

Copyright is owned by the Author of the thesis. Permission is given for a copy to be downloaded by an individual for the purpose of research and private study only. The thesis may not be reproduced elsewhere without the permission of the Author.

Benthic communities of the Whanganui River catchment: the effects of land use and geology



**A thesis presented in partial fulfilment of the requirements
for the degree of Master of Science in Ecology
at Massey University,
Palmerston North,
New Zealand**

**Jonathan Vaughan Horrox
1998**

TABLE OF CONTENTS

ABSTRACT:	1
 CHAPTER 1:	
General introduction.	2
 CHAPTER 2:	
Effects of land use and geology on lotic macroinvertebrate communities in the Whanganui River.	7
 CHAPTER 3:	
Longitudinal changes in macroinvertebrate community structure along a large New Zealand river.	42
 CHAPTER 4:	
Morphological characteristics of the New Zealand freshwater mussel <i>Hyridella menziesi</i> in the Whanganui River and its catchment.	63
 ACKNOWLEDGEMENTS:	83

ABSTRACT

ABSTRACT

The response of macroinvertebrate communities and freshwater mussels to variation associated with land use and geology was investigated in small headwaters of the Whanganui River catchment in the North Island of New Zealand. Conductivity and water clarity were higher in streams with soft tertiary/quaternary sedimentary geology, independent of land use. These soft geology catchments, when forested, had distinctive community structure often involving high relative abundances of Ephemeroptera. Pastoral agriculture resulted in a lower diversity and abundance of pollution sensitive taxa. Impacts of pastoral land use were accentuated by soft sedimentary geology culminating in low diversity and abundance in pasture streams with soft geology. Multivariate analysis showed that community structure varied significantly depending on the combination of land use and geology type present, with land use as the most significant factor. However, variations in geology may mask the effects of land use between catchments.

In the main Whanganui River, taxonomic diversity and numbers of pollution sensitive taxa decreased downstream. This was correlated with reduced periphyton biomass and increased suspenoids. Estimates of macroinvertebrate community structure differed between artificial substrate and kick samples collected from the same sites.

Shell morphology of the freshwater mussel *Hyridella menziesi* was not correlated with water chemistry. One site near the northern boundary of the Whanganui River catchment contained mussels with distinct shell morphology. A possible explanation involving stream capture by tectonic movements is considered. Shell erosion was correlated with channel width suggesting shell erosion is greater in larger waterways. Lack of demarcation in shell growth annuli meant accurate estimates of mussel age were not possible. Poor demarcation is likely to result from non seasonal patterns of environmental variation.

Chapter One

General introduction

INTRODUCTION

The character of streams and rivers reflects the integration of physical and biological processes occurring in the catchment (Johnson and Gage, 1997). Most of the Whanganui River catchment (Fig. 1.1) was once heavily forested but much of this land has been cleared for pastoral agriculture. Some of these areas have been retired allowing the native vegetation to re-grow. The land is young in geological terms consisting predominantly of Pliocene-Pleistocene marine sediments or volcanics which are younger than most of the North Island. Various unconsolidated rock types combined with low gradient provide suitable soft bottom habitats for the New Zealand freshwater mussel *Hyridella menziesi*. Many ecological features of this species are well documented in lakes but little is known of the populations present in the Whanganui River, or other lotic habitats.

The landscape influences its water bodies through multiple pathways and mechanisms, operating at different spatial scales (Allan and Johnson, 1997). Large scale factors influencing stream ecosystem function such as geology and land use (Winterbourn, 1986) have an important influence on the physicochemical characteristics of streams (Richards *et al.*, 1997). Macroinvertebrate community structure is closely linked to these characteristics making them appropriate indicators for investigating environmental change (Stark, 1993). Changes in land use practices such as those related to agricultural development are predicted to alter habitat characteristics, invertebrate community composition and water quality (Lenat, 1984; Corkum, 1990; Quinn and Hickey, 1990b). Catchment geology also affects the structure of aquatic ecosystems over a range of spatial scales through its influence on water chemistry (Maasdam and Smith, 1994), catchment vegetation, flow variability and substrate characteristics (Cummins *et al.*, 1984; Richards *et al.*, 1997). Catchments with a combination of pastoral agriculture and soft sedimentary geology can have distinctive community structure such as low abundance and diversity (Quinn and Hickey, 1990a; Davies-Colley and Stroud, 1995). The substrate of many streams in the Whanganui River catchment are dominated by erodable Tertiary aged sedimentary rock. Little is known about the basic ecology of invertebrate communities in these mudstone streams, and the influence of land use upon them.

Hyridella menziesi is abundant throughout many lakes in both the North Island and South Island. However, little is known about this species in the Whanganui River catchment. Anecdotal evidence concerning the distribution and abundance of *Hyridella menziesi* in the Whanganui River catchment suggest that the species has declined since the turn of the century. Clearly, more information concerning their response to environmental variation will be beneficial to understanding reasons for their decline.

Increased knowledge of ecological conditions that occur in the main river and its tributaries will provide greater understanding of environmental and ecological characteristics of the Whanganui River. Chapter Two investigates the effects of pastoral land use and geology type on macroinvertebrate communities in headwater streams, in particular those from catchments with soft sedimentary geology. Longitudinal changes in macroinvertebrate communities along the main Whanganui River in response to tributary inputs are explored in Chapter Three. The use of both artificial substrate and kick sampling were used to assess these changes and their respective abilities to do so were compared. In Chapter Four, morphological and spatial characteristics of the New Zealand freshwater mussel *Hyridella menziesi* are investigated in relation to physicochemistry and habitat from six sites in the Whanganui river and its tributaries.



Figure 1.1. Map showing the position of the Whanganui River catchment in the North Island of New Zealand.

REFERENCES

- Allan, D.J. and Johnson, L.B. 1997. Catchment style analysis of aquatic ecosystems. Freshwater Biology **37**: 107-111.
- Corkum, L.D. 1990. Intrabiome distributional patterns of lotic macroinvertebrate assemblages. Canadian Journal of Fisheries and Aquatic Sciences **47**: 2147-2157.
- Cummins, K.W., Minshall, G.W., Sedall, J. R., Cushing, C. E. and Petersen, R. C. 1984. Stream Ecosystem Theory. Verhandlungen der Internationalen Vereinigung für theoretisch und angewandte Limnologie **22**: 1818-1827.
- Davies-Colley, R.J. and Stroud, M.J. 1995. Water quality degradation by pastoral agriculture in the Whanganui River catchment. Hamilton: National Institute of Water and Atmospheric Research. Consultancy Report DoC050/1.
- Johnson, L.B. and Gage, S.H. 1997. Landscape approaches to the analysis of aquatic ecosystems. Freshwater Biology **37**: 113-132.
- Lenat, D.R. 1984. Agriculture and stream water quality: a biological evaluation of erosion control practices. Environmental Management **8**: 333-344.
- Maasdam, R. and Smith, D.G. 1994. New Zealand's National River Water Quality Network 2. Relationships between physico-chemical data and environmental factors. New Zealand Journal of Marine and Freshwater Research **28**: 37-54.
- Quinn, J.M. and Hickey, C.W. 1990a. Characterisation and classification of benthic invertebrate communities in 88 New Zealand rivers in relation to environmental factors. New Zealand Journal of Marine and Freshwater Research **24**: 387-409.
- Quinn, J.M. and Hickey, C.W. 1990b. Magnitude of effects of substrate particle size, recent flooding and catchment development on benthic invertebrates in 88 New Zealand rivers. New Zealand Journal of Marine and Freshwater Research **24**: 411-427.

- Richards, C., Haro, R.J., Johnson, L.B. and Host, G.E. 1997. Catchment and reach-scale properties as indicators of macroinvertebrate species traits. Freshwater Biology **37**: 219-230.
- Stark, J.D. 1993. Performance of the Macroinvertebrate Community Index: effects of sampling method, sample replication, water depth, current velocity, and substratum on index values. New Zealand Journal of Marine and Freshwater Research **21**: 463-478.
- Winterbourn, M.J. 1986. Effects of land development on benthic stream communities. New Zealand Agricultural Science **20**: 115-118.

Chapter Two

Effects of land use and geology on lotic macroinvertebrate communities in the Whanganui River

ABSTRACT

Benthic macroinvertebrate communities were sampled during 1996 and 1997 in streams from the Whanganui River catchment varying in geology and land use. Mudstone streams had higher conductivity and lower water clarity, independent of land use. The mayfly species *Coloburiscus humeralis*, *Zephlebia dentata* and *Deleatidium* spp. as well as *Pycnocentria funerea*, a caddisfly were characteristically abundant in forested streams with soft sedimentary geology. Pastoral agriculture resulted in lower diversity and abundance of pollution sensitive taxa. Impacts of pastoral land use were accentuated by soft sedimentary geology culminating in low diversity and abundance in mudstone pasture streams, with greater dominance of the gastropod *Potamopyrgus antipodarium*. Multivariate analysis showed that community structure varied significantly depending on the combination of land use and geology type present.

Keywords: benthic macroinvertebrate communities, mudstone, geology, pastoral agriculture, native forest, water clarity, relative abundance, diversity.

INTRODUCTION

Human alteration of landscapes has widespread implications for the well being of aquatic ecosystems (Naiman *et al.*, 1995). Land use such as pastoral farming, inevitably has a major effect on the physicochemical characteristics of streams (Richards *et al.*, 1997). As well as land use characteristics, the dominant surficial geology type can account for a large proportion of the variation in macroinvertebrate distribution patterns (Richards *et al.*, 1996).

Changes in land use practices such as those related to agricultural development are predicted to alter habitat characteristics, invertebrate community composition and water quality (Lenat, 1984; Corkum, 1990; Quinn and Hickey, 1990a). Changes in invertebrate community structure in response to land development has been well documented in both New Zealand (e.g., Allan, 1959; Winterbourn, 1986; Quinn and Hickey, 1990b; Smith *et al.*, 1993; Quinn *et al.*, 1997; Townsend *et al.*, 1997) and overseas (Dance and Hynes, 1980; Reed *et al.*, 1994; Richard and Host, 1994; Ventura and Harper, 1996).

Changes in community structure resulting from agricultural development include decreases in diversity and taxonomic richness, increases in invertebrate biomass, and reductions in the number of Ephemeroptera, Trichoptera and Plecoptera. A dominant feature of changing land use is the alteration or loss of riparian vegetation which then results in increases in light, nutrient and sediment levels, and alterations of flow and temperature regimes (Cowie, 1985; Winterbourn, 1986; Dons, 1987; Fahey and Rowe, 1992; Williamson *et al.*, 1992; Prat and Ward, 1994; Quinn *et al.*, 1994). Catchment vegetation types such as pine and native forest often have higher amounts of particulate organic matter than that of pasture (e.g., Friberg *et al.*, 1997). Increases in CPOM can alter community energetics and subsequently invertebrate functional feeding groups allowing for greater abundance of facultative shredders (e.g., Harding and Winterbourn, 1995).

The underlying geology of a catchment plays a key role in structuring aquatic ecosystems at both macro and micro habitat level through its influence on water quality, catchment vegetation, flow variability and substrate characteristics. Geology may also have a major influence on aquatic ecosystems through the regulation of dissolved ion input (Cummins *et al.*, 1984; Lay and Ward, 1987; Close and Davies-Colley, 1990). Areas containing volcanic rocks and tephra in New Zealand often have higher phosphorus concentrations (Currie and Gilliland, 1980; Timperley, 1983), granite and limestone can increase nitrate levels (Lay and Ward, 1987), and bicarbonate ions created from the dissolution of limestone bedrock can increase alkalinity (Stumm and Morgan, 1981). Geology may also influence the composition of catchment vegetation and the rate of organic matter turn-over, both of which can influence stream ecosystem function (Miller, 1968; Lay and Ward, 1987). Large scale geological characteristics also influence stream hydrology and thermal characteristics (Wiley *et al.*, 1997). For example streams supplied predominantly by surficial runoff rather than ground water have more variable flow and temperature regimes (Haro and Wiley, 1992; Richards *et al.*, 1997). Flow variation may affect aquatic communities directly (Statzner *et al.*, 1988; Quinn and Hickey, 1990b; Poff and Allan, 1995), or indirectly in combination with gradient, bed stability and the size distribution of substrate (Brussven, 1984; Winterbourn, 1986; Jowett and Richardson, 1990).

It is evident from the above studies that geology and land use both play important roles in shaping invertebrate community structure. Despite this, the focus of research concerning the effects of land use on water quality and invertebrate community structure in New Zealand (Quinn *et al.*, 1992; Harding and Winterbourn, 1995) has been focused primarily in cobble bottom streams with hard geology. The substrate of many streams in the Whanganui River catchment and others in the central North Island are dominated by erodable Tertiary aged sedimentary rock. Little is known about the basic ecology of invertebrate communities in these mudstone streams. These types of catchments however are among those most highly influenced by agricultural activity (Maasdam and Smith, 1994). To successfully manage these streams it is necessary to know how impacts of land use on water quality and invertebrate community structure may differ with catchment geology. In this study I survey invertebrate communities and associated measures of water quality in 48 streams in the Whanganui River catchment that exhibit different geological characteristics, and compare the influence of land use on those communities.

STUDY SITES

All study sites were third to fifth order streams draining into the Whanganui River in the central North Island of New Zealand. Tertiary sandstone, mudstone, and siltstone are the most common geology types in the catchment. Streams around Mount Ruapehu flow through bedrock of volcanic origin composed of Andesites. In the northern catchment, streams flow through Jurassic sandstone and siltstone with Ignimbrite substrates in many headwaters. Near the city of Wanganui, geology is dominated by less stable Quaternary sands and silts. Sites were chosen based on land use and surface geology. Streams were therefore grouped as having either harder Quaternary Andesite and Jurassic substrates or softer mudstone substrates of Tertiary/Quaternary age, and land use types as predominantly pasture or native vegetation. Most of the agricultural activity consisted of lower intensity sheep/beef grazing. Native vegetation consisted of mixed broadleaf/podocarp forest with areas of beech and native grassland in some of the catchments situated on the volcanic plateau of Mount Ruapehu. Aborted attempts to farm many marginal areas before and during the 1940's meant that some of the lower altitude forests are of relatively young age. Stream geology was determined using a combination of topographical maps, geological maps, and the substrate type found when sampling. Land use/geology classes will be referred to as mudstone pasture or forest for

substrates formed by Tertiary/Quaternary sediments and hardstone pasture or forest for Andesitic/Jurassic substrates. Study sites ranged in altitude between < 20 m and 840 m a.s.l.

METHODS

Sampling protocol

Sampling was conducted on two occasions. Between March 25 and April 28, 1996, 33 sites differing in geology and land use (Fig. 2.1) and February, 1997, 30 sites all in catchments of sand/silt/mudstone geology's were sampled (Fig. 2.2). Fifteen of these sites had also been sampled during the 1996 sampling period.

On both occasions sampling of macroinvertebrates was conducted by collecting four replicate 250 μm mesh, 0.11m² surber samples which were preserved in 70% alcohol. Samples were sieved through a 250 μm Endecott sieve and animals identified and enumerated with a 40 \times microscope using available keys (McFarlane, 1951; Winterbourn, 1973; Chapman and Lewis, 1976; Cowley, 1978; Winterbourn and Gregson, 1989; Towns and Peters, 1996). If taxa could not be identified to species they were assigned to apparent morphospecies.

Concurrent with each invertebrate collection a range of physicochemical parameters were measured (Table 2.1 and 2.2). Conductivity and temperature were measured using an Orion 122 conductivity meter and pH with an Orion 250A pH meter. A black disk was used to measure water clarity (Davies-Colley, 1988). Velocity was measured using a Kempton A.OTT current meter. Between 3 and 10 measures of depth and velocity were made in a transect at a right angle across the stream. Large rivers or those with greater channel variability required more measurements. Substrate size composition was assessed visually as a proportion and converted to a substrate index following Jowett *et al.* (1991). Stability was assessed using the bottom component of the Pfankuch index (Pfankuch, 1975). Channel slope was determined as the distance between three 20 m contours on a 1:50 000 topographical map and converted to degrees. The percentage of forest cover was calculated by tracing onto acetate sheets the catchment from topographical maps. Areas of forest were then cut, weighed and converting to a proportion of the total weight.

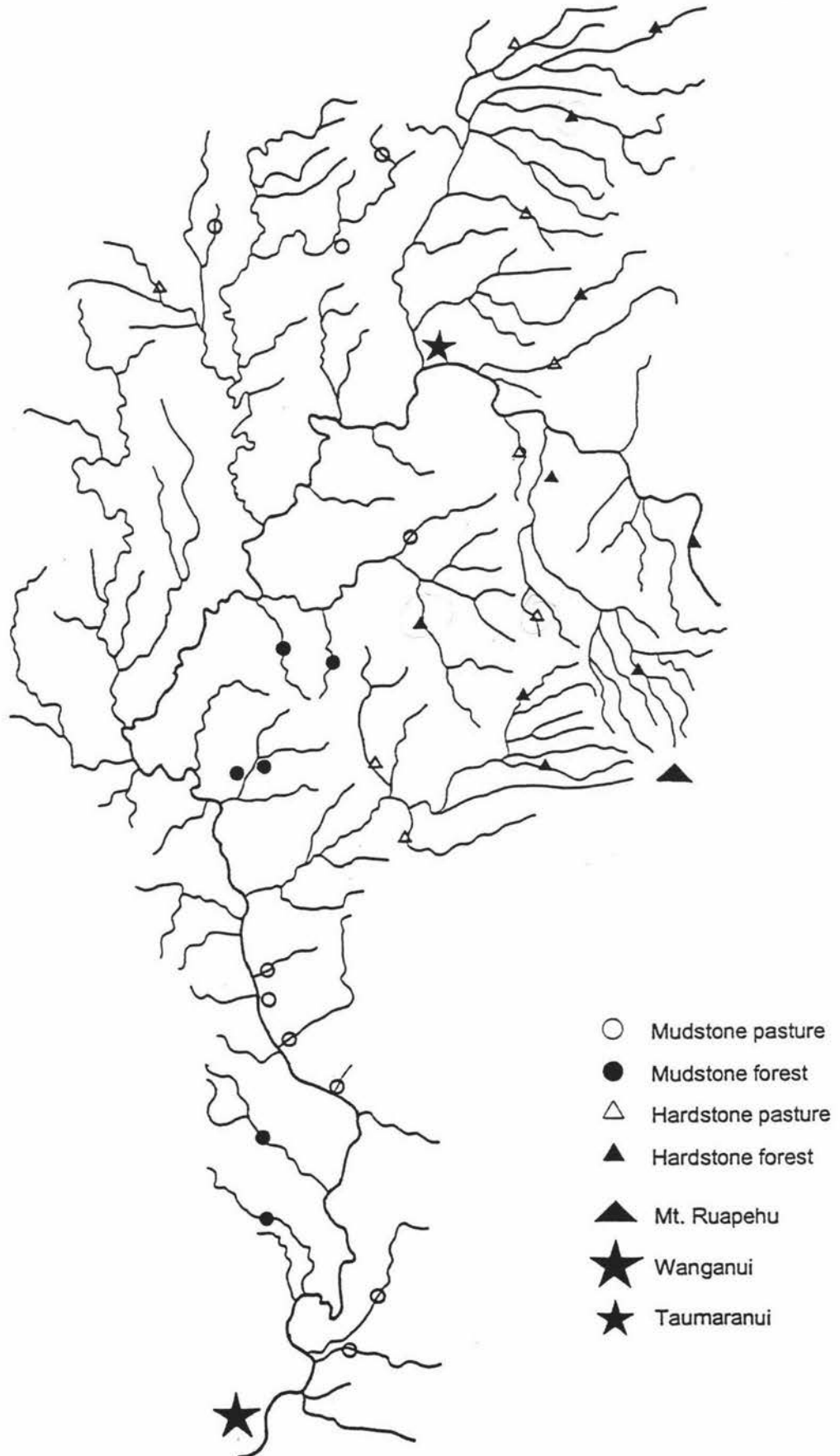


Figure 2.1. Location of the 33 streams sampled between March 25 and April 28, 1996, showing the geology/land use classes and major population centres.

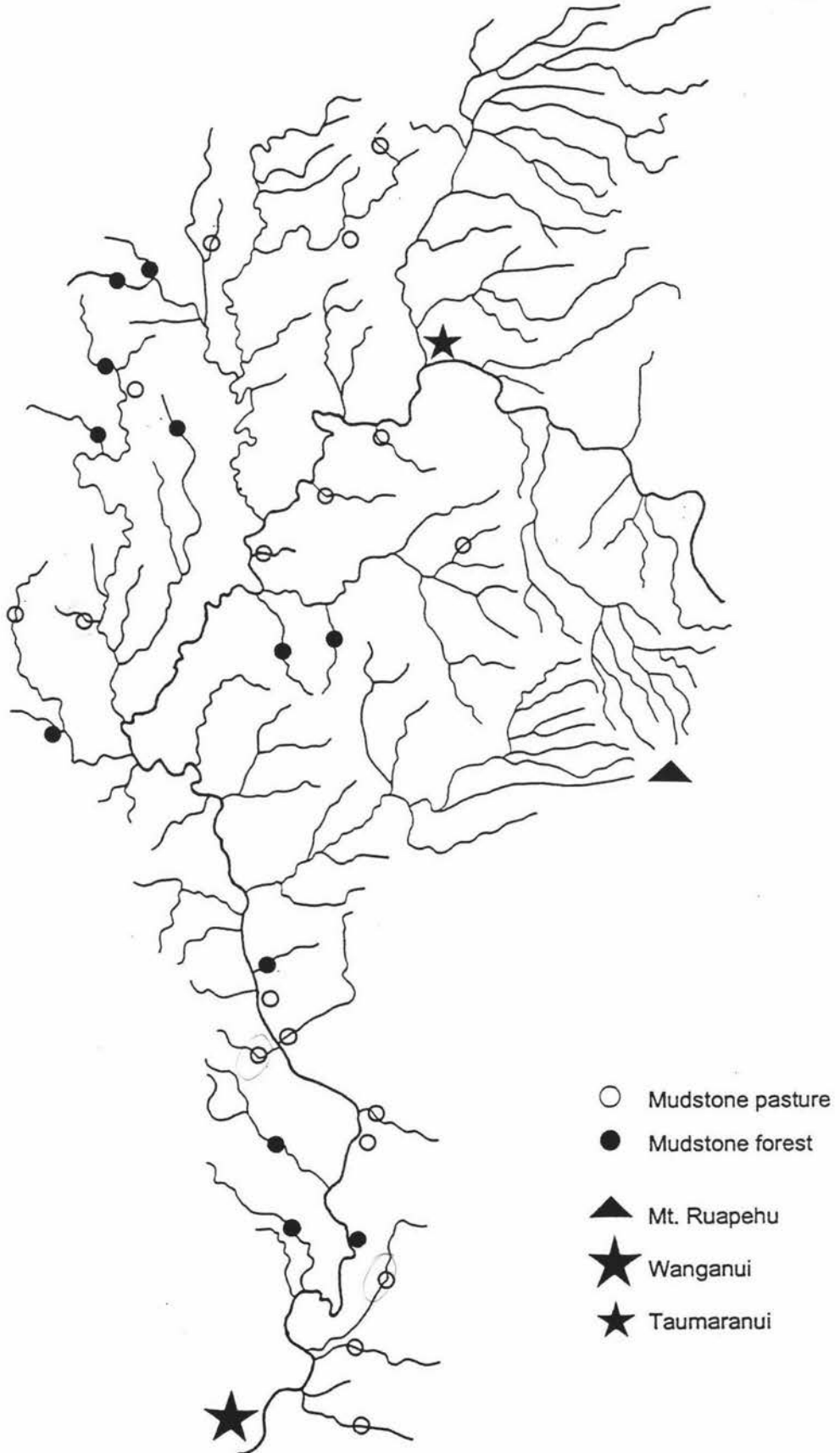


Figure 2.2. Location of the 30 mudstone streams sampled February, 1997, showing the land use classes and major population centres.

