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Effects of High Pressure on DNA and its Components

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Abstract

There have been many speculations for the environment in which life originated but it has still yet to be determined what environmental chemical and physical conditions were necessary for the evolution of self-replicating chemical systems. While it has been determined that DNA, RNA and their components are chemically unstable at high temperatures, there has currently been only a small number of studies into the role of high pressures on the chemical and physical stabilities.

High-pressure NMR spectroscopy has been used here to study the effects of high temperatures/pressures on the chemical stability of DNA and its components. This has been done with the use of a specialised commercial high-pressure NMR cell capable of withstanding pressures up to 250 MPa. In addition to this, a custom safe handling apparatus and pump system was developed for the operation of this cell.

Studies into the effects of high pressures on the rate of hydrolysis of cytosine and cytidine at 100 °C were performed by measuring the rates of hydrolysis with time under various pressure conditions. These results have shown that the rates of hydrolysis of cytosine and cytidine increase considerably with pressure.

The effects of high pressure on the physical stability of DNA were determined by performing dissociation (melting) experiments on several different DNA sequences under multiple pressure conditions. It was found that the melting point of a small DNA hexamer decreased slightly with pressure whereas the melting points of larger dodecamers increased overall with pressure. It was also found that the melting point of an i-motif structure decreased with increasing pressure.

The effects of high pressure on the chemical stability of cytosine were again studied, this time for cytosine residues within both single- and double-stranded DNA. DNA samples for bacteriophage ΦX174 were incubated under various temperature and pressure conditions. Results for these studies have yet to be determined as the incubated DNA is yet to be sequenced.

It has been discovered that high pressures have a negative effect on the chemical stability of DNA constituents while having an overall small positive effect on the physical stability of DNA.

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List of Nomenclature and Abbreviations

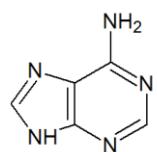
Nomenclature

The building blocks of DNA/ RNA

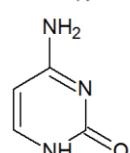
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Common Bases

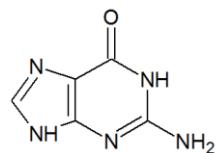
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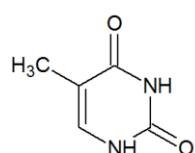
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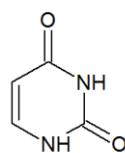
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- Thymine

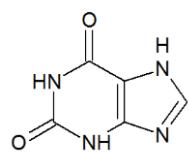


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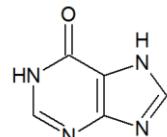


Other Bases

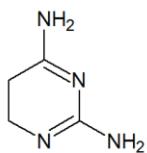
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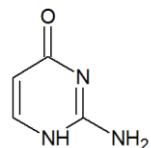
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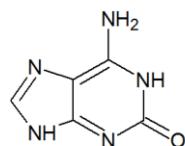
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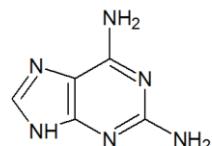
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- Isoguanine

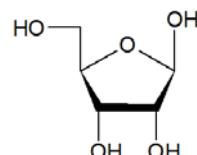


- Diaminopurine

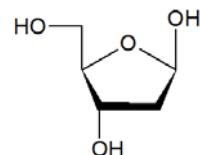


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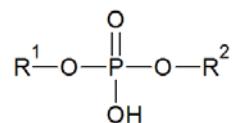
- Ribose



- 2-Deoxyribose

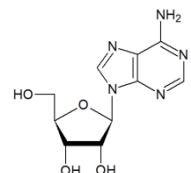


- Phosphodiester

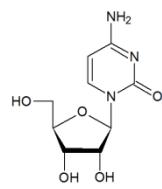


- Nucleosides

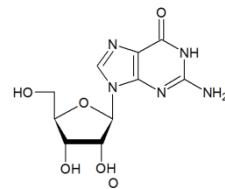
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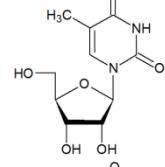
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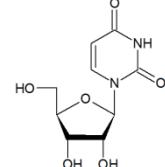
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- Thymidine



