

Copyright is owned by the Author of the thesis. Permission is given for a copy to be downloaded by an individual for the purpose of research and private study only. The thesis may not be reproduced elsewhere without the permission of the Author.

***Uterine involution in Holstein-Friesian cows  
genetically selected for high or low mature  
body weights***

A thesis presented in partial fulfilment of the requirements for the  
Master of Veterinary Science  
At Massey University  
Palmerston North, New Zealand

**Fadi Zaki Daoud**  
**2008**

## Abstract

The main objective in dairy herds is to produce milk as economically and efficiently as possible. To achieve maximum production, cows must calve regularly; hence the fertility of the herd affects the productivity of the farm dramatically. High fertility requires early uterine involution, early onset of oestrous cycles postpartum and optimal oestrus detection and conception rates (Pelssier 1976). The modern high producing, lactating dairy cow in North America is subfertile (Thatcher et al., 2006) as managed under current production systems. However, whether such subfertility occurs in high-producing cows under our pastoral system in New Zealand is not fully known. However, recent movement of genetic material across the globe, in the form of semen or frozen embryos might suggest that New Zealand herds could be drifting in a similar direction in subfertility.

The main objective of the present study was to investigate the effect of genetics on uterine involution in the context of a pastoral system by comparing two strains Holstein/Friesian cows that had been genetically selected for high (H) or low (L) mature body weight. Involution was studied through the following measurements: (i) cervical diameter, as assessed by measurement *per rectum*, (ii) plasma concentrations of the prostaglandin  $F_{2\alpha}$  metabolite, 15-keto-13,14-dihydro-prostaglandin  $F_{2\alpha}$  (PGFM), (iii) urinary concentrations of the collagen breakdown product pyridinoline and (iv) bacteriology of the cervical canal.

From the results, it was concluded that, whilst H cows exhibited similar physical involution characteristics to those of L cows ( $P>0.5$ ), they had higher levels of post-calving bacterial contamination ( $P<0.05$ ). On Days 4, 7 and 10 post partum, the anaerobic bacterial load was significantly ( $P<0.05$ ) greater in H than L cows. On Day 10, both strains peaked with mean

anaerobic bacterial counts of  $23.4 \times 10^5$  cfu and  $0.41 \times 10^5$  in H and L cows, respectively. Similarly, on Days 7 and 10, the total bacterial load (aerobic plus anaerobic) was also greater ( $P < 0.05$ ) in H than L cows. On Day 7 mean total bacterial counts were  $2.27 \times 10^6$  cfu in H cows. Peak numbers of bacteria in H cows were attained on Day 10 ( $3.39 \times 10^6$  cfu). Values in L cows were maximal on Day 7 ( $1.18 \times 10^6$  cfu).

PGFM Concentrations in L cows were significantly ( $P < 0.05$ ) higher than in H cows on Days 17, 28 and 35. Although total pyridoline and deoxypyridoline /creatinine ratio concentrations differed between strains and times, the strain.time interaction term was not significant. However, on Day 11, values in L cows tended ( $P = 0.07$ ) to be higher than in H cows.

Simple correlations established that different parameters e.g cervical diameter; PGFM, PYD and bacterial contamination are highly correlated and moving simultaneously together and are not independent of each other. Accordingly, the relationships between actual and predicted intervals between calving and; first oestrus, first insemination and conception, were calculated based on principle component and partial regression analyses of parameters of uterine involution. Indices calculated from these parameters as predictors of reproductive outcomes, were significantly correlated with intervals between calving and first oestrus ( $P < 0.05$ ) and calving and first insemination ( $P < 0.01$ ), but not significant with conception rate ( $P > 0.5$ ).

Taken together, these data show that uterine involution is impaired in H compared to L strain cows, inasmuch as there is a greater degree of bacterial contamination and a more sluggish pattern of PFGM secretion in H cows. Collagen remodelling may also be

attenuated in H than L cows, although differences between strains only tended towards significance.

More work need to be carried out to further investigate the reason for these differences whether it is genetically based or higher production is negatively affecting the above mentioned traits at a cellular level.

## List of figures

Figure number	page number
<b>Figure 2.1</b> Differences in elution time between (a) standard, and an example of a (b) sample where PYD/DPD eluted at a different time with multiple peaks before and after the expected elution time of PYD/DPD.	49
<b>Figure 3.1</b> Changes of cervical diameter with time after calving. Differences between days were significant ( $P<0.01$ ), but there were no differences between strains	56
<b>Figure 3.2</b> Mean ( $\pm$ sem) numbers of colony forming units of organisms cultured from cervical swabs collected from H and L strain cows over the post partum period (a) Aerobic bacteria(b) Anaerobic bacteria.	59
<b>Figure 3.3</b> Mean ( $\pm$ s.e.m.) total colony forming units of aerobic and anaerobic bacteria cultured from cervical swabs of H and L strain cows over the post partum period.	60
<b>Figure 3.4</b> Proportion of H and L strain cows from which positive microbiological cultures were obtained from cervical swabs collected over the post partum period. Differences between days, but not strains, were significant ( $P<0.05$ ).	61
<b>Figure 3.5</b> Mean ( $\pm$ sem) peripheral PGFM concentrations in H and L strain cows over the postpartum period	63
<b>Figure 3.6</b> Relationship between plasma PGFM concentrations and cervical diameter in H and L cows during the post partum period. Data are for individual measurement from each cow at each time point. The correlation is significant ( $P<0.001$ ).	64
<b>Figure 3.7</b> Relationship between numbers of (a) aerobic bacteria, (b) anaerobic bacteria and (c) total bacterial count and plasma concentrations of PGFM in H and L cows over the post partum period.	65
<b>Figure 3.8</b> Urinary contents of pyridoline and deoxypyridoline, as expressed as the ratio of total pyridoline and deoxypyridoline /creatinine (nmol pyridoline + deoxypyridoline /mmol creatinine) in H and L strain cows over the post partum period. Differences between time points, but	

not between strains, were significantly different from each other ( $P<0.01$ ) and approached significance ( $0.05<P<0.10$ ) where indicated (+). 67