Table 2.1. Physical and chemical characteristics recorded from 33 streams in the Whanganui River catchment, between March and April, 1996, which have mudstone or hardstone substrates and drain pasture or indigenous forest.

Sites	Width (m)	Mean depth (cm)	Mean velocity (cm s ⁻¹)	Slope (degrees)	Stability	Conductivity (μScm^{-2})	Black disk (m)	pH	Substrate size index	Altitude (metres)	Map reference 1:50 000
<i>Mudstone Pasture</i>											
Kawautahi	6.3	20	52	0.41	31	93.3	0.53	6.87	0.049	240	S19 055390
Makirikiri	2	17	78	0.28	45	299	0.6	7.51	0.029	40	S22 934474
Motuaruhe	7.1	16	59	0.37	31	200	0.23	7.52	0.059	40	S21 956754
Ohura	5	34	34	0.92	45	141	0.8	7.19	0.029	240	S18 028770
Otahu	2.5	12	26	0.37	40	81	1.6	7.03	0.040	180	S18 979688
Upokongaru	3	72	51	0.18	50	393	0.35	7.65	0.029	40	S22 935473
Hapurua	3.1	43	45	0.28	35	83	0.57	7.01	0.033	180	R18 833670
Whatauma	7.1	50	69	1.83	28	255	0.11	7.6	0.045	40	S21 920780
Kaukore	6.4	17	60	1.31	28	154	0.7	n.a.	0.060	60	R21 862898
Waimarino.Pipiriki	1.8	21	72	9.09	30	136	0.17	7.3	0.050	60	R21 864861
<i>Mudstone Forest</i>											
Crocodile.Creek	4.3	22	56	1.83	26	95	1.4	7.78	0.061	180	R20 876130
Kaiwhakauka	18.6	35	52	1.22	20	92	1.9	7.02	0.061	140	R19 899271
Morinui	11.2	25	53	0.61	22	115	1.7	6.95	0.062	200	S19 953260
Ahuahu	2.3	54	40	0.46	45	147	1.2	7.48	0.035	120	R21 860703
Kaurapaoa	4.3	37	71	0.2	28	211	1.05	7.47	0.056	80	S21 893606
Whirinaki	2.7	14	55	2.62	27	102	1.7	7.2	0.042	140	R20 852115
<i>Hardstone Pasture</i>											
Kaitieke	10.5	32	71	0.61	19	114	0.85	7.01	0.058	180	S19 060131
Kakahi	6	15	68	1.22	22	86	1.65	7.3	0.058	280	S19 016549
Mangakara	3	64	79	3.66	34	94	1.95	7.23	0.049	180	R18 769658
Mangakahu	7.1	40	78	0.46	24	85	1.25	7.18	0.056	200	S18 134720
Orautohu	11.1	36	87	0.92	25	87	1.35	7.68	0.053	310	S20 025054
Pungapunga	10.4	15	96	0.41	28	90	2.45	6.98	0.048	240	S18 200552
Ruatiti	13.1	54	81	0.23	22	78	1.1	7.15	0.057	240	S20 999080
Waimiha	19.5	61	90	0.23	27	60	2.2	6.93	0.047	140	S17 152869
<i>Hardstone Forest</i>											
Makatote	16.4	35	68	1.83	18	74	7.3	6.86	0.065	740	S20 168128
Maramataha	8.8	24	47	0.37	26	52	1.7	6.8	0.049	240	S17 528266
Pumice.Quarry	2.7	43	46	4.57	20	45	3.2	7.01	0.048	380	S19 180420
Upper.Retaruke	22.7	21	55	0.37	22	107	2.15	6.75	0.061	200	S19 440320
Ongarue	11.4	36	62	0.61	30	55	3.25	6.8	0.050	340	S17 245885
Taringamotu	7.2	26	95	0.76	27	73	2.5	6.93	0.052	340	S18 224616
Waimarino.Ruapehu	10.1	36	31	0.73	23	55	3.7	7.05	0.050	740	S20 173180
Whakapapanui	23.8	50	118	1.73	32	120	9.7	7.24	0.052	830	S19 270254
Whanganui	7.5	63	56	1.83	26	94	8.3	7.71	0.050	680	T19 337353

Table 2.2. Physical and chemical characteristics recorded from 30 streams in the Whanganui River catchment, between the 14 and 26 February, 1997, which have mudstone substrates and drain pasture or indigenous forest.

Site	Width (m)	Mean depth (cm)	Mean velocity (cm s ⁻¹)	Slope (degrees)	Stability	Conductivity (μScm^{-2})	Black disk (m))	pH	Substrate size index	Chlorophyll <i>a</i> ($\mu\text{g cm}^{-2}$)	Altitude (m)	Map reference 1:50 000
<i>Pasture</i>												
Ohura	2.3	14.0	129.3	0.92	45	235	1.1	7.4	0.029	0.00035	240	S18 028770
Matarawa	1.9	0.3	69.0	0.25	43	610	0.7	7.3	0.040	0.00530	10	S22 923 393
Makirikiri	2.0	17.7	106.7	0.28	45	213	1.3	7.8	0.029	0.00048	40	S22 934474
Whangamomona	3.6	0.3	198.7	0.10	33	124	0.3	7.5	0.044	0.00004	160	R19 608207
Te Maire	8.1	9.3	116.3	0.72	29	269	0.5	7.3	0.349	0.00150	155	S19 998478
Operiki	2.4	12.3	95.0	1.99	21	450	2.3	7.2	0.061	0.00081	90	S21 971 696
Kokakonui	4.4	11.7	98.3	0.97	32	289	1.3	7.5	0.060	0.00201	150	S19 958416
Motuaruhe	7.1	16.3	133.8	0.37	31	303	2.4	7.3	0.059	0.00195	40	S21 956754
Otahu	2.5	18.3	61.0	0.37	40	109	1.9	7.1	0.040	0.00376	180	S18 979688
Whataumu	4.2	15.0	107.0	1.83	28	395	0.6	7.5	0.045	0.00023	40	S21 920780
Waimarino	2.1	10.3	58.7	9.09	30	269	0.5	7.4	0.050	0.00300	60	R21 864861
Kauwautahi	4.8	18.7	139.0	0.41	31	172	1.5	7.3	0.050	0.00095	240	S19 055390
Tauwhata	3.0	10.3	108.0	0.85	39	161	1.7	7.5	0.050	0.00185	140	R19 882348
Hapurua	2.1	10.3	58.7	0.28	35	137	0.5	7.5	0.033	0.00143	180	R18 833670
Upokongaru	3.0	72.0	61.0	0.18	50	261	1.1	7.2	0.029	0.00020	40	S22 935473
Paparata	1.8	11.0	85.3	1.79	34	166	3.3	7.4	0.047	0.00136	200	R19 743464
Otaupare	7.6	20.3	46.0	0.90	24	243	3.5	7.1	0.064	0.00108	50	R21 882808
<i>Forest</i>												
Mangakara East	3.0	11.3	115.3	3.66	34	104	3.1	7.2	0.049	0.00085	180	R18 769658
Heao	2.9	9.8	129.8	0.25	46	160	0.8	7.2	0.032	0.00088	170	R19 793475
Kaukore	6.4	16.7	129.3	1.31	28	193	0.3	7.3	0.060	0.00291	60	R21 862898
Ahuahu	2.3	23.7	90.3	0.46	45	147	1.1	7.4	0.035	0.00014	120	R21 860708
Morinui	11.2	27.2	166.8	0.61	22	203	2.1	7.8	0.062	0.00161	200	S19 953260
Karapaoa	4.3	37.4	161.7	0.20	26	180	1.3	8.1	0.056	0.00306	80	S21 893606
Pitangi	2.5	14.0	75.3	0.89	32	300	2.0	7.9	1.416	0.00119	20	S22 960582
Tangarakau	14.5	24.0	183.3	0.34	41	153	1.7	7.1	0.047	0.00030	180	R19 733470
Mangare	3.2	0.5	102.5	0.4	40	94	0.5	7.3	0.049	0.00003	160	R19 602276
Mangakara West	6.3	14.8	157.5	1.15	30	76	2.1	7.2	0.045	0.00066	290	R18 763664
Matauru	4.8	25.0	157.0	1.15	27	99	1.8	7.1	0.057	0.00055	155	R19 707457
Kaiwhakauka	18.6	29.7	159.7	1.22	20	198	2.6	7.3	0.061	0.00088	140	R19 899271
Morgans	1.3	6.7	102.7	2.98	17	180	3.3	8.3	0.079	0.00007	180	R19 689460

Algal biomass was assessed (in 1997 only) by extracting photosynthetic pigments (chlorophyll *a* and phaeophytin) in 90% acetone, for 24 hours at 5 °C, from five small cobbles (mean circumference = 10 cm) collected on each sampling occasion.

Wavelength absorbtion was measured on a Jenway 6105 Spectrophotometer. Total pigment concentration was calculated using the method of Moss (1967 a, b) and corrected for stone surface area (Graham *et al.*, 1988).

Data analysis

Data were $\log_{10}(x+1)$ transformed prior to analysis where appropriate to remove heterogeneity of variance. Differences in physicochemical and biological measures among site classes were analysed with one-way analysis of variance (ANOVA) using the SYSTAT statistical package (SYSTAT, 1996). Correlations between the physicochemical variables and six indices of community structure were calculated. These indices were the Simpson's index of evenness (Simpson, 1949), Berger-Parker dominance index (Berger and Parker, 1970), Margelefs index (Clifford and Stevenson, 1975), EPT (Lenat, 1988), MCI (Stark, 1985) and QMCI (Stark, 1993).

Simpson's index of equability or evenness is given by:

$$D = \sum \frac{(n_i - 1)}{(N(N - 1))}$$

and the Berger-Parker dominance index by: $D = N_{\max}/N$ where n_i = number of individuals in the i -th species, N = total number of individuals, S = species number and N_{\max} = number of individuals in the most abundant species. Indices that provided an indication of diversity were taxa richness, and Margalefs index by: $D = (S-1)/1n/ N$ where S = species number and N = total number of individuals. EPT scores consisted of the number of Ephemeroptera, Plecoptera and Trichoptera taxa present. The MCI weights taxa according to their tolerance of organic enrichment. Taxa that are characteristic of higher water quality score more highly than those commonly found in polluted conditions (Stark, 1993). The MCI is given by:

$$MCI = \frac{\text{site score}}{\text{no. of scoring taxa}} \times 20$$

where the site score is the sum of individual taxon scores.

The QMCI (Stark, 1993) is given by:

$$QMCI = \sum_{i=1}^{i=S} \frac{(n_i \times a_i)}{N}$$

where S = the total number of taxa in the sample, n_i is the number of individuals in the i -th scoring taxon, a_i is the score for the i -th taxon, and N is the total number of individuals in the sample.

Community structure was analysed with detrended correspondence analysis (DECORANA), cluster analysis (using $2W/(A+B)$ and the group average), and two-way indicator species analysis (TWINSpan) using the PC-ORD statistical package (McCune and Mefford, 1995). Pseudo-species cut levels used were 1, 10, 100 and 1000. Multi-response permutation procedure (MRPP), a non parametric discriminant analysis was carried out also using the PC-ORD statistical package to assess if there were significant differences in overall community structure between classes.

RESULTS

Between year effects

Physicochemical characteristics

At sites which were re-sampled in 1997, conductivity and water clarity were the only two environmental variables to change significantly (Table 2.3) and both were higher than the previous year (Fig. 2.3). However, both variables were correlated between years (Table 2.3).

Community structure

Of the biological indices only the Berger-Parker Dominance index differed significantly between 1996 and 1997 (Table 2.3). Relative abundances of the major taxonomic groups did not differ statistically between years. Overall there was no significant difference in community structure between 1996 and 1997 (MRPP $r = 0.001$, $P = 0.37$). TWINSpan (Fig. 2.4) indicated close similarities in community structure of most but not all site pairs between years.

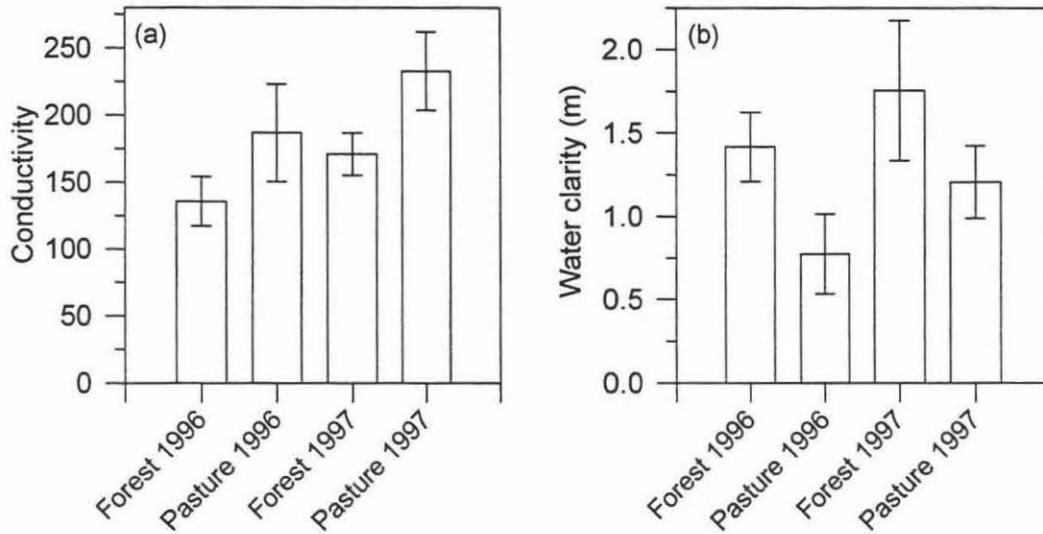


Figure 2.3. Mean (± 1 S.E.) (a) water clarity and (b) conductivity (μScm^{-2}) measured in 15 streams in the Whanganui River catchment, between March and April, 1996, and February, 1997, which have mudstone substrates and drain pasture or indigenous forest.

Land use and geology effects 1996

Physicochemical characteristics

The size of streams varied with mean depth and width ranging from 15 cm to 64 cm and 1.8 m to 23.8 m, respectively. Water clarity was highest in the hardstone forest class with low levels occurring in the mudstone pasture class (Fig. 2.5a). Conductivity displayed the reverse pattern with higher conductivity in mudstone streams (Fig. 2.5b).

Mudstone pasture streams had the lowest channel stability and smallest substrate size. Hardstone streams were wider with faster current velocity. Neither pH nor depth differed significantly among land use/geology classes (Table 2.4). Catchment forest cover was positively correlated with black disk visibility and substrate size and negatively correlated with stability and conductivity (Table 2.5).

Community structure

Of the 112 taxa encountered, 104 were found across all four land use/geology classes. In mudstone pasture streams the gastropod *Potamopyrgus antipodarium* was numerically dominant and on average accounted for 45% of the individuals present (Fig. 2.6).

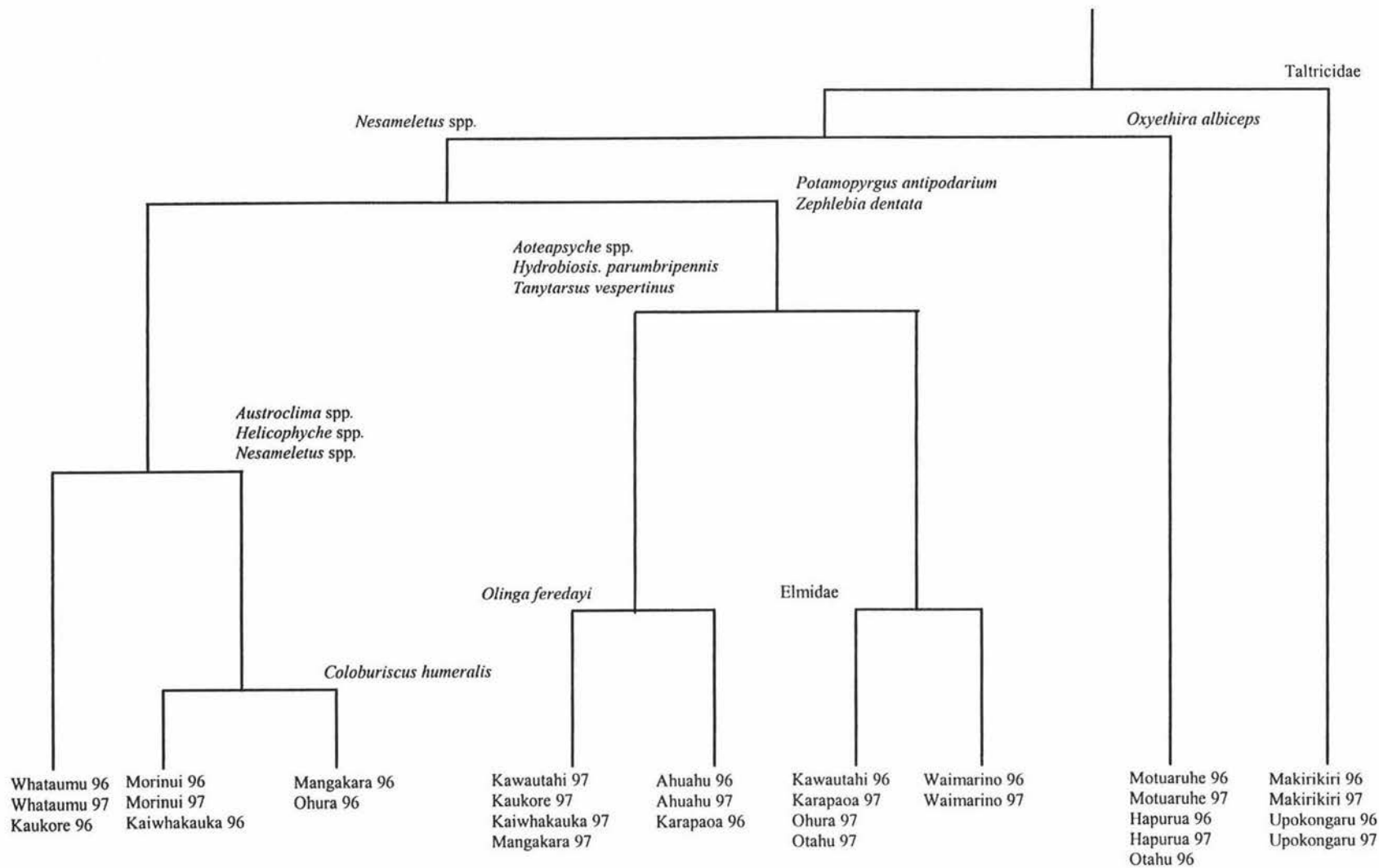


Figure 2.4. TWINSpan classification of 15 mudstone streams sampled between March and April, 1996, and February, 1997.

Table 2.3. Paired t-tests and Pearson correlations between environmental and biotic measurements for 15 streams sampled in March to April, 1996 and February, 1997, which have mudstone substrates and drain pasture or indigenous forest.

Variable	<i>t</i> statistic	<i>P</i> value	<i>r</i> value	<i>P</i> value
Conductivity	2.95	0.01	0.67	0.01
Velocity	-1.31	0.21	0.29	0.29
Water clarity	2.71	0.02	0.73	0.00
Total abundance	1.16	0.27	0.12	0.68
pH	1.19	0.25	0.14	0.61
Taxa richness	0.69	0.50	0.81	0.00
Margalefs index	-0.77	0.46	0.58	0.02
Simpson's index	-1.27	0.23	0.34	0.22
Berger-Parker index	2.63	0.02	0.48	0.07
MCI	0.29	0.77	0.38	0.17
QMCI	1.12	0.28	0.48	0.07
EPT score	0.28	0.78	-0.34	0.22

Ephemeroptera dominated in the mudstone forest streams. In contrast, hardstone streams had a greater relative abundance of Trichoptera (36%) and Coleoptera (21%) than those containing soft substrate (12% and 8%, respectively). Proportions of Coleoptera and Diptera were higher and Ephemeroptera and Trichoptera lower as a result of agricultural development.

Diversity as assessed by taxonomic richness and Margalefs index was lowest in the mudstone pasture class and highest in the mudstone forest class with similar levels of diversity in the two hardstone groups (Fig. 2.7a). While abundance did not differ significantly among groups hardstone pasture streams clearly had the most individuals with similar abundance in the other three groups (2.7b). Neither Simpson's or Berger-Parker indices differed significantly among classes (Table 2.4).

MCI scores were highest in those classes with forested catchments whereas QMCI values did not differ significantly (Fig. 2.8b) (Table 2.2) although it revealed a similar trend to the MCI. The number of Ephemeroptera, Plecoptera and Trichoptera (EPT) was greatest in mudstone forested streams (Fig. 2.8a). MCI values were correlated positively with water clarity, catchment forest cover, substrate size, and negatively with conductivity, stability and pH while QMCI was correlated negatively with conductivity and positively with slope (Table 2.5).