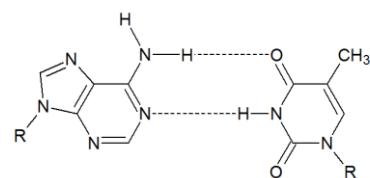
- Uridine



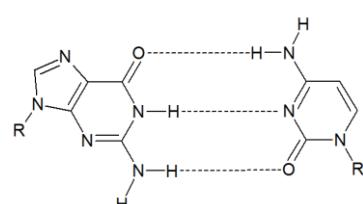
- Base pairing

Watson-Crick

- A-T

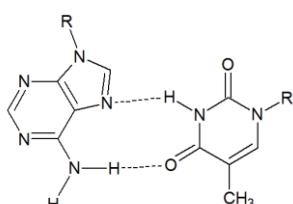


- G-C

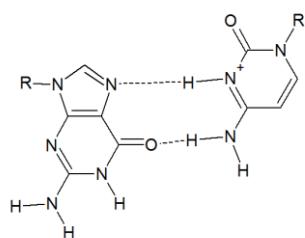


Hoogsteen

- A*T



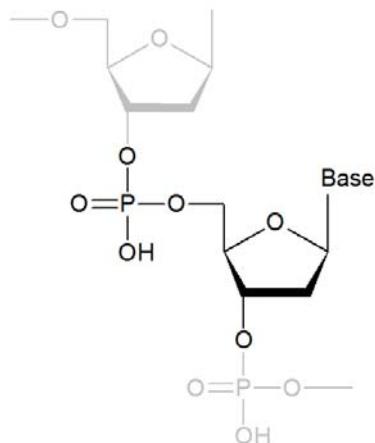
- G*C⁺



Nucleic Acids

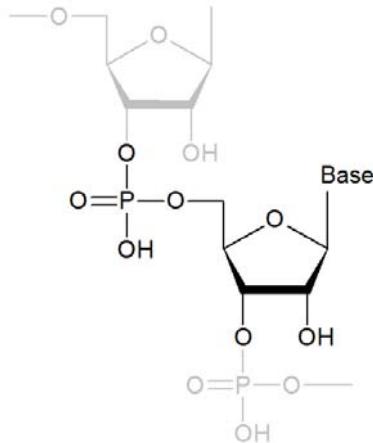
- DNA - Deoxyribonucleic acid. Each DNA nucleotide is comprised of a nucleobase, a phosphate and a 2-deoxyribose sugar. The phosphate and sugar form the backbone of DNA. The nucleobases are attached to the anomeric carbon of the sugar unit.

- DNA nucleotide



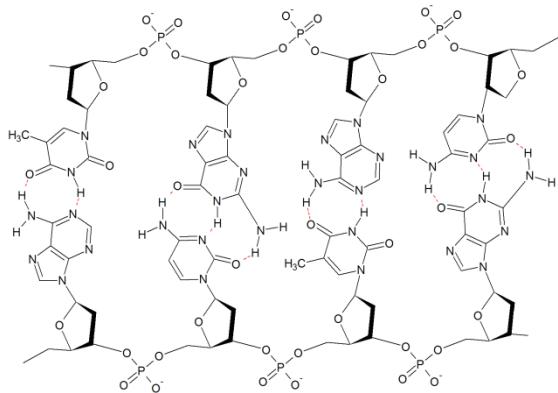
- RNA - Ribonucleic acid. Each RNA nucleotide is comprised of a nucleobase, a phosphate and a ribose sugar. The phosphate and sugar form the backbone of RNA. The nucleobases are attached to the anomeric carbon of the sugar unit.

- RNA nucleotide



- Nucleic acid structure. A series of nucleotides joined to form a oligonucleotide chain. Watson-Crick base pairing between two

complementary oligonucleotides under standard biological conditions results in the DNA structure as shown below. Three dimensionally this is a right-handed double helix with approximately 10.5 bases per turn.

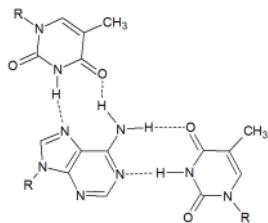


- Other DNA structures

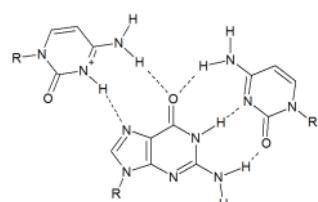
Triplexes. A triplex is formed when a C/T rich segment of a DNA sequence folds back along an A/G rich segment. This results in a triplex comprised of a pair of strands bonded by Watson-Crick pairings with a third strand bound via Hoogsteen pairings to the A/G rich strand involved in the Watson-Crick pairing.

