Copyright is owned by the Author of the thesis. Permission is given for a copy to be downloaded by an individual for the purpose of research and private study only. The thesis may not be reproduced elsewhere without the permission of the Author.

THE ONSET OF PUBERTY AND HERBAGE INTAKE IN DIFFERENT SELECTION LINES OF ANGUS CATTLE

A thesis presented in partial fulfilment of the requirements for the degree of

Master of Applied Science (MApplSc)

in Animal Science



Institute of Veterinary, Animal and Biomedical Sciences

Massey University

Palmerston North, New Zealand

José María Martínez-Marecos

This thesis is dedicated to my parents

José Ramón Martínez and

Elisa Mercedes Marecos de Martínez

ABSTRACT

Martínez-Marecos, J. M. (2003). The onset of puberty and herbage intake in different selection lines of Angus cattle. MApplSc Thesis, Massey University, Palmerston North, New Zealand. 121 pp.

Puberty onset and herbage dry matter intake was measured in four lines of Angus cattle selected for High and Low EBV-600 day weight (HG and LG respectively) and High and Low EBV-milk (HM and LM respectively). The heifers were generated on four industry farms in the North Island of New Zealand.

Heifers from the HG line were younger at puberty (438.6 \pm 9.3 days P < 0.01) than heifers from the HM (459.4 \pm 7.1 days) and LM (476.1 \pm 7.8 days) lines. No difference in age at puberty between HG and LG (455.1 \pm 11.8 days) lines were found. Average weight at puberty across all genetic lines was (349.6 \pm 9.9 kg). There was no difference among the four genotypes for this trait.

Ninety-five percent of the animals reached puberty by the end of the trial and there was no difference in the percentage of animals reaching puberty by genetic line. Animals coming from Farms 1 and 3 tended to reach puberty earlier than animals coming from Farms 2 and 4. There was no differences in pregnancy rate among the genetic lines and the overall pregnancy rate was 90 %.

Intake was measured on two occasions using n-alkanes (M1 and M2 respectively) and the pre- and post-grazing technique (M3 and M4 respectively). Average liveweight (LW) and estimated herbage intake at M1 was 240.2 ± 0.4 kg and 3.47 ± 0.1 kg DM respectively. The values at M2 were 287.2 ± 1.9 kg LW and 6.50 ± 0.36 kg DM. No differences in estimated herbage intake among the genetic lines were detected in M1 or

M2. In M3 heifers had an average LW of 247.1 ± 0.7 kg and mean estimated intakes of 4.86 ± 0.26 ; 4.17 ± 0.26 ; 4.37 ± 0.26 and 3.00 ± 0.26 kg DM for the HG, LG, HM and LM lines. The LM line having a significantly (P < 0.05) lower estimated intakes than the other lines. Average LW at M4 was 272.5 ± 0.6 kg. Animals from the LM (7.28 \pm 0.19 kg DM) line had significantly (P < 0.05) higher intakes than animals from the LG (6.52 kg \pm 0.18 DM) and HM (6.71 \pm 0.18 kg DM) lines. Intakes from the HG (6.99 \pm 0.18 kg DM) animals was not significantly different from intakes of the other genetic lines.

In general the HG heifers outperformed the heifers from the other lines in puberty onset and feed conversion efficiency. However, in a self-replacing beef cowherd, the higher maintenance cost and lower milk production of dams from the HG line should be considered. Combining the growth characteristics of the HG lines with an appropriate level of milk production into a selection index would produce animals with the adequate combination of genes for a self-replacing beef cowherd.

ACKNOWLEDGEMENTS

This work has been a wonderful learning experience. Besides the knowledge and understanding of the production system, it taught me persistency, perseverance and patience. First I would like to express my gratitude to my chief supervisor Associate Professor Steve Morris for his encouragement and for keeping everything in perspective. Thanks Steve for reading my draft scripts endless times!!!!!!