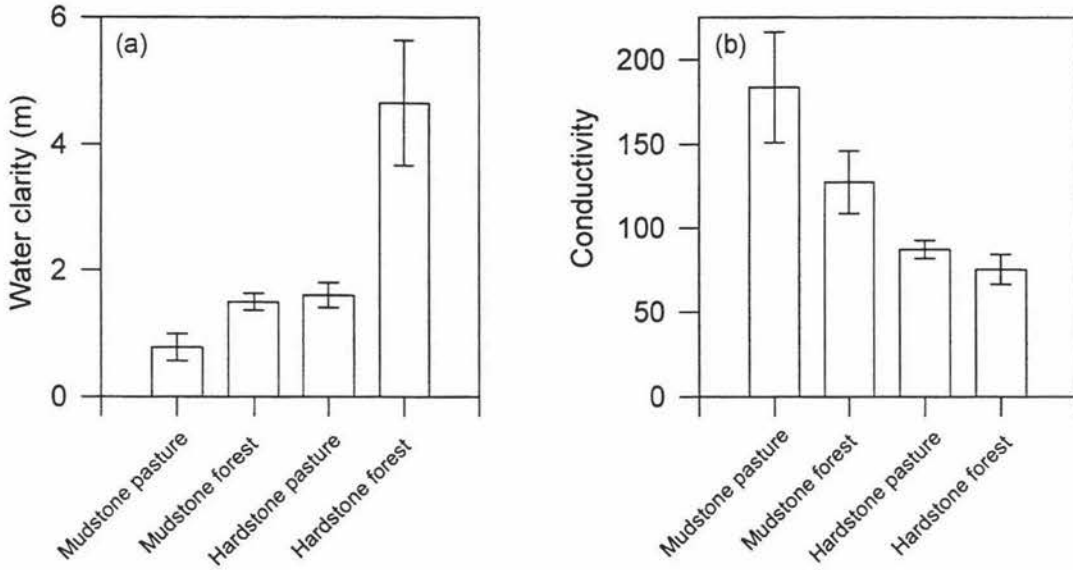


Figure 2.5. Mean (± 1 S.E.) (a) water clarity and (b) conductivity (μScm^{-2}) measured in 33 streams in the Whanganui River catchment, between March and April, 1996, which have mudstone or hardstone substrates and drain pasture or indigenous forest.

Table 2.4. One-way analysis of variance tests between environmental and biotic measurements for 33 streams in the Whanganui River catchment, between March and April, 1996, which have mudstone or hardstone substrates and drain pasture or indigenous forest.

Variable	F Ratio	D.F.	P value	Variable	F Ratio	D.F.	P value
Physical				Biological			
Conductivity	7.91	3,29	0.00	Total abundance	1.76	3,29	0.18
Water clarity	5.67	3,29	0.00	Taxa richness	3.2	3,29	0.04
Stability	6.3	3,29	0.00	Margalefs index	3.02	3,29	0.05
Substrate size	3.63	3,29	0.02	Simpson's index	1.33	3,29	0.29
Forest cover	8.45	3,29	0.00	Berger-Parker index	2.34	3,29	0.09
Width	4.98	3,29	0.01	MCI	7.86	3,29	0.00
Depth	0.84	3,29	0.48	QMCI	2.16	3,29	0.11
Velocity	3.52	3,29	0.03	EPT score	2.34	3,29	0.09
pH	1.99	3,29	0.14	Axis 1 scores	6.0	3,29	0.00
Slope	0.44	3,29	0.73	Axis 2 scores	4.13	3,29	0.02
Altitude	14.7	3,29	0.00				

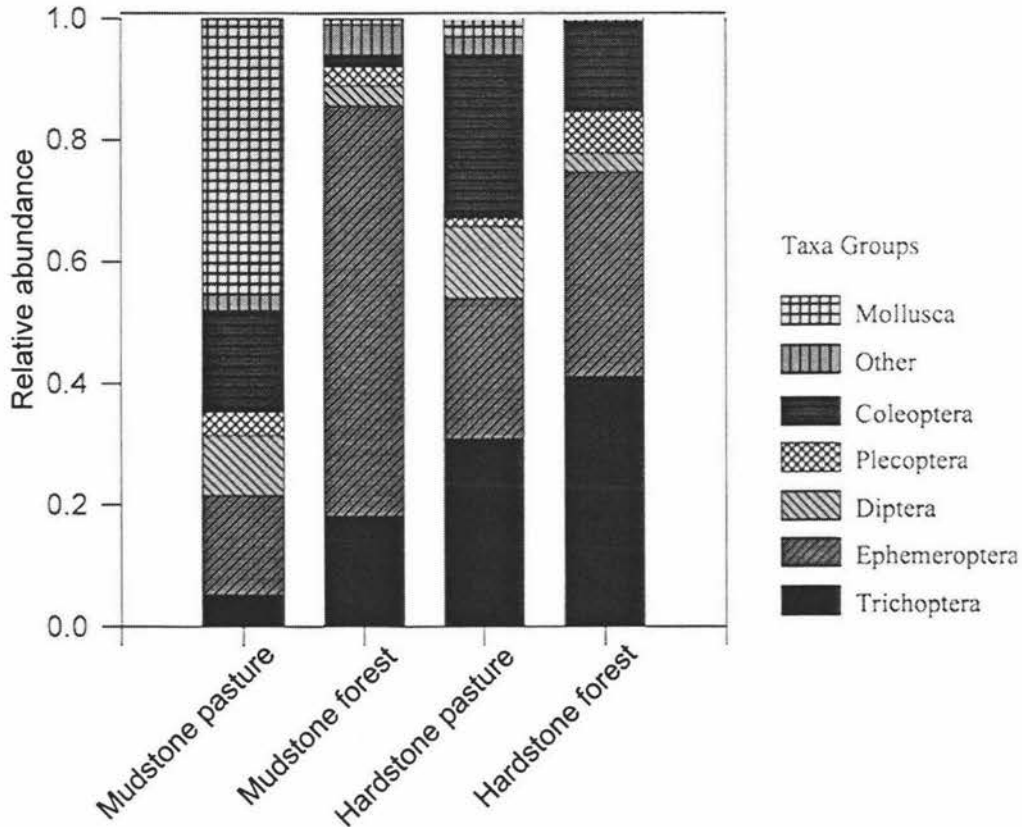


Figure 2.6. Mean relative abundance of individuals from seven taxonomic groups measured in 33 streams in the Whanganui River catchment, between March and April, 1996, which have mudstone or hardstone substrates and drain pasture or indigenous forest.

In the ordination analysis land use classes (of both geology types) were segregated on axis 1 (Fig. 2.9) with geology separated on axis 2, particularly between forested streams. The percentage of variance explained by axis 1 and 2 was 32% and 11%, respectively. Increasing axis 1 scores were indicative of sites with less pollution sensitive taxa particularly in the mudstone pasture class. *P. antipodarium* was strongly associated with these high axis 1 scores while *Deleatidium* spp. commonly occurred at sites that scored low on axis 1. Mudstone substrate streams were located higher on axis 2 with *Austroclima* spp., *Pycnocentria* spp. and *Zephlebia dentata* commonly occurring here as well. Elmids preferred streams low on axis 2. A number of variables were significantly correlated with the first DECORANA axis. These included positive correlations with conductivity, stability, and negative correlations with water clarity, slope, substrate size and catchment forest cover. Velocity was negatively correlated with the second DECORANA axis (Table 2.5).

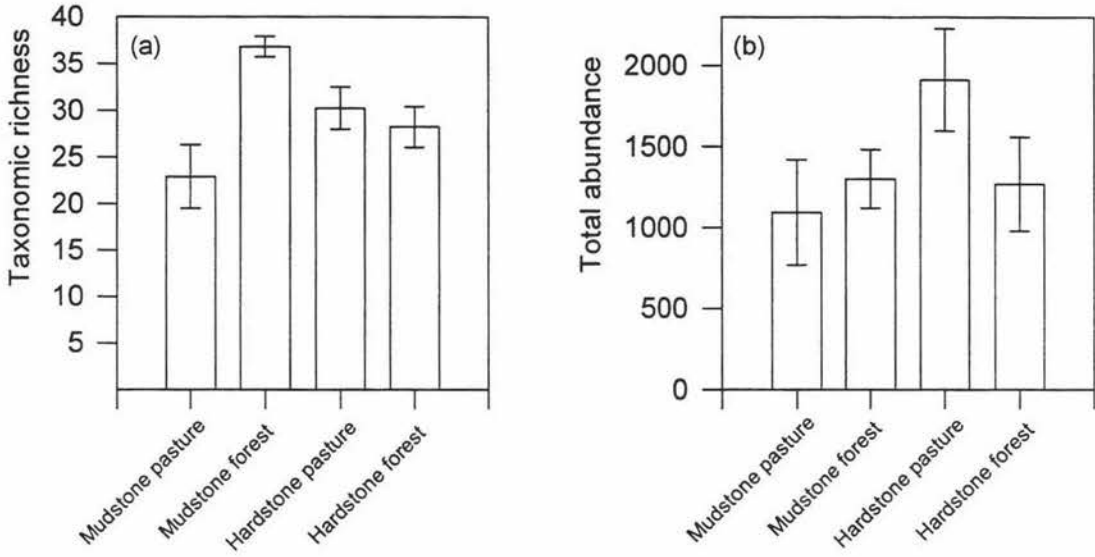


Figure 2.7. Mean (± 1 S.E.) (a) taxonomic richness and (b) total abundance measured in 33 streams in the Whanganui River catchment, between March and April, 1996, which have mudstone or hardstone substrates and drain pasture or indigenous forest.

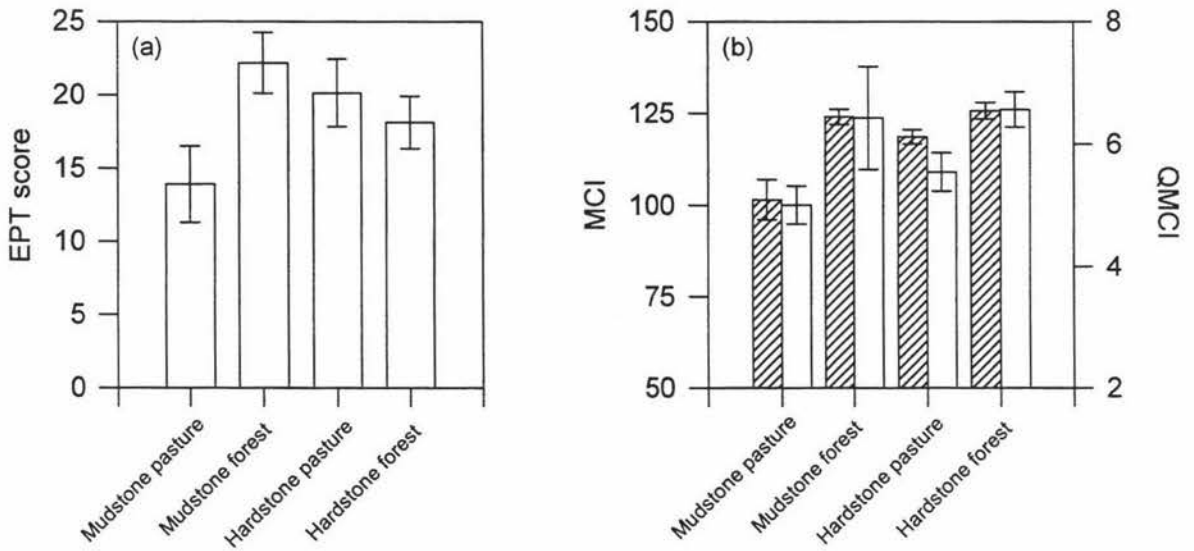


Figure 2.8. Mean (± 1 S.E.) (a) EPT scores and (b) MCI (diagonal hatching) / QMCI (no hatching) measured in 33 streams in the Whanganui River catchment, between March and April, 1996, which have mudstone or hardstone substrates and drain pasture or indigenous forest.

Table 2.5. Pearson correlations between environmental and biotic measurements for 33 streams in the Whanganui River catchment, between March and April, 1996. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$.

	Water clarity	Conductivity	pH	Substrate size	Stability	Width	Depth	Velocity	Forest cover %	Slope (degrees)	Altitude
Conductivity	**0.55	-	-	-	-	-	-	-	-	-	-
pH	-0.28	*0.37	-	-	-	-	-	-	-	-	-
Substrate size	*0.4	*-0.40	-0.23	-	-	-	-	-	-	-	-
Stability	*-0.36	***0.52	0.35	***-0.84	-	-	-	-	-	-	-
Width	**0.5	-0.33	*-0.38	***0.58	***-0.59	-	-	-	-	-	-
Depth	0.05	0.03	0.19	-0.13	0.05	0.18	-	-	-	-	-
Velocity	0.09	0.1	0.1	0.34	-0.23	0.35	0.15	-	-	-	-
Forest cover %	-0.39*	**0.45	-0.34	*0.43	*-0.42	0.32	0.17	0.04	-	-	-
Slope	-0.04	-0.20	0.14	0.23	-0.26	-0.17	0.01	0.08	0.16	-	-
Altitude	***0.58	***-0.6	**0.54	0.13	*-0.36	*0.39	0.17	0.07	0.26	0.21	-
Taxa richness	0.26	***-0.56	-0.25	**0.47	**0.46	0.16	-0.09	0.35	*0.41	-0.05	0.07
Total abundance	0.2	-0.27	-0.25	*0.39	*-0.36	0.14	*-0.44	0.16	0.07	0.08	0.31
Margalefs index	*0.36	***-0.59	-0.22	**0.53	**0.53	0.25	0.07	-0.2	**0.46	0.18	0.09
Simpson's index	-0.18	0.28	-0.06	-0.28	*0.35	-0.01	-0.1	0.02	-0.2	-0.34	-0.07
Berger-Parker index	-0.27	*0.36	0.04	-0.34	*0.42	-0.06	-0.03	0.04	-0.27	-0.30	0.19
MCI	***0.59	***-0.70	*-0.37	**0.46	***-0.49	0.32	0.03	-0.17	**0.48	0.22	0.32
QMCI	0.28	**0.49	-0.18	0.22	-0.26	0.11	-0.06	-0.22	0.28	***0.54	0.26
EPT score	0.28	***-0.62	-0.27	***0.49	**0.45	0.2	-0.07	-0.13	0.32	0.32	0.09
Axis 1 scores	**0.38	**0.5	0.26	***-0.64	***0.62	-0.33	0.06	-0.05	**0.48	-0.6	-0.27
Axis 2 scores	0.02	0.14	0.27	-0.24	0.2	-0.25	0.21	*-0.36	0.13	-0.04	-0.07

TWINSPAN analysis formed five groups (Fig. 2.10) comprising of two silt bottom streams (E) and four main groups (A, B, C and D). Group A and D consist of streams with predominantly soft substrates while B and C consisted of those with mainly hard substrates. There were significant differences among communities in the four land use/geology classes when analysed using MRPP ($r = 0.078$, $P = 3 \times 10^{-6}$). An average distance (Sorenson) of 0.87 within the mudstone pasture class indicated that community structure varied most among streams within this class whereas average distance within the mudstone forest, hardstone pasture and hardstone forest classes were 0.57, 0.67 and 0.65, respectively. This is also evident in the DECORANA analysis (Fig. 2.9).

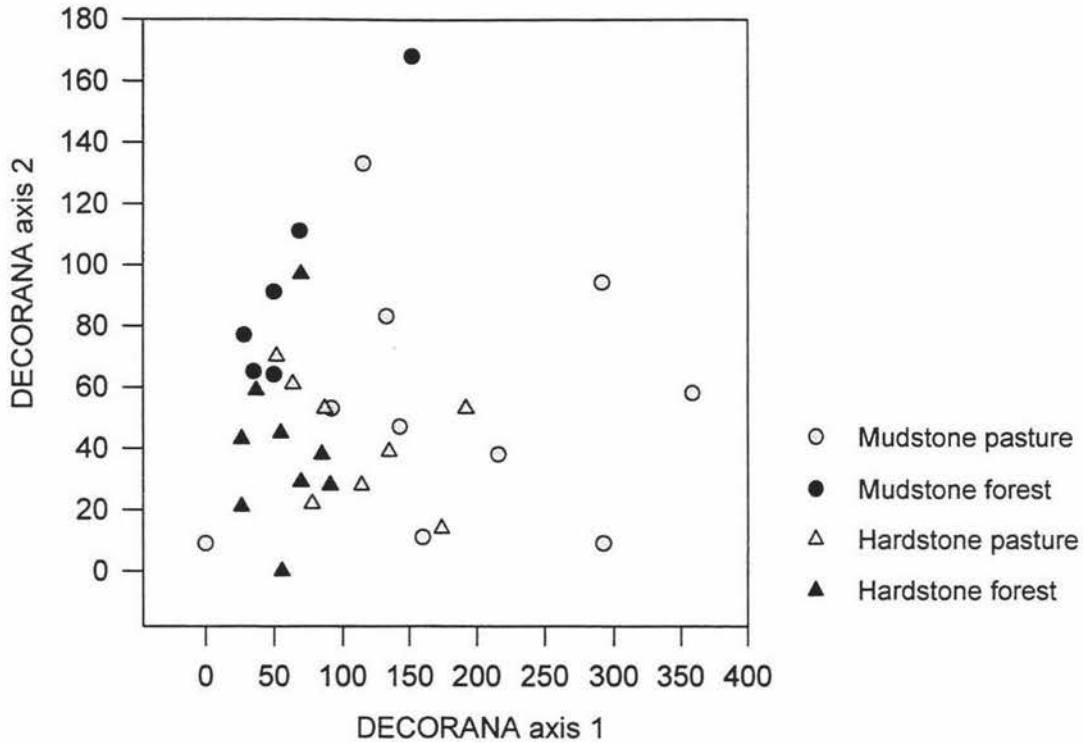


Figure 2.9. Axis 1 as a function of axis 2, for DECORANA analysis of community structure measured in 33 streams in the Whanganui River catchment, between March and April, 1996, which have mudstone or hardstone substrates and drain pasture or indigenous forest.

Land use effects 1997

Physicochemical Characteristics

At sites sampled in 1997 only two physicochemical parameters differed significantly between indigenous forest and pastoral land uses (Table 2.6). Conductivity was higher in the pasture streams (Fig. 2.11a) while velocity was lowest at these sites.

Community structure

The MCI and QMCI biotic indices and taxonomic richness were higher in the forested class (Fig. 2.11b). However, measures of evenness did not differ (Table 2.6). As in 1996, the pasture class had high numbers of Mollusca (34%), mainly *P. antipodarium*. Crustacea (Talitridae, Amphipoda) were also common at pasture sites but principally at Upokongaro and Makirikiri. Therefore this abundance of Crustacea is not representative of the whole pasture class. Ephemeroptera were relatively more abundant in the forested class (40% opposed to 28%). *Deleatidium* spp., *Austroclima* spp. and *Nesameletus* spp. were all common in the forest class.

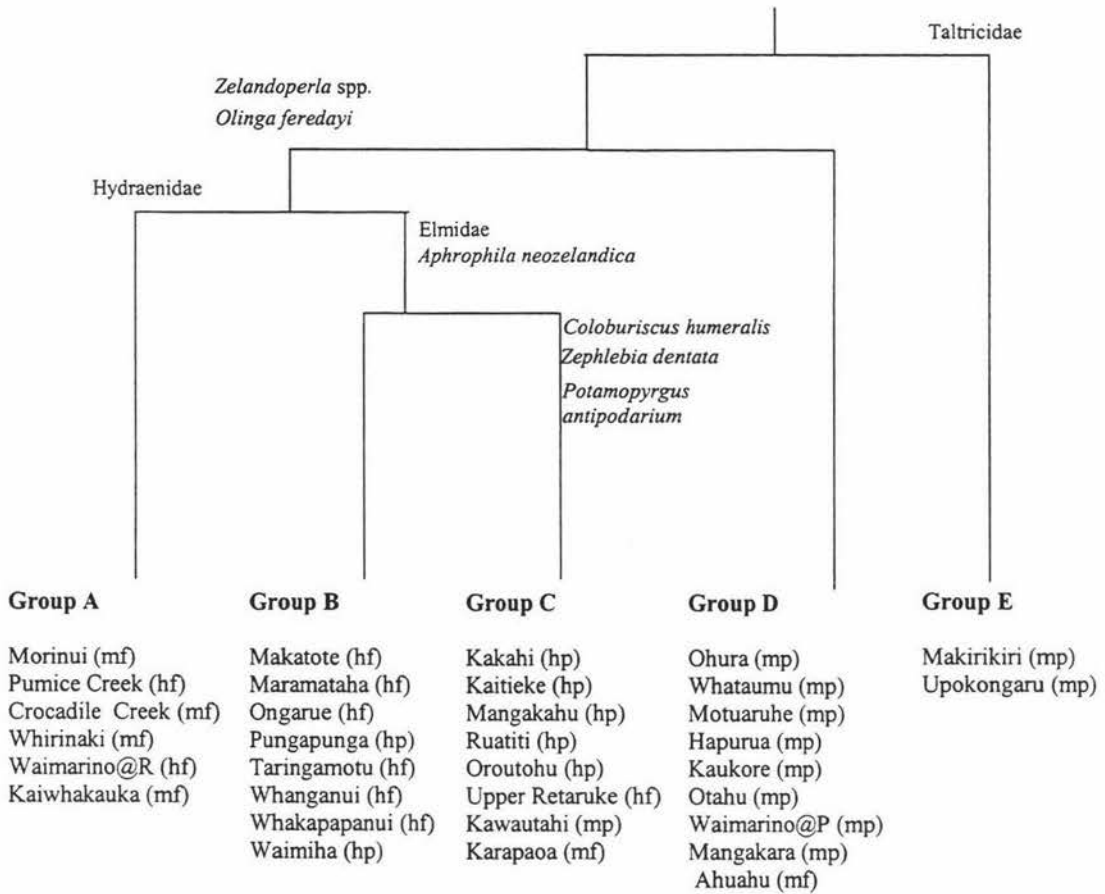


Figure 2.10. TWINSpan analysis of 33 streams in the Whanganui River catchment sampled between March and April, 1996. m = mudstone, h = hardstone, f = forest, p = pasture.

Trichoptera were also abundant at forested sites and included *Pycnocentria funeria*, *Zelolessica cheira*, and *Olinga feredayi*. Plecoptera were rare in both classes but more abundant at forested sites (Fig. 2.12).

Overall community structure of the bush and pasture classes differed significantly (MRPP, $r=0.025$, $P=0.01$). Cluster analysis and TWINSpan did not group sites distinctly in terms of land use although the initial groups formed using cluster analysis did comprise predominantly of bush or pasture sites. DECORANA also grouped bush sites to the lower left of a plot of axis 1 on 2 although there was considerable spread. (Fig. 2.13).

