Copyright is owned by the Author of the thesis. Permission is given for a copy to be downloaded by an individual for the purpose of research and private study only. The thesis may not be reproduced elsewhere without the permission of the Author.

# THE USE OF BREATH HYDROGEN TESTING TO EVALUATE CARBOHYDRATE MALABSORPTION IN DOGS

A thesis presented in partial fulfilment of the requirements for the degree of Master of Veterinary Science in Small Animal Medicine at Massey University

> Sally Ann Bissett 1997



#### Abstract

The use of the breath hydrogen test in this thesis has focussed primarily on the study of carbohydrate assimilation in healthy dogs and in dogs with gastrointestinal disease. The gastrointestinal handling of dietary carbohydrates and the rationale, applications, and limitations of the breath hydrogen test have been reviewed. Studies were undertaken to investigate the effect of food particle size on carbohydrate digestion and the effect of dehydration on breath hydrogen concentrations in healthy dogs. In addition, breath hydrogen testing was used to assess the assimilation of four commonly used commercial carbohydrate sources in dogs with gastrointestinal disease. In each study, expired breath samples were collected at regular intervals after the ingestion of a carbohydrate test meal. The excretion of hydrogen in the breath was compared between groups, mainly by analysis of the areas under the breath hydrogen concentration versus time curves.

The reduction of food particle size was found to cause starch granule disruption and significantly decreased the amount of malassimilated rice. The assimilation of corn, however, did not appear to be altered by reducing the food particle size. Instead, an earlier rise of breath hydrogen concentrations occurred, indicating more rapid orocolic transit and/or fermentability of the smaller corn particles.

Five percent dehydration, induced by food and water deprivation, was found to significantly elevate breath hydrogen concentrations and was associated with a significantly greater number of flatus-contaminated breath samples. This increased breath hydrogen excretion associated with increased flatulence in dehydrated dogs was attributed to a greater "net" production of hydrogen within the gastrointestinal tract.

Finally, breath hydrogen concentrations were not found to vary significantly between four different extrusion cooked carbohydrate sources (wheat, potato, corn, rice). However, it could not be concluded that these carbohydrates were assimilated to a similar extent as *in vitro* fermentation results revealed marked differences in the amount of hydrogen produced per gram of carbohydrate fermented at different fermentation times. Individual dogs with gastrointestinal disease appeared to differ in their ability to assimilate the four different carbohydrate sources. In addition, dogs with exocrine pancreatic insufficiency were found to malabsorb significantly more carbohydrate than the dogs with mild inflammatory bowel disease.

In conclusion, food processing to reduce carbohydrate particle size appears to increase the assimilation of rice. Methods to reduce the particle size of rice should be considered in the formulation of veterinary therapeutic diets for the management of diarrhoea. Mild dehydration appears to increase breath hydrogen excretion. This suggests that breath hydrogen tests should not be performed on animals that are suboptimally hydrated until their hydration deficits have been restored. The marked individual variation of carbohydrate assimilation noted in dogs with gastrointestinal disease, suggests that clinicians should consider altering the carbohydrate source offered to dogs with intractable malassimilation, with the aim of finding the carbohydrate best tolerated. Finally, direct comparison of breath hydrogen concentrations should not be used to compare the digestibility of different complex carbohydrates unless an accurate means is available of comparing the amount of hydrogen produced per gram of substrate fermented *in vivo*.

#### Acknowledgements

The lams Company, Lewisburg, Ohio, generously funded this Masters project, contributed the carbohydrate sources, and performed the *in vitro* fermentation work used in the study comparing the assimilation of wheat, corn, potato and rice in dogs with diarrhoea (Chapter 5). A special thanks to Greg Sunvold for taking the time to review manuscripts, for communicating from afar, and for organising the *in vitro* fermentation data.

The statistical analysis of the research was kindly performed by Charles Lawoko (Chapter 3) and Steve Haslett (Chapters 4 and 5). Their contribution to the studies and their patient explanations were greatly appreciated.

I would also like to thank the nursing staff, students and colleagues who assisted with the collection of breath samples and the care of dogs used in this Masters project. In particular, Pauline Gordon, Jill Hogan, Angus Fordham, Robin Gear, Alison Meyer, Kate Carthew, Nicola Smith and Linda Macknight were of great help in this regard, and their efforts were thoroughly appreciated. Thank you also to Alan Anderson, Steve Lees and Anne Tunnicliffe for organising the availability and transport of colony dogs, and to the clients who generously parted with their dogs for several days in order for breath hydrogen tests to be performed.

