The assay using milk samples (the strips were numbered from 1 to 12 according to their sample collection date) without treatment/preparation resulted in coagulation of the milk fat/protein with the Anti-P4-Au at the bottom of the strip. The bottom of samples 6 to 11 showed the fat and protein deposits.	99
Figure 3.22	Comparison of [P4] from two batches of LFIA analysis using diluted milk samples from cow 1. The two batches analysed from the same assay were not consistent. Therefore the milk samples required further treatment.	100
Figure 3.23	NZVP milk and serum samples of cow 1 did not have the same P4 cycle profile.	101

Figure 3.24	Standard curve spiked with a milk sample.	103
Figure 3.25	The sensor strips (the strips were numbered from 1 to 12 according to their sample collection date) from fat-removed, diluted milk samples.	104
Figure 3.26	The P4 standard curve was used to calculate the P4 in milk samples from cow 1.	105
Figure 3.27	The sensor strips (the strips were numbered from 13 to 34 according to their sample collection date) were allowed to develop in the solution well which contained the analysed solution for 20 min. The strips were scanned and analysed using the strip reader. The variation in colour intensity of the sample strips suggested a difference in [P4] in the milk samples from cow 1.	106
Figure 3.28	The P4 cycle from cow 1 analysed from LFIA, the samples showed only a partial estrous cycle which resembled a typical bovine estrous cycle.	107
Figure 3.29	The LFIA analysed P4 cycle was overlapped with the ELISA analysed P4 cycle of cow 1.	108
Figure 3.30	LFIA analysed milk sample (after removal of fat and protein) showed a full P4 cycle of cow 1.	109
Figure 3.31	Overlapped ELISA and LFIA P4 cycles of cow 1. The profiles of the P4 cycles matched but with an overestimation of the P4 concentration from LFIA.	110
Figure 3.32	LFIA analysis of milk samples from cow 2 after protein and fat removal.	111
Figure 3.33	Overlap of the LFIA P4 cycle and the ELISA P4 cycle. The general trend of the P4 cycle was demonstrated by using the LFIA method for milk sample analysis on cow 2.	112

Figure 3.34	A repeat of the P4 cycle of cow 2 analysed by LFIA with newly conjugated Anti-P4-Au.	113
Figure 3.35	Overlap of the repeat LFIA analysed P4 cycle and ELISA analysed P4 cycle for cow 2.	113
Table 3.1	Antibody titration concentrations	81
Table 3.2	Absorbance and λ_{\max} obtained from antibody titration	82

Chapter 4

Figure 4.1	A) Proposed structure of a polyester granule. The small water insoluble inclusions are formed with a amorphous polyester core with polyester synthase covalently attached to the surface. B) Electron microscopy image of <i>Pseudomonas aeruginosa</i> accumulating polyester granules.	121
Figure 4.2	Model of polyester granule formation. A) Micelle model, and B) Budding model.	122
Figure 4.3	Genetically fused 3R-GBP-1, bead formation enzyme (polyester synthases) and extra binding domain protein (IgG) enable the formation of polyester inclusion within the bacterial cell.	123
Figure 4.4	TEM images of PHA beads after incubation with colloidal gold, with PHA beads A displaying the ZZ(-)PhaC protein at the surface and PHA beads B containing the engineered protein 3xGN-G ₅ -ZZ(-)PhaC on the surface with gold nanoparticles. Figure A shows no gold attached but Figure B does.	124
Figure 4.5	Scheme of the surface of a bifunctional gold/antibody binding PHA bead.	124
Figure 4.6	There are many possible applications for GEPI, including biosensing studies.	125

