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FUSARIA AND FUSARIUM TOXINS

IN MAIZE

A thesis presented in partial fulfilment of the requirement for the degree of Doctor of Philosophy in Veterinary Pathology and Public Health at Massey University

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ABSTRACT

Many species of <u>Fusarium</u> are commonly associated with cereals, particularly maize, but in New Zealand, little is known of their significance as mycotoxin producers. These studies have examined the prevalence of fusaria and other fungi in maize and maize fields and have investigated the presence and sources of some major <u>Fusarium</u> toxins in maize.

Fungi in maize, husk, litter and soil from maize fields and in grain at harvest and in storage were assayed. The distribution of fungi was found to be uneven within maize, husk and litter substrates within a field but in soil was more homogeneous. Sampling techniques were therefore developed to ensure representative subsamples were obtained from each source.

Similarly isolation procedures were chosen to ensure adequate recovery of fungi. Dilution and direct platings were used to provide information on total populations and on fungi actually invading kernels, with two media, PDA-D and PCNB. The medium used showed no significant influence on either viable counts or kernel contamination rates nor on the number of different <u>Fusarium</u> spp recovered on the two media, but PDA-D supported a greater overall variety of fungi. The numbers of genera and of <u>Fusarium</u> spp recorded by direct plating were significantly higher than with dilution plating.

The total population and the number of different genera and of $\underline{Fusarium}$ spp were compared for the four "field" substrates. A total of 25 genera was isolated, most being recovered from soil and litter. $\underline{Fusarium}$ was present in all samples. $\underline{Acremonium}$, $\underline{Cladosporium}$, $\underline{Penicillium}$ and \underline{Mucor} occurred regularly. The four substrates gave up to ten different $\underline{Fusarium}$ spp, $\underline{F.}$ $\underline{graminearum}$, $\underline{F.}$ $\underline{culmorum}$ and $\underline{F.}$ $\underline{acuminatum}$ being the most frequent. Husk and litter samples gave the highest viable counts for both total fungi and $\underline{Fusarium}$ spp.

Field samples of maize kernels showed 13 genera and ten <u>Fusarium</u> spp. At harvest time total genera increased to 17 but <u>Fusarium</u> spp remained constant. While the total genera remained constant at 17 in stored samples, the number of <u>Fusarium</u> spp dropped to three, only F.

<u>subglutinans</u>, <u>F. graminearum</u> and <u>F. poae</u> being detected. The contamination rate of kernels by fusaria also changed significantly from field samples (75.8%) to harvest samples (58.3%) to only 1.5% in stored maize.

As with <u>Fusarium</u>, <u>Acremonium</u> and <u>Mucor</u> populations decreased from harvest to storage but other genera (e.g. <u>Aspergillus</u>, <u>Beauvaria</u>) were only found in stored maize. The frequency of occurrence of <u>Penicillium</u> remained stable over the whole period.

Three analytical methods, TLC, GC and GC-MS were used for screening maize, poultry ration samples and cultures of <u>Fusarium</u> isolates for five <u>Fusarium</u> toxins. The GC-MS method was the most reliable and sensitive for detection and quantitation of DON, DAS and T-2 toxin, but not for quantitation of ZEA, due to derivatisation problems. TLC and TLC-densitometry were sensitive and reliable enough for detection and quantitation of ZEA and MON respectively. Although the GC results were closer to the GC-MS results, a high percentage of false positives, particularly for T-2 toxin, was noticed.

Of the examined maize samples, 85% were contaminated with fungal toxins. The majority contained ZEA and three samples were each contaminated with four toxins. No MON was detected.

Many isolates, particularly of $\underline{F.}$ graminearum, were found to be ZEA-producers. Some 63% produced ZEA at >2 ppm. T-2 toxin was produced by 46% of the isolates but at low levels (<1.7 ppm). Low levels of DON and DAS were produced by a few isolates. MON was produced by 30% of isolates, particularly $\underline{F.}$ subglutinans, and in large amounts (up to 64 ppm).

This thesis is the first report on the natural occurrence of <u>Fusarium</u> toxins in New Zealand maize. T-2 toxin and DAS have not been reported as natural contaminants in this country. MON production has also not been reported in New Zealand.

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ABBREVIATIONS USED IN THIS THESIS

3-ADON 3-acetyl-deoxynivalenol 15-acetyl-deoxynivalenol 15-ADON

N, O-Bis (trimethylsilyl) acetamide BSA

N, O-Bis (trimethylsilyl) trifluoroacetamide BSTFA

Colony-forming units per gram CFU/q

CDA Czapek-dox solution agar CLA Carnation leaf agar DAN Diacetylnivalenol DAS Diacetoxyscirpenol

2,4-DNPH 2,4-dinitrophenylhydrazine

DON Deoxynivalenol

ECD Electron capture detector

ΕI Electron impact

FID Flame ionization detector

FUS-X Fusarenon-X

Gas liquid chromatography GC

GC-ECD Gas liquid chromatography with electron capture detector GC-FID Gas liquid chromatography with flame ionization detector

Gas chromatography-mass spectrometry GC-MS

GYFP Glucose-yeast-extract-peptone

HFB Heptofluorobutyryl

HFBI Heptofluorobutyryl-imidazole Initial moisture content IMC LEM Leukoencephalomalacia MAS Monoacetoxyscirpenol MC Moisture content MID Multiple ion detection

MON Moniliformin

Mycotoxin standards mixture MTM

Mass spectrometry MS

MS-MS Mass spectrometry-mass spectrometry

Mass/charge ratio of ion fragments in mass spectrometry M/Z

Neosolaniol NEO Nivalenol NIV

Pentachloronitrobenzene PCNB PDA Potato dextrose agar

PDA-D Potato dextrose agar-dichloran

ppb Part per billion (ng/g) Part per million (mg/kg) ppm PSA Potato sucrose agar RIA Radioimmunoassays

Sterile distilled water sdw Selective ion monitoring SIM TAS

Triactoxyscirpenol

TLC Thin layer chromatography

TMS Trimethylsilyl

N-trimethylsilyl-imidazole TMS I **TCMS** Trimethylchlorosilane TIM Total ion monitoring

ZEA Zearalenone