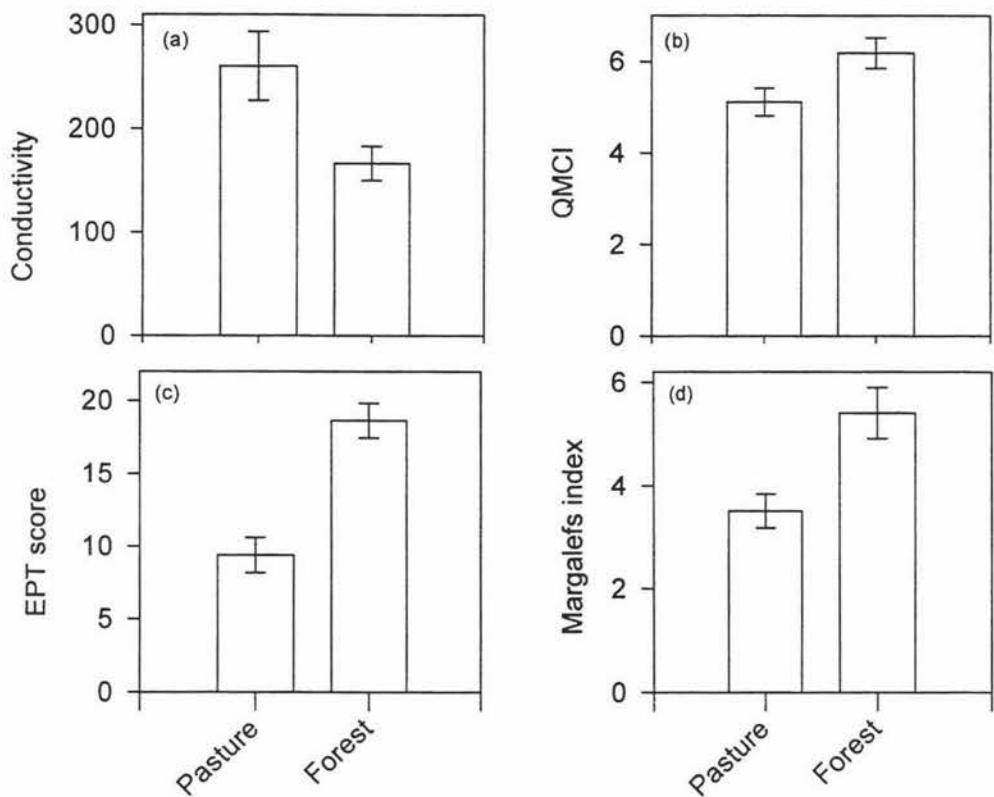


Figure 2.11. Mean (± 1 S.E.) (a) conductivity, (b) QMCI, (c) EPT score and (d) Margalefs index measured in 30 streams in the Whanganui River catchment, February, 1997, which have mudstone substrates and drain pasture or indigenous forest.

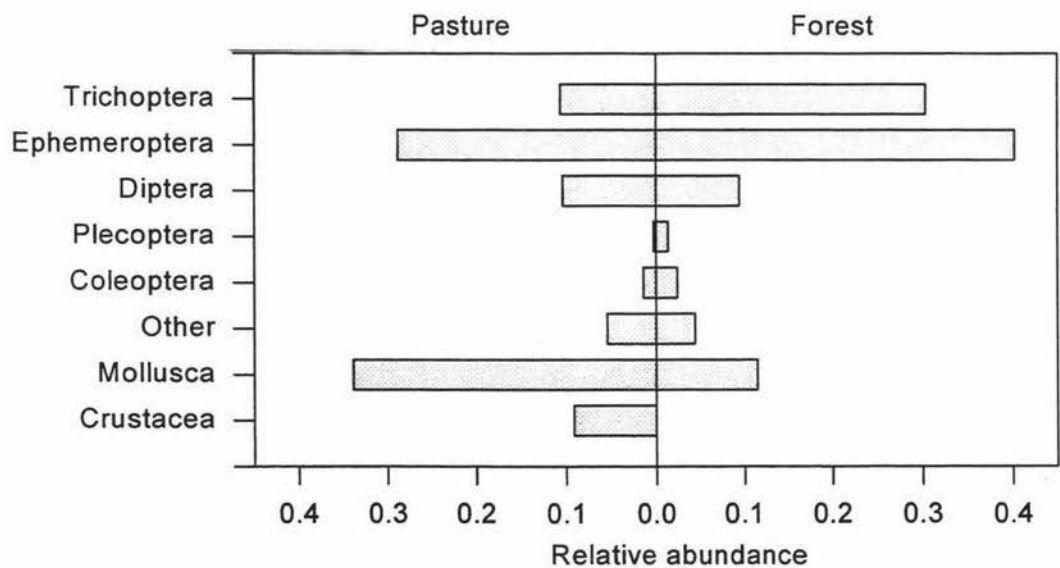


Figure 2.12. Mean relative abundance of individuals measured in 30 streams in the Whanganui River catchment, February, 1997, which have mudstone substrates and drain pasture or indigenous forest.

Table 2.6. One-way analysis of variance tests between environmental and biotic measurements for 30 streams in the Whanganui River catchment, February, 1997, which have mudstone substrates and drain pasture or indigenous forest.

Variable	F Ratio	D.F.	P value	Variable	F Ratio	D.F.	P value
Physical				Biological			
Conductivity	7.69	1,29	0.01	Chlorophyll <i>a</i>	0.16	1,28	0.70
Velocity	7.14	1,29	0.01	Total abundance	1.60	1,29	0.22
Water clarity	0.82	1,29	0.37	Taxa richness	9.46	1,29	0.01
Stability	1.51	1,29	0.23	Margalefs index	10.1	1,29	0.00
Substrate size	1.08	1,29	0.31	Simpson's index	1.01	1,29	0.32
Forest cover %	49.91	1,29	0.00	Berger-Parker index	1.94	1,29	0.18
Width	2.44	1,29	0.13	MCI	17.92	1,29	0.00
Depth	0.55	1,29	0.46	QMCI	6.65	1,29	0.02
pH	0.48	1,29	0.49	EPT score	15.84	1,29	0.00
Slope	0.26	1,29	0.62	Axis 1 scores	3.55	1,29	0.07
Altitude	1.46	1,29	0.24	Axis 2 scores	9.54	1,29	0.0

Table 2.5. Pearson correlations between environmental and biotic measurements for 30 streams in the Whanganui River catchment, February, 1997, which have mudstone substrates and drain pasture or indigenous forest. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$.

Variable	Chlorophyll <i>a</i>	Water clarity	Conductivity	pH	Substrate size	Stability	Width	Depth	Velocity	Forest cover %	Slope	Altitude
Water clarity	0.18	-	-	-	-	-	-	-	-	-	-	-
Conductivity	0.28	-0.07	-	-	-	-	-	-	-	-	-	-
pH	-0.15	0.07	0.14	-	-	-	-	-	-	-	-	-
Substrate size	0.21	0.11	0.23	0.28	-	-	-	-	-	-	-	-
Stability	-0.18	-0.34	-0.12	-0.33	-0.34	-	-	-	-	-	-	-
Width	0.19	0.10	-0.08	-0.31	0.14	-0.32	-	-	-	-	-	-
Depth	0.21	**0.46	-0.03	0.00	0.03	-0.16	0.31	-	-	-	-	-
Velocity	-0.20	-0.02	-0.32	0.07	-0.04	-0.19	***0.50	0.01	-	-	-	-
Forest cover %	0.04	0.32	***-0.52	0.07	0.17	-0.31	0.32	*0.37	0.18	-	-	-
Slope	0.24	0.30	0.01	0.03	0.20	**0.49	-0.09	0.20	-0.17	0.17	-	-
Altitude	-0.19	0.16	***-0.71	-0.10	-0.24	-0.11	0.17	0.14	*0.43	0.27	0.10	-
Taxa richness	0.12	0.09	-0.19	-0.05	0.16	-0.29	0.35	0.34	0.12	*0.42	0.07	0.17
Total abundance	0.09	0.23	-0.08	-0.13	0.12	0.00	0.06	*0.39	-0.09	0.21	-0.30	-0.10
Margalefs index	0.11	0.1	-0.26	-0.00	0.13	-0.29	0.30	0.20	0.19	0.34	0.12	0.24
Simpson's index	*-0.37	*0.03	-0.13	0.13	-0.09	0.23	*-0.39	-0.5	-0.13	-0.10	0.05	-0.05
Berger-Parker index	-0.21	-0.09	0.06	-0.05	-0.06	0.39	-0.34	-0.27	-0.16	-0.28	0.01	-0.21
MCI	-0.00	*0.41	-0.39	-0.06	0.08	***-0.5	0.28	0.34	0.34	***0.62	**0.48	*0.44
QMCI	-0.05	0.22	-0.25	-0.02	-0.00	**0.33	0.13	0.05	0.25	0.25	0.17	0.23
EPT score	0.05	0.19	-0.30	0.02	0.13	*-0.43	*0.38	*0.39	0.18	***0.57	0.14	0.28
Axis1 scores	0.02	*0.06	0.22	0.03	-0.12	***0.60	*-0.41	0.03	***-0.5	-0.23	*-0.40	*-0.43
Axis2 scores	-0.16	-0.54	0.26	0.02	0.01	0.23	-0.03	-0.34	-0.03	*-0.47	-0.34	-0.18

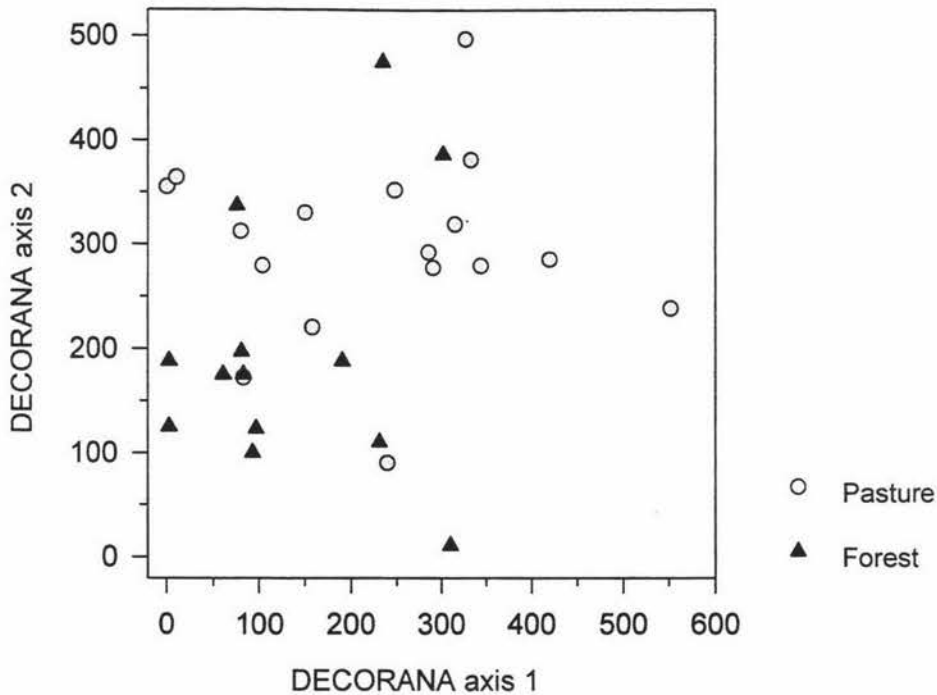


Figure 2.13. DECORANA analysis of community structure measured in 30 streams in the Whanganui River catchment, February, 1997, which have mudstone substrates and drain pasture or indigenous forest.

The percentage of variation explained by DECORANA axis 1 and 2 was 33% and 19%, respectively. Taltricidae, *P. antipodurium* and *Physa* spp. were associated with high axis 1 scores, with the mayflies *Nesameletus* spp., *Acanthophlebia cruentata* positioned at the opposite end. On the high end of axis 2 sites with *Hydrobiosis umbripennis* contrasted those at the lower with greater abundance of the mayfly's *Deleatidium* spp. and *Nesameletus* spp.

DISCUSSION

Many physical and biological factors varied among the four land use and geology classes. Multivariate analysis of community structure suggests that macroinvertebrate assemblages have different characteristics depending on geology type and catchment land use and that these differences are significant. A distinction between geological classes provided by physical and biological variables was apparent and is accentuated by pasture land use.

Other studies have noted the importance of geomorphology and climate in influencing macroinvertebrate communities (Harding, 1994) and algal biomass (Friberg and Winterbourn, 1997). Variations in geology may also mask the affects of land use between catchments (Richards *et al.*, 1997).

Few environmental measurements or biotic characteristics were statistically different between 1996 and 1997 suggesting some continuity of sites between years. Lower flow conditions were responsible for greater conductivity (e.g., Henderson, 1996) and clarity in 1997. Temporal variation in community structure is considered to be a result of the combined effects of disturbance frequency and habitat heterogeneity (Townsend, 1989; Poff and Ward, 1990; Scarsbrook and Townsend, 1993; Townsend and Hildrew, 1994). While species diversity was similar between 1996 and 1997 numbers of dominant taxa differed significantly so the abundance of many common taxa may relate more to sampling period rather than site location. No major changes occurred in the main taxonomic orders between 1996 and 1997 apart from a large increase in Amphipods at two sites, both of which had sand substrate and low current velocity, conditions that suit aquatic Amphipoda.

Conductivity was higher in mudstone catchments of both land use types due to greater levels of erosion which continually expose fresh material to weathering (Biggs and Gerbeaux, 1993; Maasdam and Smith, 1994; Biggs, 1995; Davies-Colley and Stroud, 1995). There was high relative and numerical abundances of Ephemeroptera, principally *Coloburiscus humeralis*, as well as *Zephlebia dentata* and *Deleatidium* spp. in mudstone forested streams and it appears that Trichoptera and Coleoptera prefer streams in harder Andesitic/Jurassic catchments. Cased caddisflies may be less adept at adhering to softer, smoother substrate and require greater surface irregularities to resist removal, therefore making them more prone to dislocation than mayflies. The cased caddisflies *Olinga feredayi* and *Helicopsyche* spp. were found almost exclusively in hardstone streams. Contrasting to this another cased caddisfly, *Pycnocentria funerea* appeared to be almost exclusively found in mudstone forested streams and was the main species of caddis fly found in this class. *Pycnocentria funerea* are known to favour woody substrate (Henderson pers. comm.) and this may be related to greater amounts of this material present in these streams.

Dominance by the snail *Potamopyrgus antipodarium* in mudstone pasture streams can be attributed to the affects of agricultural development such as mild enrichment (e.g., Scott *et al.*, 1994; Harding and Winterbourn, 1995; Quinn *et al.*, 1997). Agricultural development was responsible for increases in the relative proportion of Diptera and Coleoptera, and a decrease in Ephemeroptera and Trichoptera. Similar species were found within all classes but the proportions of these taxa differed. In 1997, a third of all Trichoptera individuals in the mudstone forest class consisted of the cased caddis fly species *P. aureola* but were collected almost exclusively from one stream. *P. aureola* does not appear to be common in mudstone streams judging from abundance levels in both the 1996 and 1997 studies. This species commonly occurs in larger lowland streams of moderate to swift flow and feed on algae and fine detritus (Cowley, 1978). However, Pitangi stream where nearly all of the individuals were collected is small with comparatively moderate to low algal growth, velocity and channel stability. Most larvae consisted of small early instar individuals which coincides with what would be predicted for this species in autumn (Cowley, 1978). It is possible that chance oviposition by a few females could account for the large number of *P. aureola* larvae present in a particular reach as has been shown for an Australian cased caddisfly (Bunn and Hughes, 1997). Trichoptera were still a large proportion of the taxa found in streams suffering less from the impacts associated with agricultural development.

Conductivity was also higher in pasture streams independent of geology as found elsewhere (Close and Davies-Colley, 1990; Collier, 1995). In both years conductivity was an important chemical measure of land use as streams with predominantly forested catchments had lower conductivity. Parameters such as low turbidity, conductivity and a higher proportion of indigenous forest cover within a catchment appeared to relate to community structure and the number of highly scoring pollution sensitive taxa associated with the MCI water quality index. However, when accounting for the lower relative abundance of these high scoring tolerant species the indigenous forest streams appeared to have more of them hence a QMCI equivalent to any other class.

Many studies have found agricultural development to have a negative effect on species richness (Allan, 1959; Dance and Hynes, 1980; Winterbourn *et al.*, 1981; Lenat, 1988; Quinn and Hickey, 1990b; Scott *et al.*, 1994; Harding and Winterbourn, 1995).

In this study pasture streams had lower diversity although taxa richness was higher in hardstone pasture opposed to hardstone forest streams which is due to a greater abundance of individuals at the hardstone forest sites. Improved algal productivity brought about by increased levels of light (e.g., Behmer and Hawkins, 1986; Friberg and Winterbourn, 1997) and enrichment (Quinn *et al.*, 1992) may also be responsible for higher taxonomic richness and total abundance in the hardstone pasture streams. Increased periphyton biomass may lead to higher invertebrate abundance in mildly impacted streams (e.g., Jowett and Richardson, 1990). However, water quality at the hardstone pasture sites was still relatively high as indicated by low conductivity, turbidity and high MCI scores for these streams. The mean MCI score for the hardstone pasture class was 118 which is not considered polluted as scores of over 100 are considered indicative of 'healthy' streams (Stark, 1993). Predictably taxa diversity and the presence of pollution sensitive taxa positively related to the percentage of catchment in native forest as occurred in 1996 and elsewhere (Allan, 1959; Dance and Hynes, 1980; Winterbourn *et al.*, 1981; Lenat, 1988; Quinn and Hickey, 1990b; Scott *et al.*, 1994; Harding and Winterbourn, 1995). Diversity and pollution sensitive taxa were also negatively influenced by turbidity and suspensoids as extrapolated from visibility measurements (Davies-Colley and Stroud, 1995) which also coincides with other studies (Wagener and LaPerriere, 1985; Quinn *et al.*, 1992; Richards and Host, 1994).

Turbidity levels were higher in mudstone streams regardless of land use. However, these levels were accentuated by the use of land for agricultural purposes (Ryan, 1991; Quinn *et al.*, 1992; Harding and Winterbourn, 1995; Townsend *et al.*, 1997). If conductivity and turbidity are used to measure the effects of land use on waterways allowances must be made for inherent differences in conductivity and suspensoids that occur between streams when attempting to quantify water quality across streams that differ in geology type. Bed stability was lower in mudstone pasture streams compared to the other three classes. Due to the high erodibility of soft sedimentary rock (Maasdam and Smith, 1994) and a lack of indigenous vegetation (Hanchet, 1990; Townsend *et al.*, 1997) higher levels of unstable sediment which require lower critical stream velocities to move them (Newbury, 1984; Carson and Griffith, 1987; Ashworth and Fergusson, 1989) were found in mudstone pasture streams (e.g., Collier, 1995).

Lower abundance and diversity in mudstone pasture streams reflects the susceptibility of these catchments to the impacts brought about from agricultural development. Increased turbidity, suspended sediment and silt deposition have been implicated in other studies leading to reduced invertebrate species richness (Wagener and LaPerriere, 1985; Quinn *et al.*, 1992; Richards and Host, 1994) and abundance (Ryder, 1989; Quinn *et al.*, 1992). Decreases in macroinvertebrate abundance are thought to result from reduced primary production (Quinn *et al.*, 1992), a reduction in food quality (Stumn and Morgan, 1981; Gregory, 1983; Sweeney and Vannote, 1986), bed clogging and low interstitial oxygen (Ryder, 1989). Other studies have found that periphyton biomass and productivity can be increased (Quinn and Hickey, 1990b), decreased (Harding and Winterbourn, 1995) or unaffected (Collier, 1995; Townsend *et al.*, 1997) by agricultural activity. Through agricultural development, light access to the stream channel will increase as a result of the removal of riparian vegetation. However, a corresponding increase in suspensoids (Ryan, 1991) particularly in catchments with more erodable geology's may shade the benthos (Van Nieuwenhuyse and LaPerriere, 1986; Ryan, 1991; Davies-Colley *et al.*, 1992) countering light increases above the water column. In this study chlorophyll *a* concentrations were similar between the pasture and native forest classes and appeared to be unrelated to any other measured biotic or abiotic variables. Geology type may override any influence of land use on periphyton.

The many physical and biological variables correlated with axis 1 of the DECORANA analysis suggests that while not all variables differ significantly between land use and geology, or between themselves, a combination of many are responsible for influencing invertebrate community structure. The selection of sites contained in this study are not completely dichotomous in their land use or geological properties and this gradient is evident. However, it is clear from multivariate analysis that invertebrate community structure differs among land use and geological classes, and that these differences are a result of variations in environmental conditions distinct to these classes.

REFERENCES

- Allan, K.R. 1959. Effect of land development on stream bottom faunas. Proceedings of the New Zealand Ecological Society **7**: 20-21.
- Ashworth, P.J. and Ferguson, R.I. 1989. Size-selective entrainment of bed loads in gravel bed streams. Water Resources Research **25**: 627-634.
- Behmer, D. J. and Hawkins, C. P. 1986. Effects of overhead canopy on macroinvertebrate production in a Utah stream. Freshwater Biology **16**: 287-300.
- Berger, W.H. and Parker, F.L. 1970. Diversity of plankton Foraminifera in deep sea sediments. Science **168**: 1345-1347.
- Biggs, B.J.F. 1995. The contribution of flood disturbance, catchment geology and land use to the habitat template of periphyton in stream ecosystems. Freshwater Biology **33**: 419-438.
- Biggs, B.J.F. and Gerbeaux, P. 1993. Periphyton development in relation to macro-scale (geology) and micro-scale (velocity) limiters in two gravel bed rivers, New Zealand. New Zealand Journal of Marine and Freshwater Research **27**: 39-53.
- Brussven, M.A. 1984. The distribution and abundance of benthic insects subjected to reservoir release flows in the Clearwater River, Idaho, USA. In: Lillehammer, A. and Saltveit, S.J., editors. Regulated rivers. Oslo: Univeritetsforlaget AS. p 167-180.
- Bunn, S. and Hughes, J. 1997. Dispersal and Recruitment in streams: evidence from genetic studies. Journal of the North American Benthological Society **16(2)**: 338-346.
- Carson, M.A. and Griffiths, G. A. 1987. Bedload transport in gravel channels. Journal of Hydrology New Zealand **26**: 1-151.