Figure 4.7	Binding affinity of a ZZPhaC polyesters for various antibodies, where (----) represents weak binding affinity, and (++++) represents strong binding affinity.	126
Figure 4.8	The dip coating set up for 3GNZZPhAC beads. The Au coated mylar chip was dip coated with 3GNZZPhAC beads followed by washing of the surface with Milli-Q H ₂ O. To reduce aggregation of the 3GNZZPhAC beads, the bead solution was under continuous stirring while the Au chip underwent the dip coating process.	128
Figure 4.9	Flow cell with an inlet and an outlet and a single channel.	129
Figure 4.10	Glass surfaces coated with gold acting as a sensor chip for the SPR sensing system.	130
Figure 4.11	Reference glass slide with double-sided tape.	131
Figure 4.12	The flow cell channel was sealed with the reference glass slide and with inlet and outlet tubes attached to the flow cell system.	132
Figure 4.13	The flow cell channel was sealed with the Au coated glass slide and with inlet and outlet tubes attached to the flow cell system.	132
Figure 4.14	Flow cell system setup. The single flow cell system with reference glass slide was attached to syringe pump from the inlet tube and injected with a 3GNZZPhAC bead solution at 2 mL/hr. The residue solution flowed through the flow cell channel and exited from the outlet tube.	133
Figure 4.15	The cores of 3GNZZPhAC beads were stained with Nile red dye then immobilised onto the gold surface. The fluorescence image (40x magnification) of 3GNZZPhAC beads after immobilisation showed they were still present after the washing process.	134

Figure 4.16	A SEM image (8,000x magnification) of 3GNZZPhaC beads after surface immobilisation. The larger 3GNZZPhaC beads were embedded within the solvent residues/ contaminants.	135
Figure 4.17	Aerial view of an SEM image (13,000x magnification) of 3GNZZPhaC beads after surface immobilisation. There were single layer beads scattered on the surface after cleaning. However, there were residues/ contaminants that still remained.	135
Figure 4.18	Close up (tilted 90°) SEM image (30,000x magnification) of 3GNZZPhaC beads after surface immobilisation. It's quite clear that the beads are attached to the surface with the residues/ contaminants "sticking" to it.	136
Figure 4.19	Aerial view of an SEM image (13,000x magnification) of ZZPhaC beads after surface immobilisation. The ZZPhaC beads are coagulated on the Au surface.	136
Figure 4.20	SEM image (13,000x magnification) of ZZPhaC beads after surface immobilisation. Some of the beads scattered on the surface with no aggregation.	137
Figure 4.21	Close up (tilted 90°) of an SEM image (25,000x magnification) of ZZPhaC beads after surface immobilisation. The scattered single beads were attached to the Au surface with no solvent residues after cleaning the surface (residues showed in a 3GNZZPhaC sample).	137
Figure 4.22	The Asylum Research MFP-3D model was used for surface profiling.	138
Figure 4.23	Surface profiling of the 3GNZZPhaC beads.	139
Figure 4.24	Surface profiling of 3GNZZPhaC beads on a Au coated surface.	140
Figure 4.25	Schematic diagram of a SPR sensor with immobilised PHA beads. The multifunctional PHA beads are able to bind to the gold surface as well as the analyte.	141

Figure 4.26	Sensorgram of rat Anti-P4 binding to the Au-ZZPhaC surface in FC1.	143
Figure 4.27	Sensorgram of rat Anti-P4 binding to the Au-ZZPhaC surface in FC2.	144
Figure 4.28	The rat Anti-P4 was tested on a ZZPhaC surface with a flow rate of 5 $\mu\text{L}/\text{min}$.	145
Figure 4.29	The rat Anti-P4 was tested on a ZZPhaC surface with a flow rate of 10 $\mu\text{L}/\text{min}$.	146
Figure 4.30	Binding performance of rat Anti-P4 on a ZZPhaC sensor surface with a flow rate of 5 $\mu\text{L}/\text{min}$. The sensor surface was immobilised outside of the SPR instrument, hence the FC1 and FC2 surface response units should be relatively similar.	147
Figure 4.31	The ZZPhaC beads immobilised on the commercially available Au sensor surface. The surface appeared to have a large amount of salt present which was from the buffer residues (40x magnification). FC1 and FC2 are visibly coated with salt residues and ZZPhaC beads.	148
Figure 4.32	The sensor surface with ZZPhaC beads immobilised outside of the SPR instrument. The beads were scattered along the gold surface with aggregation (500x magnification).	149
Figure 4.33	Outside surface immobilisation of ZZPhaC beads; a significant amount of aggregation occurred (15,000 magnification) and this would hinder the binding response of the Anti-P4.	149
Figure 4.34	After the first injection of ZZPhaC beads (1 ng/mL) at 5 $\mu\text{L}/\text{min}$ for 24 s on FC2, there appeared to be no binding on the Au surface.	150
Figure 4.35	The short pulse injections of ZZPhaC (1 ng/mL) at 5 $\mu\text{L}/\text{min}$ showed a very low surface response.	151
Figure 4.36	The immobilisation concentration of the ZZPhaC beads was increased to 10 ng/mL. The immobilisation step began with a short pulse of injection	

followed by a long pulse injection. The surface loading was checked by several short pulse injections. The final immobilisation level was 522.7 RU.