I am also deeply grateful to my other supervisors Associate Professor Tim Parkinson and Dr. Patrick Morel. I am also in debt with Nicolas López-Villalobos who helped me with the intricacies of statistical analysis. Thanks to Dr. Corey Matthew for the interesting conversations on pasture production. I wish to thank Mr. Graeme Poole for his assistance in running this trial and the management of the cattle involved. We really had fun times at Keebles farm.

I am deeply thankful to the New Zealand government who supported my studies throughout the New Zealand Official Development Assistance (NZODA). Special thanks to Sylvia Hooker for her support during my studies. This project was funded by Meat and Wool Innovation to whom I am grateful.

I would like to thank Professor Colin Holmes, Professor John Hodgson, Mr. Parry Matthews and their wives Ms Dorothy Holmes, Ruth Hodgson and Frances Matthews for their friendship during my stay in New Zealand.

I have known so many wonderful people during these two years that it will be impossible to name them all here. However, I should mention Carlos and Karin Hug de Belmont, Stuart and Daniela Brander, Juan Antonio Canumir and family, Ignacio

Lizarraga and Fernanda, Luciano do Rego Monteiro,.Alfredo Ordoñez and Federico Sapriza.

Thanks to two special friends in Paraguay, Carlos Alberto Lezcano and Pedro Luis Paniagua, for your support!!!!!

I must also express my immense gratitude to my family. My parents José Ramón Martínez and Elisa Mercedes Marecos de Martínez who gave me the education and the means to be here. My brothers and sisters Pedro, Rebeca, Edgar and Mechi for being there. All my love to my beautiful nieces Andrea and Veronica and my nephew Emmanuel. Finally to my two other family, Roberto and Eli.

Finally I would like to thanks my dearest Patricia, the lovely brazilian girl whom I met 15.000 kilometres away from home. All my love and gratitude for helping me during this project. I would still be writing the introduction if it weren't for you. Thanks for all!!!! Te quiero mucho!!!!!!.

TABLE OF CONTENTS

Abstract		ii:
Acknowled	lgements	v
Table of Co	ontents	vi
List of Tab	les	x
List of Figu	ıres	xiii
CHAPTER 1	INTRODUCTION	1
CHAPTER 2	LITERATURE REVIEW	3
	IMPORTANCE OF AN EARLY PUBERTY	
	Raising animals to calve at two years of age	
2.1.1.		
2.1.2.		
2.1.3.		
	ERTY DEFINITIONS	
	CHANISMS REGULATING THE ONSET OF PUBERTY	
2.3.1.		
2.3.1.		
2.3.3.		
	MONAL MECHANISMS OF THE OESTROUS CYCLE	
	ERTY AND FIRST BEHAVIOURAL OESTRUS	
	Non-pubertal oestrus	
2.5.2.		
	LUENCE OF THE DAMS' GENETIC MAKE-UP ON THE SYSTEM	
2.6.1.	Mature size and reproduction	
2.6.2.		
	NAGING THE HEIFERS TO CALVE AT TWO YEARS OLD	
	Growth before weaning	
2.7.2.	Post-weaning growth	
2.7.3.	Time and rate of gain	
2.8. MEA	ASURING HERBAGE INTAKE	
2.8.1.		
	1.1. Using n-alkanes as markers.	
	1.2. Advantages of n-alkanes over other markers	
	1.3. Choosing an n-alkane	
	1.4. Recovery of alkanes in the faeces	