- Chapman, A. and Lewis, M. 1976. An introduction to the freshwater Crustacea of New Zealand. Auckland: Collins.
- Clifford, H.T. and Stephenson, W. 1975. An Introduction to Numerical Classification. New York: Academic press.
- Close, M.E. and Davies-Colley, R.J. 1990. Base flow water chemistry in New Zealand rivers. 2. Influence of environmental factors. New Zealand Journal of Marine and Freshwater Research **24**: 343-356.
- Collier, K.J. 1995. Environmental factors affecting the taxonomic composition of aquatic communities in lowland waterways of Northland, New Zealand. New Zealand Journal of Marine and Freshwater Research **29**: 453-465.
- Corkum, L.D. 1990. Intrabiome distributional patterns of lotic macroinvertebrate assemblages. Canadian Journal of Fisheries and Aquatic Sciences **47**: 2147-2157.
- Cowie, B. 1985. An analysis of changes in the invertebrate community along a southern New Zealand montane stream. Hydrobiologia **120**: 35-46.
- Cowley, D.R. 1978. Studies on the larvae of New Zealand Trichoptera. New Zealand Journal of Zoology **5**: 639-750.
- Cummins, K.W., Minshall, G. W., Sedall, J. R., Cushing, C. E. and Petersen, R. C. 1984. Stream Ecosystem Theory. Verhandlungen der Internationalen Vereinigung für theoretisch und angewandte Limnologie **22**: 1818-1827.
- Currie, K.J. and Gilliland, B.W. 1980. Baseline water quality of the Manawatu Water Region, 1977-78. Water and Soil Miscellaneous Publications **22**: 42p.
- Dance, K.W. and Hynes, H.B.N. 1980. Some effects of agricultural land-use on stream insect communities. Environmental Pollution (Series A) **22**: 19-28.
- Davies-Colley, R.J. 1988. Measuring water clarity with a black disk. Limnology and Oceanography **4(1)**: 616-623.

- Davies-Colley, R.J., Hickey, C.W., Quinn, J.M. and Ryan, P.A. 1992. Effects of clay discharges on streams. Hydrobiologia **248**: 215-247.
- Davies-Colley, R.J. and Stroud, M.J. 1995. Water quality degradation by pastoral agriculture in the Whanganui River catchment. Hamilton: National Institute of Water and Atmospheric Research. Consultancy Report DoC050/1.
- Dons, A. 1987. Hydrology and sediment regime of a pasture, native forest, and pine forest catchment in the central North Island of New Zealand, New Zealand. New Zealand Journal of Forestry Science **17**: 161-178.
- Fahey, B.D. and Rowe, L.K. 1992. Land use impacts. In: Mosley M.P., editor. Waters of New Zealand. Wellington: New Zealand Hydrological Society Inc. p 265-284.
- Friberg, N. and Winterbourn, M.J. 1997. Effects of native and exotic forest on stream biota in New Zealand: a colonisation study. Marine and Freshwater Research **48**: 267-275.
- Friberg, N., Winterbourn, M.J., Shearer, K. A. and Larsen, S. E. 1997. Benthic communities of forest streams in the South Island, New Zealand: effects of forest type and location. Archiv für Hydrobiologie **138(3)**: 289-306.
- Graham, A.A., McCaughan, D. J. and McKee, F. S. 1988. Measurement of surface area of stones. Hydrobiologia **157**: 85-87.
- Gregory, S.V. 1983. Plant-herbivore interactions in stream ecosystems. In: Barnes J. R. and Minshall, G. W., editors. Stream ecology, application and testing of general ecological theory. New York: Plenum Press. p 157-190.
- Hanchet, S.M. 1990. Effect of land use on the distribution and abundance of native fish in tributaries of the Waikato River in the Hakarimata Ranges, North Island, New Zealand. New Zealand Journal of Marine and Freshwater Research **24**: 159-171.

- Harding, J.S. 1994. Lotic ecoregions of New Zealand [PhD thesis]. Christchurch, New Zealand: University of Canterbury.
- Harding, J.S. and Winterbourn, M.J. 1995. Effects of contrasting land use on physico-chemical conditions and benthic assemblages of streams in a Canterbury (South Island, New Zealand) river system. New Zealand Journal of Marine and Freshwater Research **29**: 479-492.
- Haro, R.J. and Wiley, M.J. 1992. Secondary consumers and the thermal equilibrium hypothesis: insights from Michigan spring brooks. In: Stanford, J. A. and Simons J. J., editors. Proceedings of the First International Conference on Ground Water Ecology; 26-29 April, 1992; Tampa, Florida. Bethesda, MD: American Water Resources Association. p 179-188.
- Henderson, I. 1996. How reliable are chemical measures of water quality? Observations of conductivity in the Turitea Stream. New Zealand Limnological Society Annual Conference; 2-4 July, 1996; Napier, New Zealand.
- Jowett, I.G. and Richardson, J. 1990. Microhabitat preferences of benthic invertebrates in a New Zealand river and the development of instream flow habitat models for *Deleatidium* sp. New Zealand Journal of Marine and Freshwater Research **24**: 11-22.
- Jowett, I.G., Richardson, J., Biggs, B.J.F., Hickey, C.W. and Quinn, J.M. 1991. Microhabitat preferences of benthic invertebrates and the development of generalised *Deleatidium* sp. habitat suitability curves applied to four New Zealand rivers. New Zealand Journal of Marine and Freshwater Research **25**: 187-199.
- Lay, J.K. and Ward, A.K. 1987. Algal community dynamics in two streams associated with different geological regions in the south-eastern United States. Archiv für Hydrobiologie **108**: 305-324.
- Lenat, D.R. 1984. Agriculture and stream water quality: a biological evaluation of erosion control practices. Environmental Management **8**: 333-344.

- Lenat, D.R. 1988. Water quality assessment of streams using a qualitative collection method for benthic invertebrates. Journal of the North American Benthological Society **7**: 222-233.
- Maasdam, R. and Smith, D.G. 1994. New Zealand's National River Water Quality Network 2. Relationships between physico-chemical data and environmental factors. New Zealand Journal of Marine and Freshwater Research **28**: 37-54.
- McCune, B. and Mefford, M.J. 1995. PC-ORD Multivariate Analysis of Ecological Data, Version 2.0. Oregon: MjM Software Design.
- McFarlane, A.G. 1951. Caddisfly larvae (Trichoptera) of the family Rhyacophilidae. Records of the Canterbury Museum **5**: 267-289.
- Miller, R.B. 1968. Calcium and Lining. Soils of New Zealand, part 2. New Zealand Soil Bureau Bulletin **26(2)**: 72-75.
- Moss, B. 1967a. A spectrophotometric method for the estimation of percentage degradation of chlorophyll's to pheo-pigments in extracts of algae. Limnology and Oceanography **12**: 335-340.
- Moss, B. 1967b. A note on the estimation of chlorophyll *a* in freshwater algal communities. Limnology and Oceanography **12**: 340-342.
- Naiman, R.J., Magnuson, J.J., McKnight, D.M. and Stanford, J.A. 1995. The Freshwater Imperative. Washinton, D.C.: Island Press.
- Newbury, R.W. 1984. Hydrological determinants of aquatic insect habitats. In: Resh, V.H. and Rosenberg, D.M., editors. The Ecology of Aquatic Insects. New York: Praeger. p 323-357.
- Pfankuch, D. J. 1975. Stream reach inventory and channel stability evaluation. U. S. Department of Agriculture Forest Service/Northern Region.

- Poff, N.L. and Allan, J.D. 1995. Functional organisation of stream fish assemblages in relation to hydrological variability. Ecology **76**: 606-627.
- Poff, N.L. and Ward, J.V. 1990. Physical habitat of lotic systems: recovery in the context of historical patterns of spatiotemporal heterogeneity. Environmental Management **14**: 629-645.
- Prat, N. and Ward, J.V. 1994. The tamed river. In: Margalefs, R., editor. *Limnology Now: a Paradigm of Planetary Problems*. Amsterdam: Elsevier Science. p 219-236.
- Quinn, J.M., Cooper, A.B., Davies-Colley, R.J., Rutherford, J.C. and Williamson, R.B. 1994. Land-use effects on New Zealand hill country streams and implications for riparian management. In: Link, G.L. and Naiman, R.J., editors. *Proceedings of the International Workshop on the Ecology and Management of Aquatic-Terrestrial Ecotones*. Seattle: University of Washington.
- Quinn, J.M., Cooper, A.B., Davies-Colley, R.J., Rutherford, J.C. and Williamson, R.B. 1997. Land use effects on habitat, water quality, periphyton, and benthic invertebrates in Waikato, New Zealand, hill-country streams. New Zealand Journal of Marine and Freshwater Research **31**: 579-597.
- Quinn, J.M. and Hickey, C.W. 1990a. Characterisation and classification of benthic invertebrate communities in 88 New Zealand rivers in relation to environmental factors. New Zealand Journal of Marine and Freshwater Research **24**: 387-409.
- Quinn, J.M. and Hickey, C.W. 1990b. Magnitude of effects of substrate particle size, recent flooding and catchment development on benthic invertebrates in 88 New Zealand rivers. New Zealand Journal of Marine and Freshwater Research **24**: 411-427.
- Quinn, J.M., Williamson, R.B., Smith, R.K. and Vickers, M.L. 1992. Effects of riparian grazing and channalisation on streams in Southland, New Zealand. 2. Benthic invertebrates. New Zealand Journal of Marine and Freshwater Research **26**: 259-273.

- Reed, J.L., Campbell, I.C. and Bailey, P.C.E. 1994. The relationship between invertebrate assemblages and available food at forest and pasture sites in three south-eastern Australian streams. Freshwater Biology **32**: 641-650.
- Richards, C., Haro, R.J., Johnson, L.B. and Host, G.E. 1997. Catchment and reach-scale properties as indicators of macroinvertebrate species traits. Freshwater Biology **37**: 219-230.
- Richards, C. and Host, G. 1994. Examining land-use influences on stream habitats and macroinvertebrates: a GIS approach. Water Resources Bulletin **30**: 729-738.
- Richards, C., Johnson, L.B. and Host, G. 1996. Landscape scale influences on stream habitats and biota. Canadian Journal of Fisheries and Aquatic Sciences **53**: 295-311.
- Ryan, P.A. 1991. Environmental effects of sediment on New Zealand streams: a review. New Zealand Journal of Marine and Freshwater Research **25**: 207-221.
- Ryder, G.I. 1989. Experimental studies on the effects of fine sediments on lotic invertebrates [PhD thesis]. Dunedin, New Zealand: University of Otago.
- Scarsbrook, M.R. and Townsend, C. R. 1993. Stream community structure in relation to spatial and temporal variation: a habitat template study of two contrasting New Zealand streams. Freshwater Biology **29**: 395-410.
- Scott, D., White, J.W., Rhodes, D.S. and Koomen, A. 1994. Invertebrate fauna of three streams in relation to land use in Southland, New Zealand. New Zealand Journal of Marine and Freshwater Research **28**: 277-290.
- Simpson, E.H. 1949. Measurement of Diversity. Nature **163**: 688.
- Smith, M.J., Cooper, A.B. and Quinn, J.M. 1993. Land-water interactions research at Whatawhata - the start. Hamilton, New Zealand: National Institute of Water and Atmospheric Research, Ecosystems. Publication 4.

- Stark, J.D. 1985. A macroinvertebrate community index of water quality for stony streams. Water and Soil Miscellaneous Publication **87**: 53p.
- Stark, J.D. 1993. Performance of the Macroinvertebrate Community Index: effects of sampling method, sample replication, water depth, current velocity, and substratum on index values. New Zealand Journal of Marine and Freshwater Research **21**: 463-478.
- Statzner, B., Gore, J.A. and Resh, V.H. 1988. Hydraulic Stream Ecology: Observed Patterns and Potential Applications. Journal of the North American Benthological Society **7**: 307-360.
- Stumm, W. and Morgan, J.J. 1981. Aquatic Chemistry. New York: J. Wiley and Sons.
- Sweeney, B.W. and Vanote, R.L. 1986. Growth and production of a stream stonefly: influences of diet and temperature. Ecology **67**: 1396-1410.
- SYSTAT. 1996. SYSTAT for Windows: Statistics, Version 6 Edition. Evanston, Illinois: SYSTAT Inc.
- Timperley, M.H. 1983. Phosphorus in spring waters of the Tuapo Volcanic zone, North Island, New Zealand. Chemical Geology **38**: 287-306.
- Towns, D.R. and Peters, W.L. 1996. Leptophlebiidae (Insecta: Ephemeroptera). Fauna of New Zealand **36**: 37-50.
- Townsend, C.R. 1989. The patch dynamics concept of stream community ecology. Journal of the North American Benthological Society **8**: 36-50.
- Townsend, C.R., Arbuckle, C.J., Crowl, T.A. and Scarsbrook, M.R. 1997. The relationship between land-use and macroinvertebrate communities in tributaries of the Taieri River, New Zealand: a hierarchically scaled approach. Freshwater Biology **37**: 177-191.

- Townsend, C.R. and Hildrew, A. G. 1994. Species traits in relation to a habitat template for river ecosystems. Freshwater Biology **31**: 265-275.
- Van Nieuwenhuysse, E.E. and LaPerriere, J.D. 1986. Effects of placer gold mining on the primary productivity in sub-arctic streams of Alaska. Water Resources Bulletin **22**: 91-99.
- Ventura, M. and Harper, D. 1996. The impacts of acid precipitation mediated by geology and forestry upon upland stream invertebrate communities. Archive für Hydrobiologia **138(2)**: 161-173.
- Wagener, S.M. and LaPerriere, J.D. 1985. Effects of placer mining on invertebrate communities of interior Alaska streams. Freshwater Biology **4**: 208-214.
- Wiley, M.J., Kohler, S.L. and Seelbach, P.W. 1997. Reconciling landscape and local views of aquatic communities. Freshwater Biology **37**: 133-148.
- Williamson, R.B., Smith, R.K. and Quinn, J.M. 1992. Effects of riparian grazing and channelisation on streams in Southland, New Zealand. 1. Channel form and stability. New Zealand Journal of Marine and Freshwater Research **26**: 241-258.
- Winterbourn, M.J. 1973. A guide to the freshwater Mollusca of New Zealand. Tuatara **20**: 141-159.
- Winterbourn, M.J. 1986. Effects of land development on benthic stream communities. New Zealand Agricultural Science **20**: 115-118.
- Winterbourn, M.J. and Gregson, K.L.W. 1989. Guide to the Aquatic Insects of New Zealand. Bulletin of the Entomological Society of New Zealand 9, 96 p.
- Winterbourn, M. J., Rounick, J.S. and Cowie, B. 1981. Are New Zealand stream ecosystems really different? New Zealand Journal of Marine and Freshwater Research **15**: 321-328.

Chapter Three

Longitudinal changes in macroinvertebrate community structure along a large New Zealand river

ABSTRACT

Both artificial substrates and kick sampling were used to sample macroinvertebrate communities and explore the affects of physicochemical changes along a large New Zealand river. Taxonomic diversity and numbers of pollution sensitive taxa decreased along five sampling sites down the Whanganui River. This was correlated to reduced periphyton biomass and increased suspenoids. *Potamopyrgus antipodarium* was highly abundant at site three and five as both sites had similar macrohabitat suitable for this species. The influence of macrohabitat was apparent from similarities in community structure between site three and five and differences in community structure between sampling techniques.

Keywords: benthic macroinvertebrate communities, macrohabitat, artificial substrate, longitudinal, sampling precision, conductivity, water clarity, periphyton, Whanganui.

INTRODUCTION

Land development practices result in changes in the structure and function of running water ecosystems through catchment modification (Winterbourn, 1986) significantly altering physicochemical parameters and algal biomass (Quinn and Hickey, 1990b). Invertebrate assemblages are integrally linked to in stream physical and chemical characteristics and for this reason are useful environmental indicators (Stark, 1993).

In developed countries the longitudinal ecological pattern of a large river is often strongly influenced by water pollution and flow regulation (Bournaud *et al.*, 1996; Zamora-Muñoz and Alba-Tercedor, 1996). Longitudinal changes of macroinvertebrate fauna may be closely related to the immediate environment or due to other causes such as physicochemical inputs from tributaries. Much of the work concerning water quality in New Zealand has been conducted on small to medium sized rivers, typically with stony substrate. As large rivers usually flow through lowland areas, they are susceptible to anthropogenic influence. However, few studies have been conducted on the effects of physicochemistry on macroinvertebrate communities in large non-cobble dominated waterways like the Whanganui River. Sampling techniques used in shallow stony streams are not always appropriate for large rivers. Consequently, artificial substrates

such as multiplate samplers, are used for sampling (see review by Rosenberg and Resh, 1982), particularly reaches that are deep and have boulder or bedrock substrate (Boothroyd and Dickie, 1989), conditions that commonly occur in the middle and lower reaches of the Whanganui River.

It has been demonstrated previously in Chapter One that catchment land use combined with substrate geology influences invertebrate community composition in smaller headwaters of the Whanganui River catchment. The aim of this study is to explore the effects of water quality on macroinvertebrate communities in the main stem of the Whanganui River and to compare the efficiency of kick net and artificial substrate sampling techniques in doing this.

STUDY SITES

Five sites were sampled along the Whanganui River between Taumaranui and Kaiwhaiki (Fig. 3.1). Site five was located 14 km upstream from the river mouth and site numbers decreased in order upstream. Altitude ranged from < 3 m to 160 m a.s.l.

Site one was situated 8 km upstream of Taumaranui where the river flows through land developed for pastoral agriculture with water extraction for hydroelectric power generation occurring on both the Whanganui and Whakapapa tributaries. However, these headwaters are partially snow fed arising at high altitude on Mt Tongariro and Mt Ruapehu in Tongariro National Park and the river is at its most 'natural' state at this site. The predominant geology type is of volcanic origin. Channel characteristics in this section of the river consist mainly of fast flowing riffle over boulder/cobble substrate.

Site two was approximately 300 m below the confluence of the Retaruke River. The main tributaries to enter the Whanganui above this point are the Ongarue, Ohura and the Retaruke. The Ongarue is a large tributary and the geology of its headwaters consist principally of the pumice soils of the Hauhungaroa range which are well forested. The Ohura river drains land predominantly in pastoral agriculture with soils formed from siltstone which are very prone to erosion. The Retaruke headwaters are located in indigenous bush with substantial areas downstream draining agriculturally developed land.

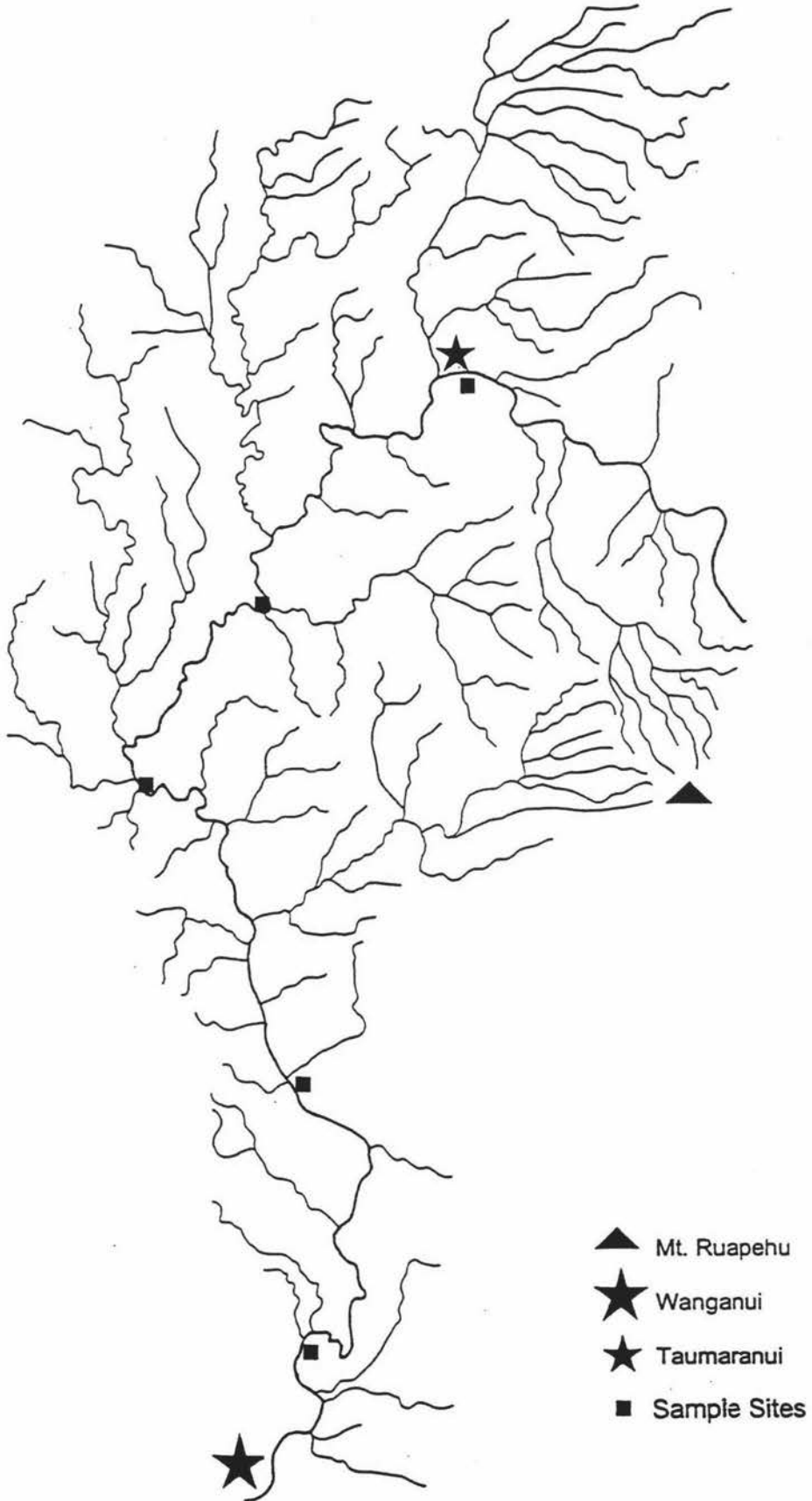


Figure 3.1. Location of the five sites along the main stem Whanganui River where artificial substrate samples and kick samples were collected between the 13 and 15 of May, 1997.

At site two geology formed by Tertiary sediments was influential in creating many deep sections with mudstone bedrock substrate mixed with riffle sections of large cobble substrate.