	152
Figure 4.37	Surface response of ZZPhaC bead immobilised FC2 and bare gold FC1. The binding performance of rat Anti-P4 on both channels was very similar; with a higher concentration of rat Anti-P4 the surface response was lower than the bare gold surface.
	153
Figure 4.38	The Au-ZZPhaC sensor chip was used to repeat the binding curve of rat anti progesterone, where FC1 contained only bare gold and FC2 was immobilised with ZZPhaC beads.
	154
Figure 4.39	The SEM image (40x magnification) of the SPR sensor chip with ZZPhaC beads. FC2 was immobilised with ZZPhaC beads and FC1 was the reference cell without a binding layer (bare gold).
	156
Figure 4.40	The SPR immobilisation method reduced the aggregation (4,000x magnification) of the ZZPhaC beads and allowed rat Anti-P4 binding to occur.
	157
Figure 4.41	A preliminary assay using 3GNZZPhaC beads and ZZPhaC beads as binding material.
	159
Table 4.1	145
Table 4.2	146
Table 4.3	153
Table 4.4	155
Table 4.5	158

Abbreviations

4TP-P4	3-(pregn-4-ene-3,20-dione-4-yl)thiopropionic acid
4TPH-P4	6-[3-[(pregn-4-ene-3,20-dione-4-yl)thiopropano-yl]amino]hexanoic acid
AFM	atomic force microscopy
Anti-P4	progesterone antibody
AuNPs	gold nanoparticles
BSA	bovine serum albumin
C	control line
CBG	corticosteroid binding globulin
CM5	carboxymethylate dextran
CL	corpus luteum
DCC	1,3-dicyclohexylcarbodiimide
DMF	dimethylformamide
DNA	deoxyribonucleic acid
dpm	disintegration per minute
EC20	the lowest concentration that can be distinguished from the background noise
EC50	the half maximal effect concentration
EC80	the highest concentration that can be distinguished from the background noise
ECLIA	electrochemiluminescence immunoassay
EDC	1-ethyl-3-(3-dimethylaminopropyl)carbodiimide
EDTA	ethylenediaminetetraacetic acid
EIAs	enzyme immunoassays
ELISA	enzyme-linked immunosorbent assay
FC1	flow cell one

FC2	flow cell two
FSH	follicular stimulating hormone
GC-MS	gas chromatography–mass spectrometry
GEPIs	genetically engineered polypeptides for inorganics
HBS-EP ⁺	SPR buffer contained 0.1 M HEPES, 1.5 M NaCl, 30 mM EDTA and 0.5% v/v Surfactant P20
hCG	gonadotropin
HEPES	4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid
HPLC	high performance liquid chromatography
IFC	integrated micro fluidic cartridge
IgG	immunoglobulin G
kDa	kilodalton
LC-MS	liquid chromatography–mass spectrometry
LFDs	lateral flow devices
LFIA	lateral flow immunoassay
LH	luteinising hormones
LOD	limit of detection
NHS	N-hydroxysuccinimide
NZVP	New Zealand Veterinary Pathology Limited
P4	progesterone
P4-PEG	<i>N</i> -(13-(carbonylamino)-4,7,10-trioxatridecanyl)- 3-(pregn-4-ene-3,20-dione-4-yl)thiopropamide
PHA	polyhydroxyalkanoate
PHB	polyhydroxybutyrate
PHBA	poly(3-hydroxybutyrate-co-3-hydroxyvalerate)
pI	isoelectric point
pK	disassociate constant

OVA	ovalbumin
RIAs	radioimmunoassays
RU	response unit
SEM	scanning electron microscopy
SPR	surface plasmon resonance
SPRI	surface plasmon resonance imaging
T	test line
THF	tetrahydrofuran
TIR	total internal reflection
ZZ domain	antibody binding domain of protein A
θ_{SPR}	surface plasmon resonance angle
λ_{max}	lambda(max)