 - Triplex pairing

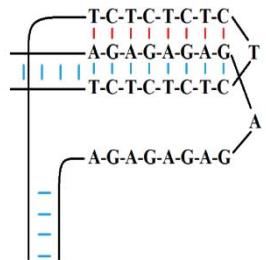
T-A*T



C-G*C⁺

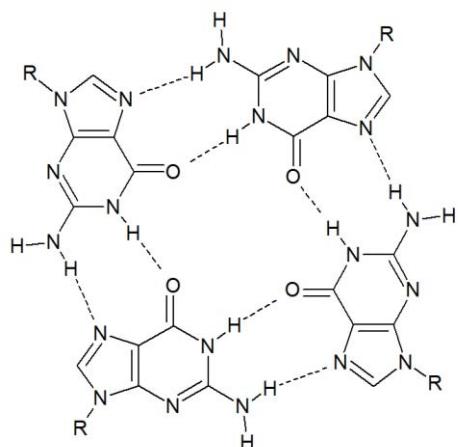


- Triplex Structure

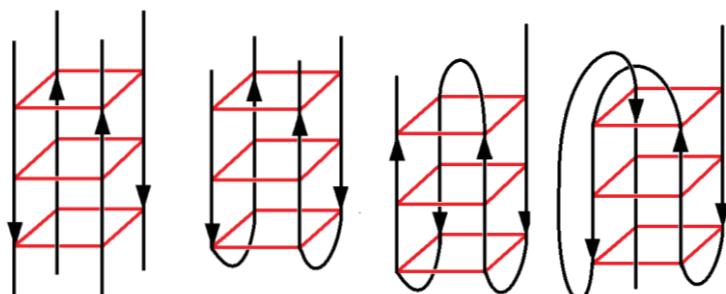


G-Quadruplexes. G-quadruplexes are a tetrad structure formed by four, G-rich sequences. Four guanine bases undergo Hoogsteen type bonding to form a G tetrad. These tetrads stack to form a quadruplex with multiple topologies.

- G-quadruplex tetrad

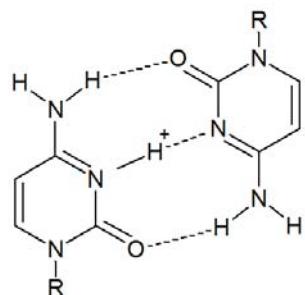


- G-quadruplex Topologies

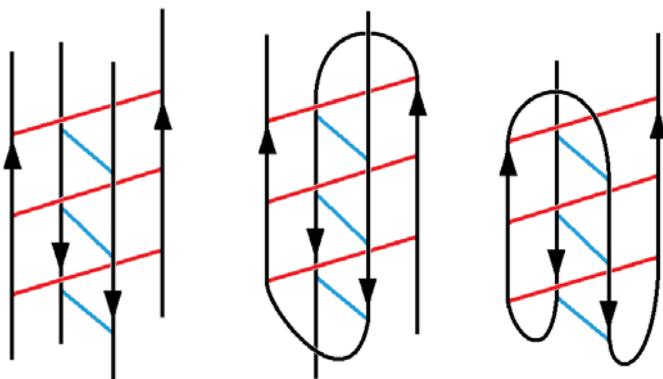


i-Motifs. i-Motifs are a tetrad structure formed by four, C-rich, DNA sequences. The structure is formed from two sets of antiparallel stranded duplexes connected by C-C⁺ Hoogsteen pairings. These pairings are intercalated between the two duplexes resulting in the four stranded structures.

- i-Motif pairing



- i-Motif structures



Abbreviations

CD spectroscopy	- Circular Dichroism spectroscopy
COSY	-Correlation Spectroscopy
dNTPs	- deoxyribonucleotide Triphosphates
HiP	- High Pressure Equipment Company
NMR spectroscopy	- Nuclear Magnetic Resonance spectroscopy
NOESY	-Nuclear Overhauser Effect Spectroscopy
RF	- Radio Frequency
SBS	- Sequencing By Synthesis
Sequence 121	- DNA sequence 5'-CAT TTA TAA ATG-3'. 16.7 % G-C
Sequence 122	- DNA sequence 5'-CAT TCT AGA ATG-3'. 33.3 % G-C
Sequence 123	- DNA sequence 5'-CAA GTC GAC TTG-3'. 50 % G-C
Sequence 124	- DNA sequence 5'-CAG GTC GAC CTG-3'. 66.7 % G-C
Sequence 125	- DNA sequence 5'-CAC CCG CGG GTG-3'. 83.3 % G-C
T_m	- Melting temperature
TMSP-d	- 3-(trimethylsilyl)-2,2',3,3'-tetradeuteropropionic acid (NMR reference)