Most importantly, I would like to express my gratitude to my supervisors. Grant Guilford and Boyd Jones were integral to the formation of this thesis and I am indebted to them for their guidance, support, and for providing the opportunity to take on this task (and others to come). Grant has been particularly generous with his time, advice and humour for which I whole-heartedly thank him.

## **Table of Contents**

Abstract	ii
Acknowledgmen	nts iv
Table of Conter	v
List of Tables	vii
List of Figures	
Chapter 1.	A Review of Gastrointestinal Handling of Dietary Carbohydrates 1-27
	-Introduction
	-Carbohydrates in the Diet
	-Carbohydrates in the Small Bowel: Digestion and Absorption 5
	-Carbohydrates in the Large Bowel: Fermentation 13
	-Carbohydrate Intolerance 21
	-References
Chapter 2.	A Review of the Breath Hydrogen Test
	-Introduction
	-Breath Hydrogen as a Measure of Intestinal Gas Formation 28
	-Applications of the Breath Hydrogen Test
	-Variables and Limitations of Breath Hydrogen Measurements 54
	-References
Chapter 3.	Effect of Food Particle Size on Carbohydrate Assimilation Assessed by
	Breath Hydrogen Testing in Dogs
	-Introduction
	-Materials and methods
	-Results 82
	-Discussion 88
	-References

Chapter 4.	Effect of 5% D	ehydration on Breath Hydrogen Concentrations in
	Dogs	
	-Introduction	
	-Materials and	methods
	-Results	
	-Discussion	
	-References	
Chapter 5.	A Comparison	of Wheat, Potato, Corn and Rice Assimilation in Dogs with
	Diarrhoea using	g Breath Hydrogen Tests
	-Introduction	
	-Materials and	methods
	-Results	
	-Discussion	
	-References	
Chapter 6.	Conclusions	

## List of Tables

Table 1.1	Main food carbohydrates 5
Table 1.2	Major intestinal brush border carbohydrases in mammals 9
Table 2.1	Examples of the methodology and criteria used for investigating SIBO by
	breath $H_2$ analysis 41
Table 2.2	Examples of the methodology and criteria used for estimating OCTT by
	breath H <sub>2</sub> analysis
Table 3.1	Protocol used to obtain expired breath samples by the anaesthetic face
	mask technique
Table 3.2	Mean $\pm$ SD breath H <sub>2</sub> data of 10 dogs after the ingestion of a whole rice or
	blended rice meal
Table 3.3	Mean $\pm$ SD breath H <sub>2</sub> data of 10 dogs after the ingestion of a chopped corn
	or blended corn meal
Table 4.1	Clinical and laboratory parameters of dehydrated and non-dehydrated
	dogs
Table 5.1	Nutritional analysis of the wheat, potato, corn and rice flours 107
Table 5.2	Composition of medium used to culture faecal microflora 108
Table 5.3	Mean in vitro $H_2$ and $CH_4$ production of wheat, potato, corn and rice
	substrates

## List of Figures

Figure 1.1	An example of the molecular structure and $\alpha$ 1,6 branching point of the
	amylopectin chain 4
Figure 1.2	Alpha-amylase hydrolysis of amylose and amylopectin chains
Figure 1.3	Active transport of glucose (or galactose) across the brush border membrane
	by the Na <sup>+</sup> -glucose cotransporter 12
Figure 1.4	Overview of carbohydrate degradation and fermentation in the large
	bowel
Figure 3.1	The collection of expired breath by the face mask technique 80
Figure 3.2	Mean $\pm$ SEM breath H <sub>2</sub> concentrations of 10 dogs after the ingestion of a
	whole rice or blended rice meal
Figure 3.3	Photomicrographs of cooked rice starch from two different rice meals 84
Figure 3.4	Mean $\pm$ SEM breath H <sub>2</sub> concentrations of 10 dogs after the ingestion of a
	chopped corn or blended corn meal
Figure 3.5	Photomicrographs of cooked corn starch from two different corn meals 87
Figure 4.1	Mean $\pm$ SEM breath H <sub>2</sub> concentrations of 10 dehydrated and non-dehydrated
	dogs after the ingestion of a wheat-containing test meal
Figure 4.2	AUC of 10 dogs tested when dehydrated and non-dehydrated over 18 hours
	after the ingestion of a wheat-containing test meal
Figure 5.1	Mean $\pm$ SEM breath H <sub>2</sub> concentrations of 12 dogs after the ingestion of
	wheat, potato, corn and rice meals
Figure 5.2	Mean $\pm$ SEM breath H <sub>2</sub> concentrations of the dogs with exocrine pancreatic
	insufficiency (EPI) and inflammatory bowel disease (IBD) 113
Figure 5.3	AUC data of 12 dogs over 18 hours after the ingestion of each of the four
	different carbohydrate

1

.