CHAPTER 3 MATERIALS AND METHODS	39
3.1. EXPERIMENTAL LAYOUT	39
3.2. BACKGROUND TO ANIMALS	
3.3. ANIMAL MANAGEMENT	41
3.4. PUBERTY DETERMINATION	42
3.4.1. Tail painting and rectal palpation	42
3.4.2. Blood plasma progesterone	
3.4.2.1. Elisa test	45
3.5. HERBAGE INTAKE MEASUREMENTS	47
3.5.1. Intake measurements using the n-alkane technique	47
3.5.1.1. Alkane capsules insertion	
3.5.1.2. Faeces collection	50
3.5.1.3. Intake paddocks	51
3.5.1.4. Herbage sampling for alkane profiles	52
3.5.2. Intake measurement using the pre-post grazing technique	53
3.5.2.1. Regression equations	
3.5.2.2. Pasture area allocation	55
3.6. SWARD MEASUREMENTS	58
3.6.1. Botanical composition	
3.6.2. Herbage nutritive value	
3.7. STATISTICAL ANALYSIS	59
Chapter 4 results	61
4.1. LIVEWEIGHT	
4.2. AGE AND WEIGHT AT PUBERTY	
4.3. PREGNANCY RATE	
4.4. INTAKE ESTIMATION USING N-ALKANES, FIRST MEASUREMENT	
4.4.1. Herbage measurements	
4.4.1.1 N-alkane profiles and sampling procedure	
4.4.2. Intakes	
4.5. INTAKE ESTIMATION USING N-ALKANES, SECOND MEASUREMENT	
4.5.1. Herbage measurements	
4.5.1.1. N-alkane profiles and sampling procedure	
4.5.2. Intakes	
4.6. INTAKE ESTIMATION USING PRE- AND POST-GRAZING, THIRD MEASUREMENT	
4.6.1. Herbage measurements	
4.6.2. Regression equation and intakes	
4.7. INTAKE ESTIMATION USING PRE- AND POST-GRAZING, FOURTH MEASUREMENT	
4.7.1 Herbage measurements	
4.7.2. Regression equations and intakes	

CHAPTER 5 DISCUSSION	87
5.1. PUBERTY	88
5.1.1. Age and weight at puberty	88
5.1.2. Genetic make-up and its effect on age and weight at puberty.	89
5.1.3. Environmental effects	
5.1.4. Environmental x genetic interactions	95
5.1.5. Regression curves	96
5.2. INTAKE ESTIMATIONS	96
5.2.1. Intakes using the n-alkane technique	97
5.2.1.1. Herbage sampling procedure	98
5.2.1.2. Herbage drying procedure	99
5.2.2. Intakes using the pre- and post grazing technique	100
5.2.3. Intake measurements implications	102
CHAPTER 6 CONCLUSIONS	103
REFERENCES	107

LIST OF TABLES

Table 2-1	Cumulative performance of cows managed to calf first at two or at three years of age
Table 2-2	Initial and subsequent calving group of cows, weaning weights, weaning age and average daily gain of their calves (19 years average data)7
Table 2-3	Pregnancy rate, age and weight at puberty and at breeding for heifers bred at their first or third oestrus.
Table 2-4	Least squares means for different reproductive and productive traits of Brahman heifers and cows of different mature size
Table 2-5	Puberty onset as influenced by different rates of gain and different periods of growth
Table 2-6	Comparison of known herbage intake of sheep and cattle with estimated intakes using dosed C_{32} alkane and natural C_{33} alkane as markers 35
Table 2-7	Alkane recovery values from different animal species and different diets.
Table 3-1	Number of animals by genetic line and farm of origin
Table 4-1	Liveweight increase (kg) and liveweight gain (kg day ⁻¹) during the experimental period (243 days) (means ± SEM)
Table 4-2	Age (days) and weight (kg) at puberty by genetic line (means \pm SEM). 65
Table 4-3	Age (days) and weight (kg) at puberty by farm of origin (means ± SEM).
Table 4-4	Number of heifers per line and number of heifers reaching puberty 67
Table 4-5	Equations and statistics from the regressions relating age at puberty (age) to weight at puberty (lw) by genetic line
Table 4-6	Percentage of pregnant heifers by mating cycle71
Table 4-7	Herbage botanical composition of the paddocks used in the First (18-21 June) and Second (22 –25 June) Collection Periods