**Figure 3.9** Relationship between total pyridoline and deoxypyridoline /creatinine ratio and cervical diameter in H and L cows over the post partum period. 67

**Figure 3.10** Relationship between actual and predicted intervals between calving and first oestrus, based on partial regression analysis of parameters of uterine involution. The relationship ( $y = 0.5236x + 22.021$ ;  $R^2 = 0.4149$ ) was significant ( $P<0.05$ ). 69

**Figure 3.11** Relationship between actual and predicted intervals between calving and first insemination, based on partial regression analysis of parameters of uterine involution. The relationship ( $y = 0.612x + 23.839$ ;  $R^2 = 0.6009$ ) was significant ( $P<0.01$ ). 69

**Figure 3.12** Relationship between actual and predicted intervals between calving and conception, based on partial regression analysis of parameters of uterine involution. The relationship ( $y = 0.1618x + 74.688$ ;  $R^2 = 0.0822$ ) was not significant. 71

**List of tables**

<b>Table number</b>	<b>page number</b>
<b>Table 2.1</b> Reagents (μl) used in PGFM radioimmunoassay	45
<b>Table 3.1</b> Reproductive outcomes	54
<b>Table 3.2</b> Relationship between uterine involution and interval between calving and first oestrus	68
<b>Table 3.3</b> Relationship between uterine involution and interval between calving and first insemination	70
<b>Table 3.4</b> Relationship between uterine involution and interval between calving and conception	71



**Table of contents**

**Abstract.....I**

**List of figures.....IV**

**List of tables..... VI**

**Table of contents.....VII**

  

***CHAPTER I: Literature Review..... 1***

  

**1.1 Introduction.....1**

**1.2 Uterine involution.....3**

  

**1.3 Management of New Zealand cows.....3**

**1.4 Post partum period.....5**

**1.5 Ovarian activity.....7**

**1.6 Endocrinology of the post partum period.....7**

**1.6.1 Role of Hypothalamo-Pituitary Axis.....8**

**1.6.2 GnRH and LH secretion.....9**

**1.6.2.1 During regular oestrus cycles.....9**

**1.6.2.2 During the postpartum period.....10**

**1.6.3 GnRH and FSH secretion.....11**

**1.6.3.1 During regular oestrous cycles.....11**

**1.6.3.2 FSH during the postpartum period.....12**

**1.6.4 Progesterone.....14**

**1.6.5 Oestrogen.....15**

**1.6.6 Prostaglandin F<sub>2α</sub>.....16**

**1.6.7 Prolactin.....19**

**1.7 Bacteriology of the post partum uterus.....20**

**1.8 Factors influencing uterine involution.....24**

**1.8.1 Abnormalities of parturition .....24**

1.8.2	Infection .....	26
1.8.3	Milk production .....	27
1.8.4	Breed differences .....	28
1.8.5	Parity .....	30
1.8.6	Season .....	30
1.8.7	Nutrition .....	32
1.9	Collagen remodelling.....	34
1.9.1	Structure of collagen.....	35
1.9.2	Collagen breakdown.....	35
1.9.3	Detection of collagen.....	36
1.9.4	Urinary collagen degradation products .....	36
1.10	Summary .....	37
 <i>CHAPTER II: Materials and Methods.....</i>		<i>40</i>
2.1	Introduction.....	40
2.2	Animals and Management.....	41
2.3	Grazing and pasture management.....	41
2.4	General reproductive management.....	42
2.5	Sampling regimen.....	42
2.6	Radioimmunoassay of PGFM concentrations.....	44
2.7	Microbiology.....	46
2.8	Measurement of collagen breakdown products (pyridoline and deoxypyridolin).....	46
2.8.1	HPLC.....	47
2.8.2	ELISA.....	49
2.9	Analysis of data.....	51
2.10	Ethics Approval.....	52
 <i>CHAPTER III: Results.....</i>		<i>53</i>
3.1	Cervical diameter.....	54
3.2	Microbiology.....	56

3.2.1	<i>Aerobic bacterial contamination.....</i>	56
3.2.2	<i>Anaerobic bacterial contamination.....</i>	56
3.2.3	<i>Total bacterial contamination.....</i>	56
3.3	<b>Prostaglandin F<sub>2α</sub> metabolite (PGFM).....</b>	61
3.4	<b>Pyridoline and deoxypyridoline.....</b>	65
3.5	<b>Relationships between parameters of uterine involution and fertility outcomes.....</b>	67
 <b><i>CHAPTER IV: Discussion.....</i></b>		72
4.1	<b>Introduction.....</b>	72
4.2	<b>Cervical Diameter.....</b>	73
4.3	<b>Bacterial contamination.....</b>	74
4.4	<b>Prostaglandin F<sub>2α</sub>.....</b>	80
4.5	<b>Collagen breakdown products .....</b>	86
4.6	<b>General conclusion.....</b>	89
<b>Reference list .....</b>		91