Site three was situated downstream of the Whangamomona river confluence. Both the Whangamomona and the Tangarakau Rivers, which enter the Whanganui further upstream, drain catchments comprising of sandstone and jointed mudstone. The banks of the Whanganui at this point are in many places steep and high restricting light entry. Riffles are fewer as the channel is confined and deep.

Macrohabitat at site four was similar to site two except levels of deposited sediment were greater. Macrohabitat is defined here as factors such as velocity and overall substrate qualities characteristic to a given reach. The last major tributary, the Manganuioteao, enters approximately 16 km upstream. This tributary drains the western indigenous forest/grassland slopes of Mt Ruapehu so water quality is higher than for other tributaries. No additional major tributaries enter between sites four and five. Substrate at site five consisted mainly of deposited silt and mudstone bedrock. Gradient and velocity were lower here than the upstream sites. Tidal influence and salt water penetration can occur at site five depending on river levels.

METHODS

Sampling protocol

Invertebrates were collected between the 13 and 15 of May, 1997. At each of the five sites, three 60 second kick samples were taken using a 250 μ m mesh kick net as the sampler moved progressively upstream. Two artificial substrates were collected from each site that had been placed in the river 6 weeks prior to their removal on the above date.

Samples were sieved through a 250 μ m Endecott sieve and animals identified and enumerated with a 40 \times microscope using available keys (McFarlane, 1951; Winterbourn, 1973; Chapman and Lewis, 1976; Cowley, 1978; Winterbourn and Gregson, 1989; Towns and Peters, 1996). If taxa could not be identified to species they were assigned to apparent morphospecies.

Measurement of physicochemical parameters at each site were undertaken at the same time as the collection of invertebrate samples. Conductivity was measured using an Orion 122 portable conductivity meter and water samples were taken back to the lab for pH analysis using an Orion 250A pH meter. Black disk measurements were used to measure water clarity (Davies-Colley, 1988). Current speed at points where artificial substrates and kick samples were collected was categorised as being of low, medium or high velocity (Table 3.1). Depth of artificial substrates at the time of placement ranged from 60 cm to 100 cm.

Algal biomass was assessed by extracting photosynthetic pigments (chlorophyll *a* and phaeophytin) in 90% acetone, for 24 hours at 5 °C, from five small cobbles (mean circumference = 10 cm) collected on each sampling occasion. Absorption was measured on a Jenway 6105 Spectrophotometer. Total pigment concentration was calculated using the method of Moss (1967 a, b) and corrected for stone surface area (Graham *et al.*, 1988).

Artificial substrates

Artificial substrates were removed using a dip net placed around them before disturbance and transferred to a sorting tray. They were then dismantled with all animals removed using a scrubbing brush and preserved in 70% alcohol. Artificial substrates based on the multiplate design of Hester and Dendy (1962), comprised of 14 square perspex plates (80 mm × 80 mm) separated by 20 mm × 20 mm square spacers. Plates were mounted at the end of a 1 metre steel rod with eight upper spaces of 3 mm and five lower spaces of 6 mm. Plates were constructed of 3 mm clear perspex uniformly roughened from belt sanding of both sides using 60 grade sandpaper. The total area available for macroinvertebrate colonisation on each substrate was 0.187 m². Substrates were stabilised on the river bed using weighted grapnel type anchors with four protruding 20 cm steel rods attached to the steel shaft of each artificial substrate. Both artificial substrate replicates at each site were attached to stable submersed objects such as imbedded logs or very large boulders using 7 mm braided polypropylene rope. Artificial substrates were not placed at or near the surface because of the risk of snagging by large floating debris and it has been shown that multiplates located nearer the benthos display a greater consistency of macroinvertebrate assemblage structure among sites (Mason *et al.*, 1973; Boothroyd and Dickie, 1989).

Fixed surface placements for artificial substrates were unsuitable because highly fluctuating water levels can leave them exposed in the lower Whanganui River. Artificial substrate placement was influenced by the logistical constraints of suitable anchorage points and localities.

Data analysis

Longitudinal trends in physicochemical and biological variables were tested for significance with Spearman rank correlations. Spearman rank correlations were also calculated between physicochemical variables and six indices of community structure. Variation of indices between sampling techniques was investigated using paired t-tests and Pearson correlations. Community structure was analysed with detrended correspondence analysis (DECORANA), multi-response permutation procedure (MRPP) and two-way indicator species analysis (TWINSPAN) using the PC-ORD statistical package (McCune and Mefford, 1995). Pseudo-species cut levels were 1, 10, 100 and 1000 per sample.

Biological indices used consisted of the MCI (Stark, 1985), QMCI (Stark, 1993), EPT (Lenat, 1988), Simpson's evenness (Simpson, 1949), Berger-Parker dominance (Berger and Parker, 1970) and Margelefs (Clifford and Stevenson, 1975). The formula for these indices are contained in Chapter Two. Data was transformed where appropriate using $\log_{10}(x+1)$ transformations to remove heterogeneity of variance.

RESULTS

Physicochemical variables

Water clarity was the only physicochemical variable to show a significant longitudinal trend ($r_s = 0.9$, $P = 0.04$) (Fig. 3.2a). Levels of chlorophyll *a* also decreased downstream (Fig. 3.2b) but a major drop occurred at site three ($r_s = 0.7$, $P > 0.05$). Conductivity (Fig. 3.2c) and pH (Fig. 3.2d) varied widely ranging between 127 to 178 μScm^{-2} , and 7.08 to 7.38, respectively.

Community structure

EPT scores showed significant longitudinal change in the artificial substrate samples ($r_s = 1$, $P = 0.00$) with EPT scores and the MCI showing significant longitudinal change in the kick samples (both $r_s = 0.9$, $P = 0.04$).

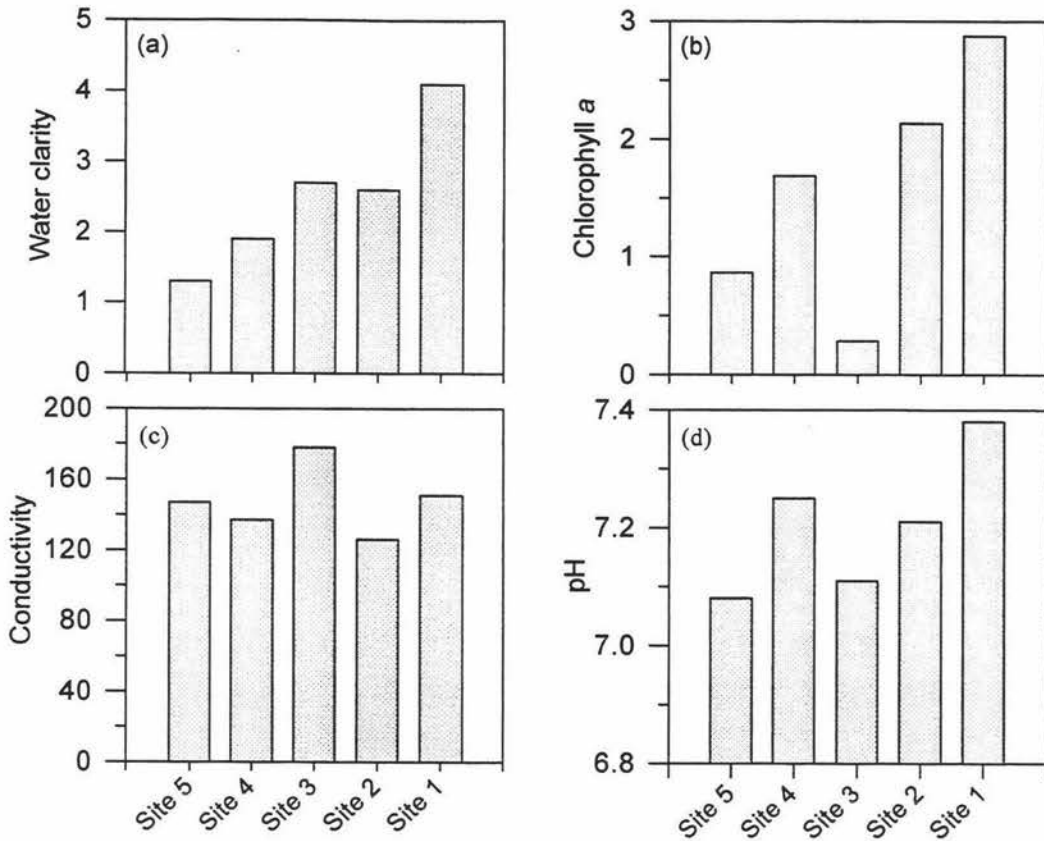


Figure 3.2. (a) Water clarity, (b) chlorophyll *a* ($\mu\text{g cm}^{-2}$), (c) conductivity (μScm^{-2}), and (d) pH measured at five sites along the Whanganui River, May, 1997.

Margalefs index from artificial substrate samples (Fig. 3.3a) decreased longitudinally downstream ($r_s = 1$, $P = 0.00$) although taxonomic richness varied widely among sites and between sampling techniques with the lowest diversity occurring at site five (Fig. 3.3b). Diversity of the communities collected with artificial substrates only exceeded that of samples collected using kick sampling at site three. Macrohabitat at site three resembled that of site one having deep, low current conditions combined with large mudstone boulders, bedrock and deposited silt substrate. Both Margalefs index and taxonomic richness from artificial substrate samples were significantly correlated with water clarity ($r_s = 0.9$, $P = 0.04$ and $r_s = 1$, $P = 0.00$, respectively).

EPT scores for both sampling techniques decreased longitudinally downstream ($r_s = 1$, $P = 0.00$) (Fig. 3.3c). Artificial substrate EPT scores were correlated with water clarity while EPT scores for kick samples correlated with pH and chlorophyll *a* (both $r_s = 0.9$, $P = 0.04$). Total abundance of individuals was similar among kick samples but varied widely among artificial substrates (Fig. 3.3d). The Berger-Parker dominance and

Simpson's evenness indices (Fig. 3.3e) were similar with high values for these at sites three and five. The Berger-Parker dominance and Simpson's evenness indices were correlated between sampling techniques ($r = -0.72$, $P = 0.01$ and $r = -0.68$, $P = 0.04$) with high numbers of the mollusc *Potamopyrgus antipodarium* and an overall lack in taxonomic diversity at sites three and five responsible for high values for these indices (Fig. 3.4). MCI (Fig. 3.3f) and QMCI scores were similar and lowest at site five followed by site three for both artificial substrates and kick samples. MCI values for kick samples decreased significantly downstream ($r_s = 0.9$, $P = 0.04$), and were positively correlated to chlorophyll *a* ($r_s = 0.9$, $P = 0.04$) and pH ($r_s = 0.9$, $P = 0.04$). Kick sample QMCI's were also correlated to pH ($r_s = 0.9$, $P = 0.04$).

At site three over 90% of the individuals present were molluscs belonging to the genus *Potamopyrgus* with the crustacean shrimp *Paratya* common in the kick samples (Fig. 3.4). *Potamopyrgus antipodarium* was clearly the most abundant individual at site three, comparable to site five. However, Mollusca accounted for only one fifth of the individuals at site two and were not present at site one. Trichoptera increased significantly above site three and accounted for over 50% of the individuals collected with artificial substrates at site one. The net spinning caddis fly, *Aoteapsyche colonica* was the most common taxa on artificial substrates at site one and two. A high proportion of Ephemeroptera (predominantly *Coloburiscus humeralis*) were collected from the site one kick samples. Graphing the DECORANA ordination axis 1 against axis 2 displayed grouping between sample types within a site and a gradient of sites along the first axis (Fig. 3.5) starting from site five to site three, site four, site two and lastly site one.

Similarities between site five and site three were evident as these samples were closely grouped on all three ordination axis. Two-way indicator species analysis (TWINSpan) classified both site five sample types, site four artificial substrate and the site three kick samples together. The remaining groups were characterised by the family Elmidae (Coleoptera) and consisted of two remaining groups. One contained all sample types from sites one and two together. The other contained the artificial substrate samples at site three and the kick sample at site four. Multi-response permutation procedure (MRPP), found invertebrate communities to differ significantly among sites for kick samples ($r = 0.33$, $P = 0.001$) and artificial substrates ($r = 0.44$, $P = 0.003$) but not between sampling techniques ($r = -0.003$, $P = 0.42$).

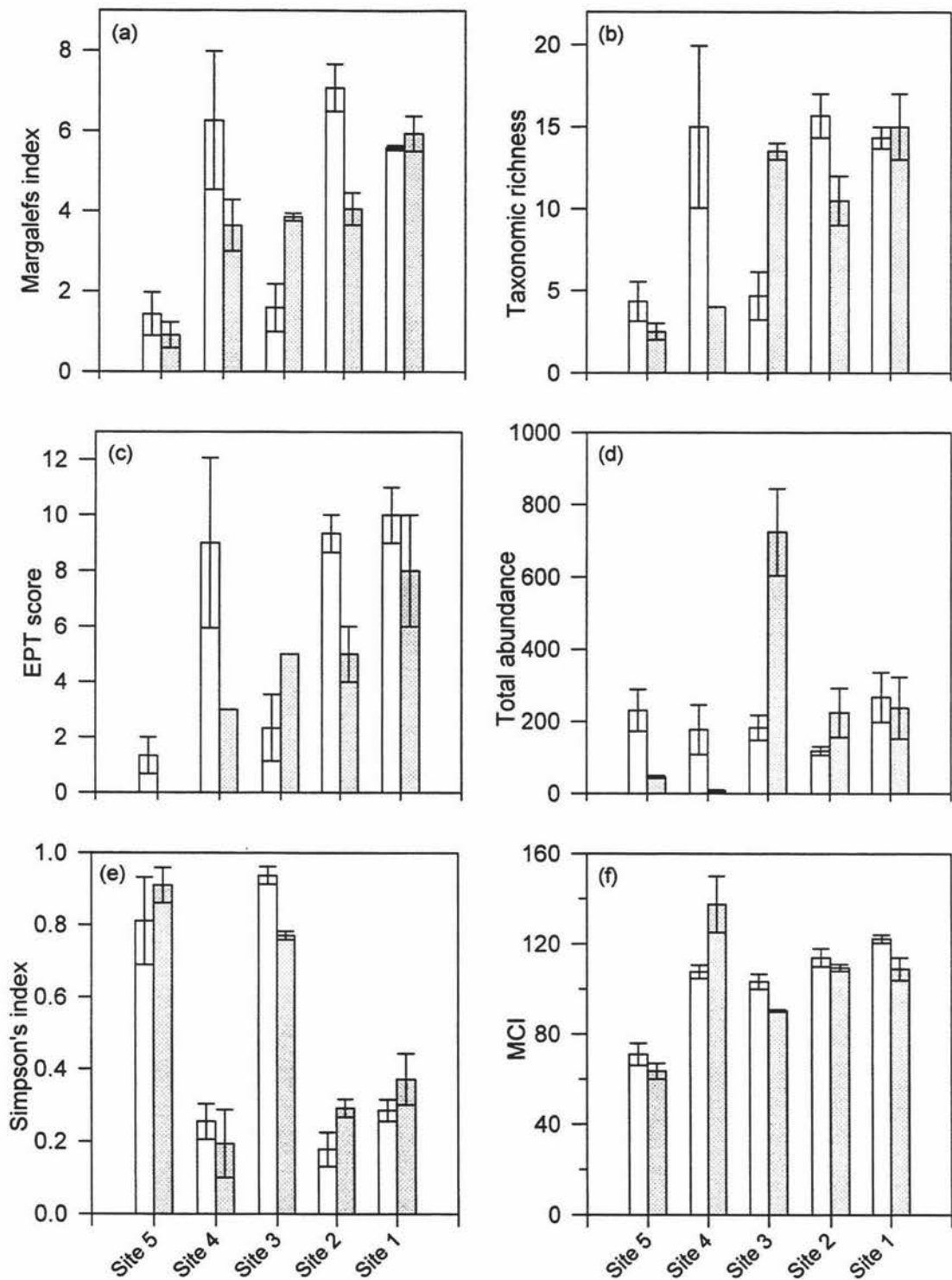


Figure 3.3. Mean (± 1 S.E.) (a) Margalefs index, (b) taxonomic richness, (c) EPT scores, (d) total abundance, (e) Simpson's index of evenness, and (f) MCI values measured at five sites along the Whanganui River, May, 1997. White fills represent kick samples and shaded fills represent artificial substrate samples.

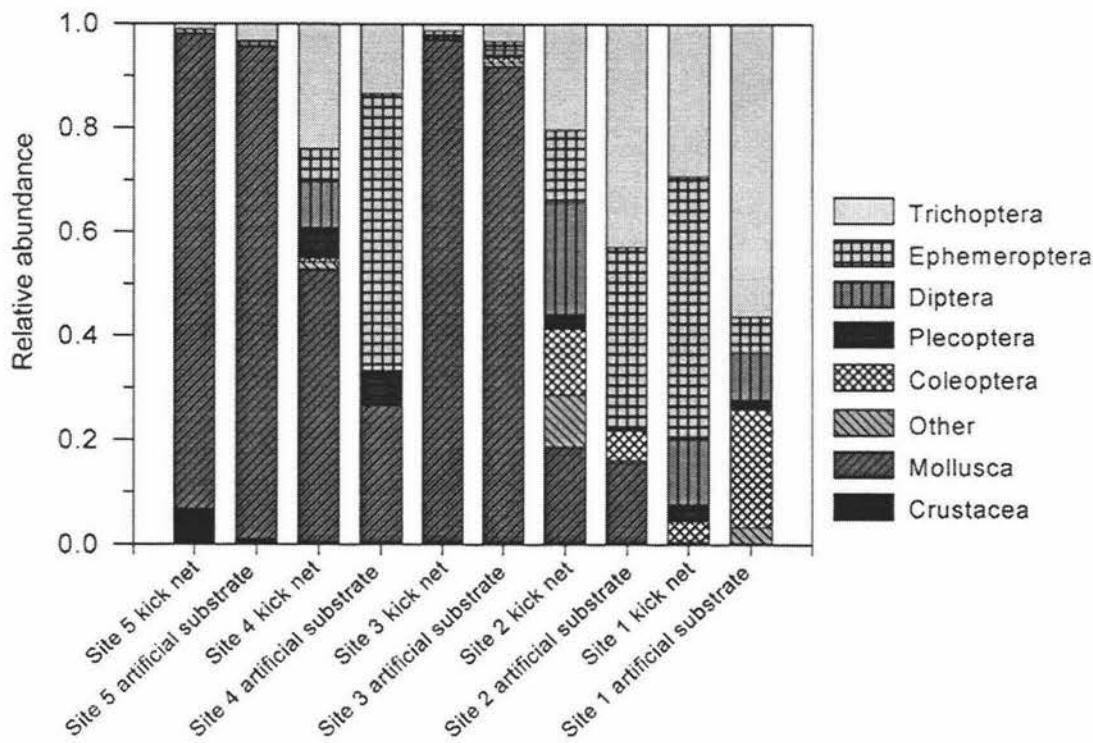


Figure 3.4. Mean relative abundance of individuals from eight taxonomic groups at five sites measured using both artificial substrate and kick sampling along the Whanganui River, May, 1997.

Table 3.1. Velocity and substrate categories for five sites along the Whanganui River, May, 1997.

Site	Artificial substrates	Kick samples	
	<i>Velocity</i>	<i>Substrate composition</i>	<i>Velocity</i>
1	medium	boulder/cobble	high
2	low	boulder/cobble and bedrock	high
3	medium	large boulder/bedrock	medium
4	low	boulder/cobble and bedrock	high
5	medium	bedrock/large boulder and silt	medium

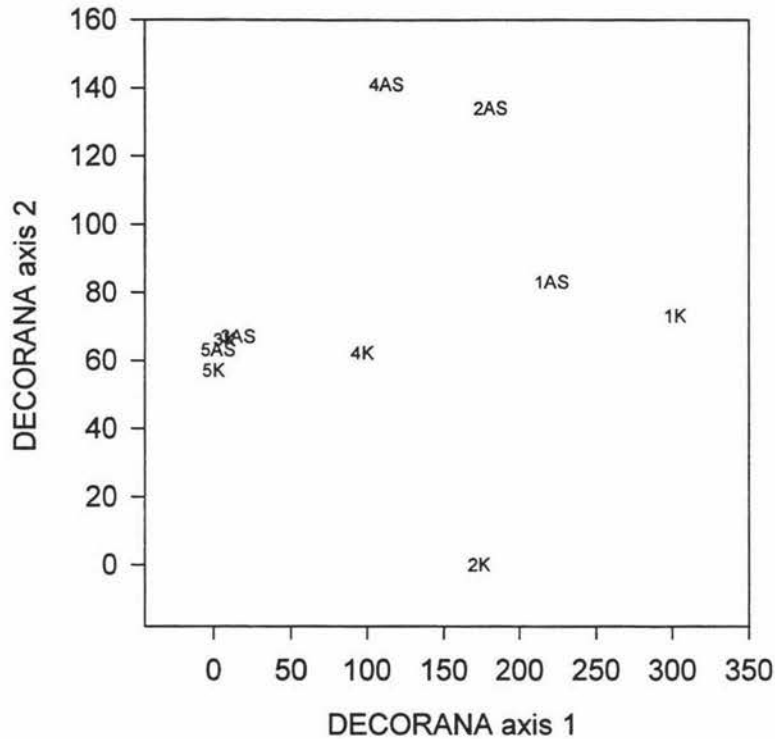


Figure 3.5. DECORANA analysis of community structure measured using both (AS) artificial substrates and (K) kick sampling along the Whanganui River, May, 1996.

DISCUSSION

Over the course of 130 km between site one and site five, variation in biological and physicochemical factors were observed. Longitudinal trends in water clarity, the MCI, Margalefs index and EPT score were observed but (excluding EPT score) were dependant on the sampling technique used. Macroinvertebrate community structure was similar between site three and five as shown by the mean relative abundance of main taxonomic groups, community dominance and evenness, and ordination. No significant difference in macroinvertebrate community structure between sampling techniques was found with multi-response permutation procedure (MRPP) or paired t-tests. However, longitudinal trends in biological measurements as well as their relationship with physicochemical measurements varied between sampling techniques.