Table 4-8	Herbage nutritive value from the First (18-21 June) and Second (22-25 June) Collection Periods
Table 4-9	Alkane concentration (g/kg DM) of herbage samples collected by taking a single or pooled herbage sample. Average of First (18-21 June) and Second (22-25 June) Collection Periods
Table 4-10	Liveweight (kg), herbage intake (kg DM) estimates using a single (S) and pooled (P) grass sample and the C_{32} : C_{33} alkane pairs plus theoretical (T) intake (kg DM) based on published formulae for the different selection lines of heifers (means \pm SEM).
Table 4-11	Herbage botanical composition of the paddocks used in the First (24-27 September) and Second (28 September –1 October) Collection Periods.
Table 4-12	Herbage nutritive value from the First (24-27 September) and Second (28 September –1 October) Collection Periods
Table 4-13	N-alkanes concentration in herbage obtained by a single or pooled sampling procedure and freeze- or oven-dried
Table 4-14	Liveweight (kg) and herbage intake (kg DM) estimates using the C_{32} : C_{33} alkane pairs and single (S) or pooled (P) grass sample oven-dried (O) or freeze-dried (F). Theoretical intake (kg DM) based on published formulae for the different selection lines of heifers (means \pm SEM) 78
Table 4-15	Herbage botanical composition of the lanes used in the pre- and post grazing trial
Table 4-16	Herbage nutritive value from lanes 18-A, 18-B, 18-C and 18-D 79
Table 4-17	Liveweight, liveweight gain, estimated herbage intake and theoretical herbage intake plus difference between estimated and theoretical values for the selection lines of heifers (means \pm SEM)
Table 4-18	Botanical composition of the lanes used in the pre and post- grazing trial.
Table 4-19	Herbage nutritive value from lanes 18-A, 18-B, 18-C and 18-D
Table 4-20	Regression equations, r-square and coefficient of variation (CV) for the pre- and post-grazing models.

LIST OF FIGURES

Figure 2-1	Sire expected progeny difference (EPD) for pregnancy rate (up to five mating years) related to standardised age ate first oestrus in daughters6
Figure 2-2	Schematic representation of the reproductive axis showing neuronal and endocrine interactions.
Figure 2-3	Hormonal mechanisms of the oestrous cycle
Figure 2-4	Distribution of pubertal and non-pubertal oestrus
Figure 2-5	Average concentration of alkanes in tropical and temperate grasses 33
Figure 2-6	Relationships between the estimated faecal recoveries of n-alkanes and their carbon chain lengths in sheep (a) and cows (b)
Figure 3-1	Experimental layout
Figure 3-2	Blood sampling procedure to determine time of first ovulation 44
Figure 3-3	Schematic representation of the procedures used in the first and second herbage intake measurements using n-alkanes
Figure 4-1	Monthly liveweight (kg) of the High EBV-600 day (●) Low EBV-600 day (●), High EBV-milk (●) and Low EBV-milk (■) heifer lines. Vertical bars represents standard errors of the means 62
Figure 4-2	Liveweight gain (kg/day) of the High EBV-600 day (●), Low EBV-600 day (▲), High EBV-milk (◆) and Low EBV-milk (■) genetic lines. Vertical bars are the standard errors of the means
Figure 4-3	Weight at puberty (kg) of the High EBV-600 day (HG), Low EBV-600 day (LG) and High EBV-milk (HM) by farm of origin. Vertical solid lines are the standard errors of the means
Figure 4-4	Cumulative percentage of animals attaining puberty by farm of origin. 68
Figure 4-5	Quadratic regression between liveweight (kg) and age (days) at puberty for the High EBV-600 day, Low EBV-600 day, High EBV-milk and Low EBV-milk lines.

Figure 4-6	•		regression			_	•		_	
	herbage	mass	(kg DM/ha))	•••••	• • • • • • • • • • • • • • • • • • • •				80
Figure 4-7	Simple	linear	regression	between	the	rising	plate	meter	readings	and
	herbage	mass	(kg DM/ha)	for the p	re- a	nd post	t-grazi	ng ever	nts	84