Varying conductivity levels reflect the impact of land use practices (Close and Davies-Colley, 1990; Collier, 1995) occurring in the respective tributaries that discharge into

the Whanganui River. Contributions either dilute or increase upstream conductivity rather than having a cumulative effect as was found with water clarity. Water clarity decreased downstream representing an accumulation of deposited silt and suspended material in the river system. Shading from increased turbidity may be responsible for the overall decrease in periphyton biomass (Van Nieuwenhuysen and LaPerriere, 1986; Ryan, 1991; Davies-Colley *et al.*, 1992). The large drop in periphyton biomass at site three may be a result of decreased light access (e.g., Quinn *et al.*, 1997) caused by the presence of stream banks greater than 15 m high. Periphyton biomass may also be reduced by high numbers of the algal grazing gastropod *Potamopyrgus antipodarium* as has been shown elsewhere for grazing invertebrates (Winterbourn and Fegley, 1989; Rosemond, 1993; Kjeldsen, 1996).

Higher periphyton biomass positively influenced the number of pollution sensitive taxa as indicated by the MCI and numbers of Ephemeroptera, Trichoptera and Plecoptera (EPT). Other studies have found *Deleatidium* spp. to increase with periphyton biomass (e.g., Suckling, 1982; Jowett and Richardson, 1990) where growth is not overly dense or filamentous (Davies-Colley *et al.*, 1992) as was found in this study. The relationship between community evenness and dominance (as determined from kick samples) with conductivity could represent the positive response of *Potamopyrgus antipodarium* to increased organic enrichment associated with pastoral agriculture (Scott *et al.*, 1994; Harding and Winterbourn, 1995). However no relationship was found between periphyton biomass and dominance, evenness or conductivity. Growth rate rather than biomass may be a more appropriate measure if high algal grazing is occurring at site three.

Longitudinal change in community structure was shown by multivariate analysis as well as significant downstream decreases in overall taxa and EPT diversity, and pollution sensitive species. Species diversity and EPT numbers collected with artificial substrates were negatively correlated with turbidity and suspended solids as inferred from water clarity (Davies-Colley and Stroud, 1995). Negative impacts on diversity associated with increased suspensoids (e.g., Quinn *et al.*, 1992) are likely to result from substrate clogging (Ryder, 1989) and decreased primary production (Davies-Colley *et al.*, 1992).

Similarities in community structure between site three and five are caused primarily by high relative abundance of *Potamopyrgus antipodarium*. Both sites had similar

substrate (very large boulders and bedrock) in areas of low to moderate velocity which provided a suitably stable macrohabitat for *P. antipodarium* to proliferate (Jowett and Duncan, 1990; Harding and Winterbourn, 1995). Increased diversity and changes in taxonomic composition from site three to site four kick samples may result from a combination of less organically enriched water from the Manganuioteao and different macrohabitat characteristics (e.g., Bournaud *et al.*, 1996) which play an important part in shaping community structure (Williams, 1980; Poff and Ward, 1990; Quinn and Hickey, 1990b; Collier, 1995). Two examples of macrohabitat differences from site three to four that are associated with increased diversity include a shift from bedrock/boulder to large cobble substrate (Quinn and Hickey, 1990b) and increased velocity benefiting taxa that have higher oxygen requirements such as Ephemeroptera, Plecoptera and Trichoptera (Nebeker, 1972; Wiley and Kohler, 1980; Williams *et al.*, 1987) which were more abundant at site four.

Part of the life cycle of the Atyid shrimp *Paratya* spp. occurs in or near the sea (Chapman and Lewis, 1976; Taranaki Catchment Commission, 1985; Quinn and Hickey, 1990a). Tidal influence occurs at site five which explains the high numbers of *Paratya* spp. found at this site. Increases in Ephemeroptera and Trichoptera at sites one and two and the absence of *P. antipodarium* at site one are most probably the product of an increase in water quality and velocity (e.g., Lenat, 1988; Quinn and Hickey, 1990b; Harding and Winterbourn, 1995).

At the top two sites, Trichoptera, principally the net spinner *Aoteapsyche colonica*, preferred the artificial substrates as has been found for similar species (e.g., Benfield *et al.*, 1974; Way *et al.*, 1995). At site three the mean abundance of *P. antipodarium* collected on the artificial substrates was three times of that collected with kick sampling where a lot of bedrock made kick sampling difficult. This is an example where artificial substrates were able to collect a greater variety and abundance of taxa because they provided a larger surface area and more suitable macrohabitat for many taxa (e.g., Dickson and Cairns, 1972; Minshall and Minshall, 1977; Taranaki Catchment Commission, 1985; Boothroyd and Dickie, 1989).

It is hard to determine whether six weeks are long enough for adequate colonisation of artificial substrates (Pearson and Jones, 1975; Fredeen and Spurr, 1978). Consideration for the accumulation of periphyton and detritus must also be taken into account as they

greatly influence the colonising community (Roby *et al.*, 1978; Winterbourn *et al.*, 1981; Rounick and Winterbourn, 1983; Winterbourn *et al.*, 1984; Boothroyd and Dickie, 1987). Initially it may appear that the maximum quantity of periphyton biomass and detritus present on an artificial substrate would be equivalent or less than that found on the nearby benthos. However, a well placed artificial substrate in moderate to highly disturbed environments will remain stationary during disturbance events that would dislodge or scour equivalent sized natural substrate, thus allowing artificial substrates to build up higher levels of periphyton and detritus (e.g., Canton and Chadwick, 1983). Therefore invertebrate communities may continue to change well after six weeks as a result of the creation of new niches (Meir *et al.*, 1979). These changes may include a decrease in taxonomic richness and an increase in taxonomic domination (Boothroyd and Dickie, 1989). Periphyton growth and detritus on the artificial substrates at sites one, two, and three was noticeably higher than the surrounding benthos which lends support to this theory.

A potential advantage of artificial substrates is the increase of sampling precision between sites (Shaw and Minshall, 1980; Lamberti and Resh, 1985). To maximise precision between artificial substrates their site placement must be similar in terms of flow, depth and light regimes (Rosenberg and Resh, 1982). However, due to the constraints of suitable anchorage sites they were often placed in areas subject to lower velocity than where the kick samples were collected. The influence of velocity was apparent between sampling techniques where kick samples at sites one, two and four were all taken from a riffle and had greater numbers of Ephemeroptera, Plecoptera and Trichoptera, taxa which often have greater oxygen requirements (Nebeker, 1972; Williams *et al.*, 1987; Wiley *et al.*, 1997). These taxa were the main component missing from most of the artificial substrate samples where velocity was lower.

The combination of both sampling techniques suggest a longitudinal decrease in taxonomic diversity and pollution sensitive taxa. This decrease is influenced by a reduction in periphyton biomass and an increase in suspensoids associated with pastoral land use. The influence of macrohabitat is evident from differences between sampling techniques, and sites three and five. This influence should be considered when assessing the effect of physicochemistry on changes in community structure and choosing macroinvertebrate sampling techniques for studies of water quality.

REFERENCES

- Benfield, E.F., Hendricks, A.C. and Cairns, J. 1974. Proficiencies of two artificial substrates in collecting stream macroinvertebrates. Hydrobiologia **45**: 431-440.
- Berger, W.H. and Parker, F.L. 1970. Diversity of plankton Foraminifera in deep sea sediments. Science **168**: 1345-1347.
- Boothroyd, I.K.G. and Dickie, B.N. 1989. Macroinvertebrate colonisation of perspex artificial substrates for use in biomonitoring studies. New Zealand Journal of Marine and Freshwater Research **23**: 467-478.
- Bournaud, M., Cellot, B., Richoux, P. and Berrahou, A. 1996. Macroinvertebrate community structure and environmental characteristics along a large river: congruity of patterns for identification to species or family. Journal of the North American Benthological Society **15(2)**: 232-253.
- Canton, S.P. and Chadwick, J.W. 1983. Aquatic and insect communities of natural and artificial substrates in a montane stream. Journal of Freshwater Ecology **2**: 153-158.
- Chapman, A. and Lewis, M. 1976. An introduction to the freshwater Crustacea of New Zealand. Auckland: Collins.
- Clifford, H.T. and Stephenson, W. 1975. An Introduction to Numerical Classification. New York: Academic press.
- Close, M.E. and Davies-Colley, R.J. 1990. Base flow water chemistry in New Zealand rivers. 2. Influence of environmental factors. New Zealand Journal of Marine and Freshwater Research **24**: 343-356.
- Collier, K.J. 1995. Environmental factors affecting the taxonomic composition of aquatic communities in lowland waterways of Northland, New Zealand. New Zealand Journal of Marine and Freshwater Research **29**: 453-465.

- Cowley, D.R. 1978. Studies on the larvae of New Zealand Trichoptera. New Zealand Journal of Zoology **5**: 639-750.
- Davies-Colley, R.J. 1988. Measuring water clarity with a black disk. Limnology and Oceanography **4**(1): 616-623.
- Davies-Colley, R.J., Hickey, C.W., Quinn, J.M. and Ryan, P.A. 1992. Effects of clay discharges on streams. Hydrobiologia **248**: 215-247.
- Davies-Colley, R.J. and Stroud, M.J. 1995. Water quality degradation by pastoral agriculture in the Whanganui River catchment. Hamilton: National Institute of Water and Atmospheric Research. Consultancy Report DoC050/1.
- Dickson, K.L. and Cairns, J. 1972. The relationship of macroinvertebrate communities collected by floating artificial substrates to the MacArthur-Wilson equilibrium model. American Midland Naturalist **88**: 68-75.
- Fredeen, F.J.H. and Spurr, D.T. 1978. Collecting semi-quantative samples of the Black Fly larvae (Diptera: Simuliidae) and other aquatic insects from large rivers with the aid of artificial substrates. Entomology **14**: 411-431.
- Graham, A.A., McCaughan, D.J. and McKee, F. S. 1988. Measurement of surface area of stones. Hydrobiologia **157**: 85-87.
- Harding, J.S. and Winterbourn, M.J. 1995. Effects of contrasting land use on physico-chemical conditions and benthic assemblages of streams in a Canterbury (South Island, New Zealand) river system. New Zealand Journal of Marine and Freshwater Research **29**: 479-492.
- Hester, F.E. and Dendy, J.S. 1962. A multi-plate sampler for aquatic macroinvertebrates. Transactions of the American Fisheries Society **91**: 420-421.
- Jowett, I.J. and Duncan, M.J. 1990. Flow variability in New Zealand rivers and its relationship to instream habitat and biota. New Zealand Journal of Marine and Freshwater Research **24**: 305-317.

- Jowett, I.G. and Richardson, J. 1990. Microhabitat preferences of benthic invertebrates in a New Zealand river and the development of instream flow habitat models for *Deleatidium* sp. New Zealand Journal of Marine and Freshwater Research **24**: 11-22.
- Kjeldsen, K. 1996. Regulation of algal biomass in a small lowland stream: field experiments on the role of invertebrate grazing, phosphorus and irradiance. Freshwater Biology **36**: 535-546.
- Lamberti, G.A. and Resh, V.H. 1985. Comparability of introduced tiles and natural substrates for sampling lotic bacteria, algae and macroinvertebrates. Freshwater Biology **15**: 21-30.
- Lenat, D.R. 1988. Water quality assessment of streams using a qualitative collection method for benthic invertebrates. Journal of the North American Benthological Society **7**: 222-233.
- Mason, W.T., Weber, C.I., Lewis, P.A. and Julian, P.C. 1973. Factors influencing the performance of basket and multiplate macroinvertebrate samplers. Freshwater Biology **3**: 409-436.
- McCune, B. and Mefford, M.J. 1995. PC-ORD Multivariate Analysis of Ecological Data, Version 2.0. Oregon: MjM Software Design.
- McFarlane, A.G. 1951. Caddisfly larvae (Trichoptera) of the family Rhyacophilidae. Records of the Canterbury Museum **5**: 267-289.
- Mier, P.G., Penrose, D.L. and Polak, L. 1979. The rate of colonisation of macroinvertebrates on artificial substrate samplers. Freshwater Biology **9**: 381-392.
- Minshall, G.W. and Minshall, J.N. 1977. Macrodistribution of macroinvertebrates in a Rocky Mountain (U.S.A.) stream. Hydrobiologia **55**: 231-249.

- Moss, B. 1967a. A spectrophotometric method for the estimation of percentage degradation of chlorophyll's to pheo-pigments in extracts of algae. Limnology and Oceanography **12**: 335-340.
- Moss, B. 1967b. A note on the estimation of chlorophyll *a* in freshwater algal communities. Limnology and Oceanography **12**: 340-342.
- Nebeker, A.V. 1972. The effect of low oxygen concentration on survival and emergence of aquatic insects. Transactions of the American Fisheries Society **101**: 675-679.
- Pearson, R.G. and Jones, N.V. 1975. The colonisation of artificial substrata by stream macroinvertebrates. Progressive Water Technology **7**: 497-504.
- Poff, N.L. and Ward, J.V. 1990. Physical habitat of lotic systems: recovery in the context of historical patterns of spatiotemporal heterogeneity. Environmental Management **14**: 629-645.
- Quinn, J.M., Cooper, A.B., Stroud, M.J. and Burrell, G.P. 1997. Shade effects on stream periphyton and invertebrates: an experiment in streamside channels. New Zealand Journal of Marine and Freshwater Research **31**: 665-683.
- Quinn, J.M. and Hickey, C.W. 1990a. Characterisation and classification of benthic invertebrate communities in 88 New Zealand rivers in relation to environmental factors. New Zealand Journal of Marine and Freshwater Research **24**: 387-409.
- Quinn, J.M. and Hickey, C.W. 1990b. Magnitude of effects of substrate particle size, recent flooding and catchment development on benthic invertebrates in 88 New Zealand rivers. New Zealand Journal of Marine and Freshwater Research **24**: 411-427.
- Quinn, J.M., Williamson, R.B., Smith, R.K. and Vickers, M.L. 1992. Effects of riparian grazing and channalisation on streams in Southland, New Zealand. 2. Benthic invertebrates. New Zealand Journal of Marine and Freshwater Research **26**: 259-273.

- Roby, K.B., Newbold, J.D. and Erman, D.C. 1978. Effectiveness of an artificial substrate for sampling macroinvertebrates in small streams. Freshwater Biology **8**: 1-8.
- Rosemond, A.D. 1993. Interactions among irradiance, nutrients and herbivore constraints on a stream algal community. Oecologia **94**: 585-594.
- Rosenberg, D.M. and Resh, V.H. 1982. The use of artificial substrates in the study of freshwater benthic invertebrates. In: Cairns Jr J., editors. Artificial substrates. Michigan, U.S.A: Ann Arbor Science. p 175-236.
- Rounik, J.S. and Winterbourn, M.J. 1983. The formation, structure and utilisation of stone surface organic layers in two New Zealand streams. Freshwater Biology **13**: 57-72.
- Ryan, P.A. 1991. Environmental effects of sediment on New Zealand streams: a review. New Zealand Journal of Marine and Freshwater Research **25**: 207-221.
- Ryder, G.I. 1989. Experimental studies on the effects of fine sediments on lotic invertebrates [PhD thesis]. Dunedin, New Zealand: University of Otago.
- Scott, D., White, J.W., Rhodes, D.S. and Koomen, A. 1994. Invertebrate fauna of three streams in relation to land use in Southland, New Zealand. New Zealand Journal of Marine and Freshwater Research **28**: 277-290.
- Shaw, D.W. and Minshall, G.W. 1980. Colonisation of an introduced substrate by stream macroinvertebrates. Oikos **34**: 259-271.
- Simpson, E.H. 1949. Measurement of Diversity. Nature **163**: 688.
- Stark, J.D. 1985. A macroinvertebrate community index of water quality for stony streams. Water and Soil Miscellaneous Publication **87**: 53p.

- Stark, J.D. 1993. Performance of the Macroinvertebrate Community Index: effects of sampling method, sample replication, water depth, current velocity, and substratum on index values. New Zealand Journal of Marine and Freshwater Research **21**: 463-478.
- Suckling, D.M. 1982. Organic wastewater effects on benthic invertebrates in the Manawatu River. New Zealand Journal of Marine and Freshwater Research **16**: 263-270.
- Taranaki Catchment Commission. 1985. A biological survey of the lower Patea river. Stratford, New Zealand: Taranaki Catchment Commission. Technical Report 84-4.
- Towns, D.R. and Peters, W.L. 1996. Leptophlebiidae (Insecta: Ephemeroptera). Fauna of New Zealand **36**: 37-50.
- Van Nieuwenhuysse, E.E. and LaPerriere, J.D. 1986. Effects of placer gold mining on the primary productivity in sub-arctic streams of Alaska. Water Resources Bulletin **22**: 91-99.
- Way, C.M., Burky, A.J., Bingham, C.R. and Miller, A.C. 1995. Substrate roughness, velocity refuges, and macroinvertebrate abundance on artificial substrates in the lower Mississippi River. Journal of the North American Benthological Society **14(4)**: 510-518.
- Wiley, M.J., Kohler, S.L. 1980. Position changes of mayfly nymphs due to behavioural regulation of oxygen consumption. Canadian Journal of Zoology **58**: 618-622.
- Wiley, M.J., Kohler, S.L. and Seelbach, P.W. 1997. Reconciling landscape and local views of aquatic communities. Freshwater Biology **37**: 133-148.
- Williams, D.D. 1980. Some relationships between stream benthos and substrate heterogeneity. Limnology and Oceanography **25**: 166-172.

- Williams, D., Taveres, A.F. and Bryant, E. 1987. Respiratory device or camouflage: a case for the caddisfly. Oikos **50**: 42-52.
- Winterbourn, M.J. 1973. A guide to the freshwater Mollusca of New Zealand. Tuatara **20**: 141-159.
- Winterbourn, M.J. 1986. Effects of land development on benthic stream communities. New Zealand Agricultural Science **20**: 115-118.
- Winterbourn, M.J., Cowie, B. and Rounick, J.S. 1984. Food resources and ingestion patterns of insects along a West Coast, South Island river system. New Zealand Journal of Marine and Freshwater Research **18**: 379-388.
- Winterbourn, M.J. and Fegley, A. 1989. Effects of nutrient enrichment and grazing on periphyton assemblages in some spring-fed, South Island streams. New Zealand Natural Sciences **16**: 57-65.
- Winterbourn, M.J. and Gregson, K.L.W. 1989. Guide to the Aquatic Insects of New Zealand. Bulletin of the Entomological Society of New Zealand 9, 96 p
- Winterbourn, M. J., Rounick, J.S. and Cowie, B. 1981. Are New Zealand stream ecosystems really different? New Zealand Journal of Marine and Freshwater Research **15**: 321-328.
- Zamora-Muñoz, C. and Alba-Tercedor, J. 1996. Bioassessment of organically polluted Spanish rivers, using a biotic index and multivariate methods. Journal of the North American Benthological Society **15(3)**: 332-352.

Chapter Four

Morphological characteristics of the New Zealand freshwater mussel *Hyridella menziesi* in the Whanganui River and its catchment

ABSTRACT

The influence of habitat and environmental characteristics on the morphology of the freshwater mussel *Hyridella menziesi* were investigated in the Whanganui River catchment, North Island, New Zealand. Of the six sites sampled, one had mussels with distinctive shell morphology. Physicochemical and habitat variation was not related to shell morphology but geographical isolation from tectonic movement is one possibility. Habitat characteristics were the most important determinant of distribution. Body weight and annuli number showed greater variation in these populations from lotic habitats, than in previous reports on lentic populations.

Keywords: freshwater mussel, *Hyridella menziesi*, shell morphology, annuli, habitat, environmental variation, lotic.

INTRODUCTION

Freshwater mussels have been used extensively in bio-accumulation studies investigating immediate and historical levels of deleterious substances such as heavy metals (e.g., Imlay, 1982; Millington and Walker, 1983; Carell *et al.*, 1987; Åberg *et al.*, 1995; Nyström *et al.*, 1996) and organic compounds (e.g., Kauss and Hamdy, 1985; Storey and Edward, 1989). Freshwater mussels display a high degree of morphological variability in response to a range of environmental variables (Tevesz and Carter, 1980). This variability can include changes in shell dimensions, shell and body weight, surface erosion and shell abnormalities (e.g., Cvancara, 1970; Green, 1972; Harmen, 1972; Hinch *et al.*, 1986; Roper and Hickey, 1994; Müller and Patzner, 1996; Kesler and Downing, 1997).

Physical characteristics such as dissolved oxygen, turbulence and temperature as well as general trophic dynamics are markedly dissimilar between lotic and lentic habitats (Nobes, 1980). The majority of morphological studies involving freshwater mussels overseas have been conducted in lentic environments. Studies of *Hyridella menziesi*, one of four New Zealand freshwater mussel species (Winterbourn, 1973), have been conducted in lakes or impounded rivers (e.g., Grimmond, 1968; Forsyth and McCallum, 1978; James, 1985; James 1989; Roper and Hickey, 1994). The aim of this study was to

investigate the influence of environmental conditions and habitat type on lotic populations of *Hyridella menziesi* in the Whanganui River catchment.

STUDY SITES

Mussel samples were collected between spring 1996 and summer/autumn 1997 from six sites within the Whanganui River catchment representing six geographic populations (Fig. 4.1). The area sampled for mussels varied greatly between sites due to major differences in mussel density and habitat type. Because of this sample sizes among the six sites varied from 11 and 71 mussels.

Table 4.1. Site characteristics and mussel sample sizes from six sample sites in the Whanganui River catchment taken between 1996 and 1997.

Site	Width (m)	Depth (cm)	Mussel sample size	Area sampled (m)	Substrate characteristics	Altitude (m)
Pipiriki	28	230	23	8×20	sediment and large boulder	60
Jerusalem	25	140	11	7×15	sediment and cobble/boulder	40
Lismore	1	38	40	1×1	sand/silt	40
Nixons	1.9	9-30	31	280	silt/soft mud	< 20
Hapurua	3.1	15-30	19	180	sand/silt	180
Kakahi	6	30-60	71	1×1	sand/silt	280

Both sites at Pipiriki and Jerusalem were situated in the main Whanganui River channel by the townships of respective names. At Pipiriki, mussels were found in pockets of sediment between large stable boulders in groups of one to three individuals. Habitat and environmental characteristics 10 kilometres downstream at Jerusalem were similar to that of Pipiriki with slightly higher levels of deposited sediment.

At Lismore geology consisted of unstable Quaternary sands and silts that form the predominant substrate type for streams in this area. Mussels were buried with none visible at the substrate surface. Catchment land use comprised of exotic forestry and dairy/sheep farming. No current flow was apparent.

The site at Nixons Creek was located in suburban Wanganui with a catchment of low gradient pasture land. Mussels occurred in groups of between 1 to 10 individuals. In many areas urban vegetation and structures provided shelter from terrestrial predation and strong current with plant roots often improving the cohesion of soft consolidated mud where mussels were commonly situated.

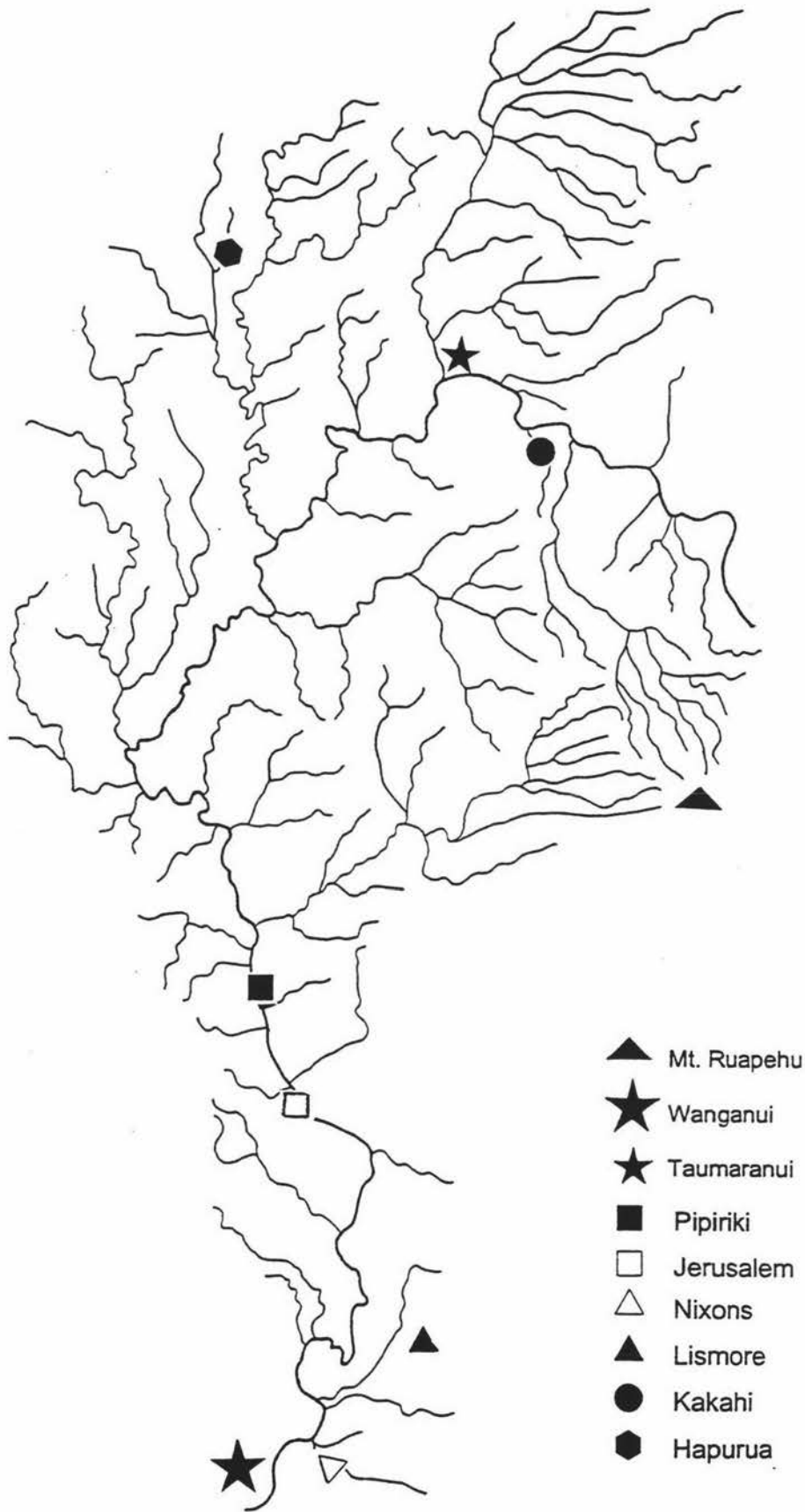


Figure 4.1. Location of the six sites where samples of the freshwater mussel *Hyridella menziesi* were collected during 1996 and 1997.

Hapurua stream is located in the north western boundary of the Whanganui River catchment and is similar geologically to the Lismore and Nixons sites. Current speed was typical of a low gradient stream. The banks were heavily eroded from cattle access which contributed to the consolidated sand/silt/mud substrate. Mussels were located sporadically in small groups or found individually.

Kakahi stream is situated 14 km east of Taumaranui with a catchment developed predominantly for pastoral agriculture. Mussels were collected from the banks of a large pool formed by a stream bend.

METHODS

Mussel densities varied markedly between populations so quantitative estimates of density were not practical due to the relative scarcity of mussels at some sites. Sites were often located and searched on the basis of anecdotal evidence that suggested mussel presence. This evidence was acquired through feedback from local newspaper articles, and consultation with residents and local iwi. A total of over 50 localities were searched for *Hyridella menziesi* between December 1995 and November 1997. Mussels were located by visual contact and hand searches of the substrate which has been found effective in other studies (Cvancara, 1970; Roper and Hickey, 1994).

Sites in the main Whanganui river channel were both searched and sampled using snorkelling equipment. Water levels were low enough at other sites to collect mussels without diving. Physicochemical measurements and invertebrate samples were collected at each site. Macroinvertebrates were sampled by taking four replicate samples collected at each site using a 250 μm mesh, 0.1 m^2 Surber sampler and preserved in 70% alcohol. Samples were sieved using a 250 μm Endecott sieve and animals identified and enumerated with a 40 \times microscope using available keys (McFarlane, 1951; Winterbourn, 1973; Chapman and Lewis, 1976; Cowley, 1978; Winterbourn and Gregson, 1989; Towns and Peters, 1996). If taxa could not be identified to species they were assigned to apparent morphospecies.

Invertebrate samples were taken from riffles instead of the slow flowing areas that mussels commonly inhabited. This was to gain an understanding of water quality and

the wider invertebrate fauna rather than to sample invertebrates characteristic to habitats similar to those occupied by *Hyridella menziesi*. Physicochemical measurements recorded consisted of conductivity, width, depth and pH.

All individuals used for morphometric analysis were greater than 45 mm long to avoid changes in morphology associated with allometric growth (Roper and Hickey, 1994). Length was measured as the maximum antero-posterior dimension of the shell, height as the maximum dorso-ventral dimension measured at right angles to length, and depth as the maximum transverse dimension of the shell with both valves in the normal closed position. Dry flesh weight was determined after careful removal of the body from the shell and air drying in an incubator at 70°C for 48 hours. Both valves were weighed after drying in the same conditions. Cavity volume was measured by weighing the amount of water that filled one valve and multiplying this by 2. Shell length, width, depth and weight were measured with vernier callipers. The left valve of each shell was examined for erosion and the area within which eroded periostracum occurred was recorded according to categories modified from Roper and Hickey (1994). These categories ranged from A to I. This technique was used to gain a rank measure of erosion between mussels rather than a quantitative measure of eroded area. Attempts were made to age mussels by counting shell annuli using the preparation technique of Grimmond (1968).

Differences between morphometric variables were examined using one-way analysis of variance (ANOVA) using the SYSTAT statistical package (SYSTAT, 1996) and Pearson correlations were conducted between physicochemical and biological stream measures to investigate relationships between morphological and environmental characteristics. Data was transformed where appropriate using $\log_{10}(x+1)$ transformations to remove heterogeneity of variance. Two physical condition indices were calculated (e.g., Roper and Hickey, 1994).

$$CI_{\text{flesh: shell weight}} = \frac{\text{dry flesh weight (mg)}}{\text{shell weight (g)}}$$

$$CI_{\text{flesh: shell volume}} = \frac{\text{dry flesh weight (mg)}}{\text{shell volume (ml)}}$$

Canonical discriminate analysis (SYSTAT, 1996) on $\log_{10}(x+1)$ transformed values of length, width, depth and shell weight was used to examine morphological differences between mussel populations. The MCI (Stark, 1985) was calculated for each site from macroinvertebrate samples (refer to Chapter Two for formula).

RESULTS

Mussel densities varied greatly between sites and small sample sizes were directly related to low mussel densities. Average measures of length, width and depth suggested that variation among most sites in these three characteristics are the result of scale differences and not changes in the relative proportion of each characteristic. Mussels from Hapurua were the one exception where mean and variance of shell depth were markedly higher than any other site (Fig. 4.2).

Shell length distributions were comparable in width for all six geographic populations (Fig. 4.3). Mean shell weight showed a similar trend to mean length, width and height but this trend was not proportional between sites (Fig. 4.4a). Average shell weights were similar and heaviest in the Nixons, Jerusalem and Pipiriki mussel populations with shells at the other three sites, especially at Hapurua, weighing less (Fig. 4.4a). The largest mean shell cavity volume was found for mussels from Jerusalem (Fig. 4.4b). Shells from Hapurua had the lowest volume (despite their high depth) due to their overall small size in other dimensions (Fig. 4.4b).

Mussel populations at Kakahi and Pipiriki had the lowest values for both condition indices indicated by steeper slopes to the top left of Fig. 4.5 and Fig. 4.6 and both indices were highly variable among populations ($F_{5,189} = 20.6$, $P = 0.00$ for flesh weight/cavity volume and $F_{5,189} = 13.6$, $P = 0.00$ for flesh weight/shell weight). Least significance difference tests indicate that Pipiriki and Kakahi are responsible for the differences found by ANOVA for both condition indices. While index values were higher in the other four populations they varied depending on which index was used and were not significantly different ($F_{5,189} = 0.66$, $P = 0.15$). For example mussels at Lismore had high flesh weight to shell weight ratios but were lower when substituting shell volume for shell weight.

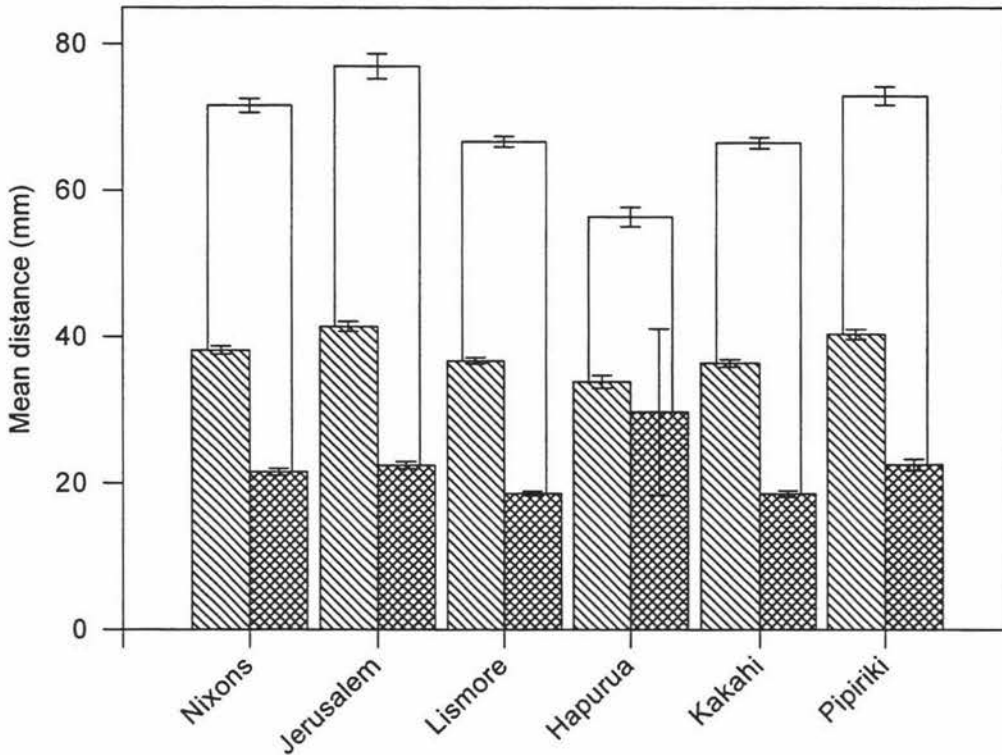


Figure 4.2. Mean (± 1 S.E.) measurements of mussel shell length (no hatching), width (diagonal hatching), and depth (reticulated hatching) from six sites in the Whanganui River catchment.

Mean shell erosion differed significantly among populations ($F_{5,189} = 19.7$, $P = 0.00$) being highest in shells from the main river sites at Jerusalem and Pipiriki, and lowest at Lismore (Fig. 4.7). Erosion varied widely among shells from Kakahi and this may result from a large sample size. Erosion was not correlated to any associated indicators of size/age like length, width and shell weight. Channel width was significantly correlated with shell erosion ($r = 0.95$, $P = 0.00$) but not with shell weight ($r = 0.64$, $P = 0.17$). Demarcation of annuli varied between shells and overall was not clear enough to provide age estimates.

MCI values ranged from 85 to 120 (Table 4.1). MCI scores of 80 or less are considered polluted while scores of 100 or more are considered healthy (Stark, 1985). Conductivity and pH ranged between 86 and 610 μScm^{-2} (Kakahi to Nixons), and 7.2 to 7.8 (Pipiriki to Lismore), respectively. It should be noted that chemical measures were not all recorded at the same time.

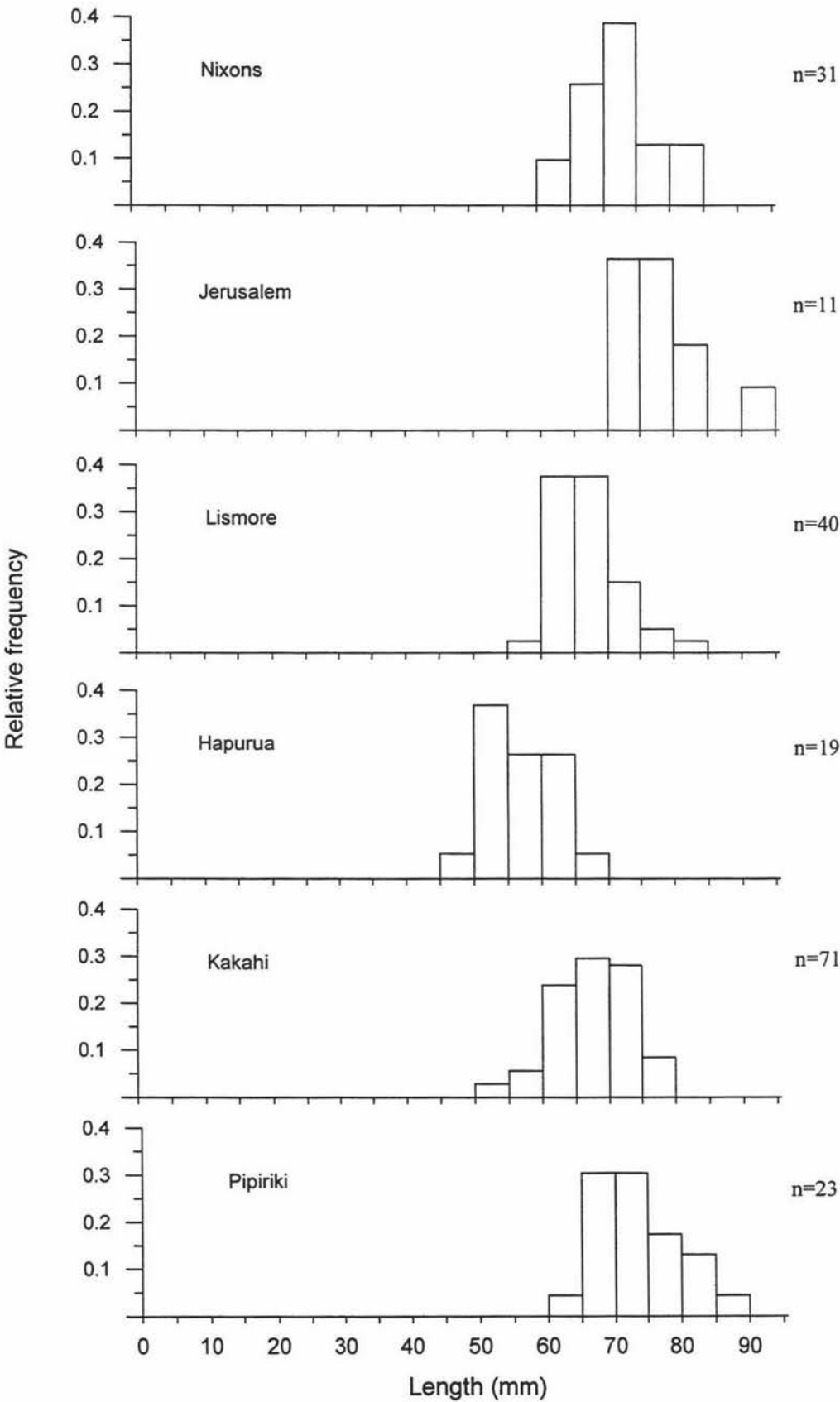


Figure 4.3. Relative frequencies of size classes for mussel shell length from six sites in the Whanganui River catchment.

Canonical discriminant analysis found significant differences between populations based on length, width, depth and shell weight ($F_{5,189} = 10.7$, $P = 0.00$). Plotting factor 1 and 2 canonical score means and values show that mussels from Hapurua are distinct from the other groups with lower factor 1 scores (Fig. 4.8). Factor 1 explained 81% of variation in the data and the classification of Hapurua mussels was of 95% certainty. There was little separation of the other sites by either factor 1 or factor 2, but Lismore and Kakahi do appear to have greater factor 2 scores. Lismore and Kakahi had almost identical means. Factor 2 was responsible for 13% of the variation in the data.

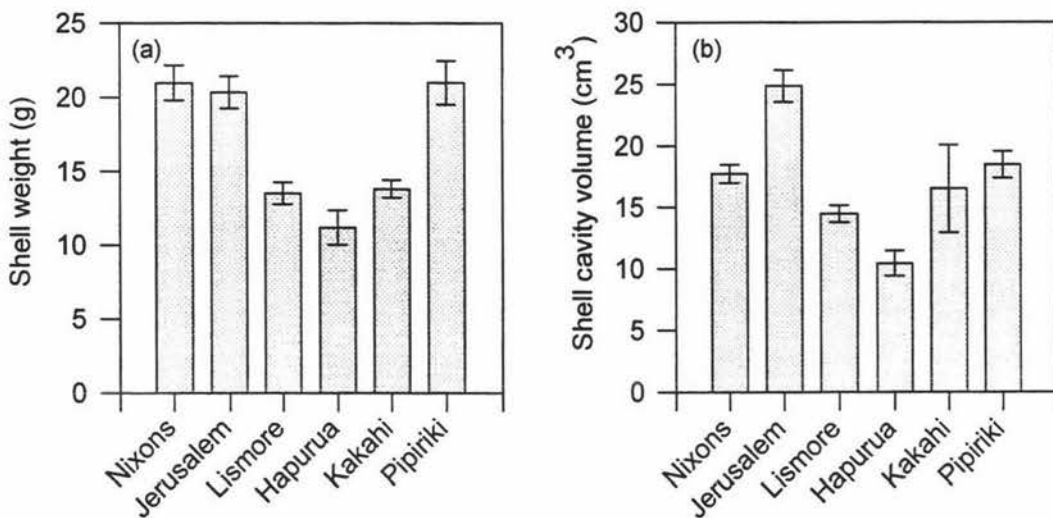


Figure 4.4. Mean (± 1 S.E.) (a) shell weight and (b) shell cavity volume of mussels from six sites in the Whanganui River catchment.

Shell length was the most important influence on factor 1 with shell weight being of primary importance along factor 2. Overall, length was the most important morphological characteristic (F to enter = 26.8). Shell depth had the lowest F to enter value (3.95) but was second to highest (F to enter = 10.4) when using a stepwise discrimination. This is due to a strong allometric correlation between shell weight and length ($r = 0.89$, $P = 0.02$).

Levels of pH were negatively correlated with factor 1 scores ($r = -0.7$, $P = 0.04$) with values ranging from 7.2 to 7.8. MCI and conductivity used to estimate water quality were not significantly correlated with mussel dimensions or factor 1 scores.

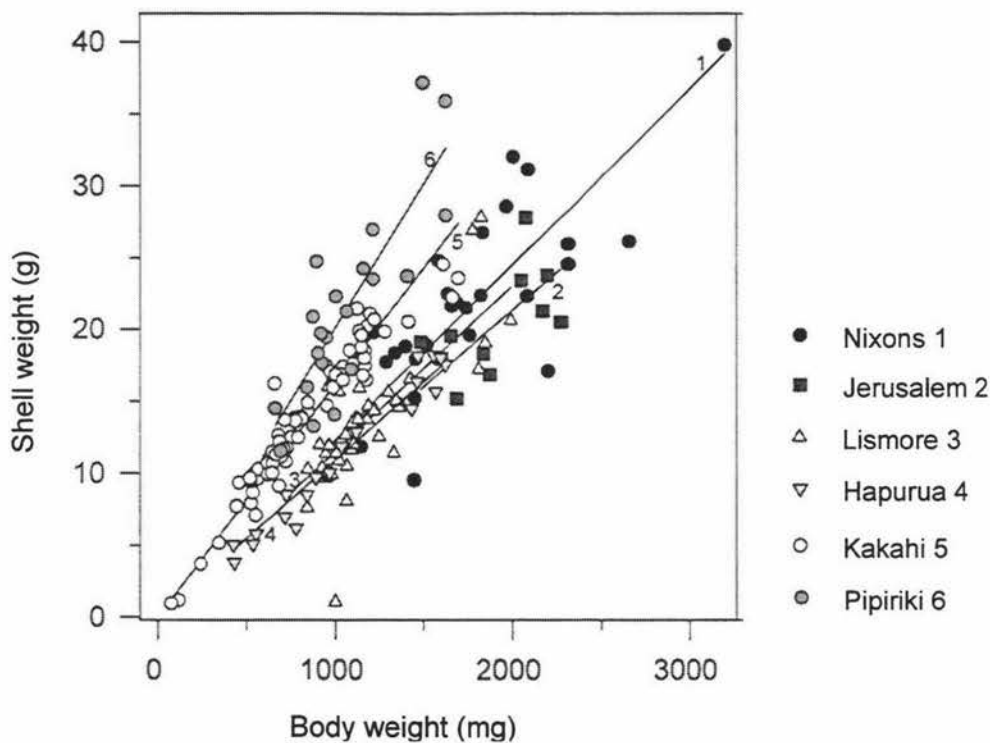


Figure 4.5. Shell weight vs. body weight for mussels from six sites in the Whanganui River catchment. Regression lines are shown.

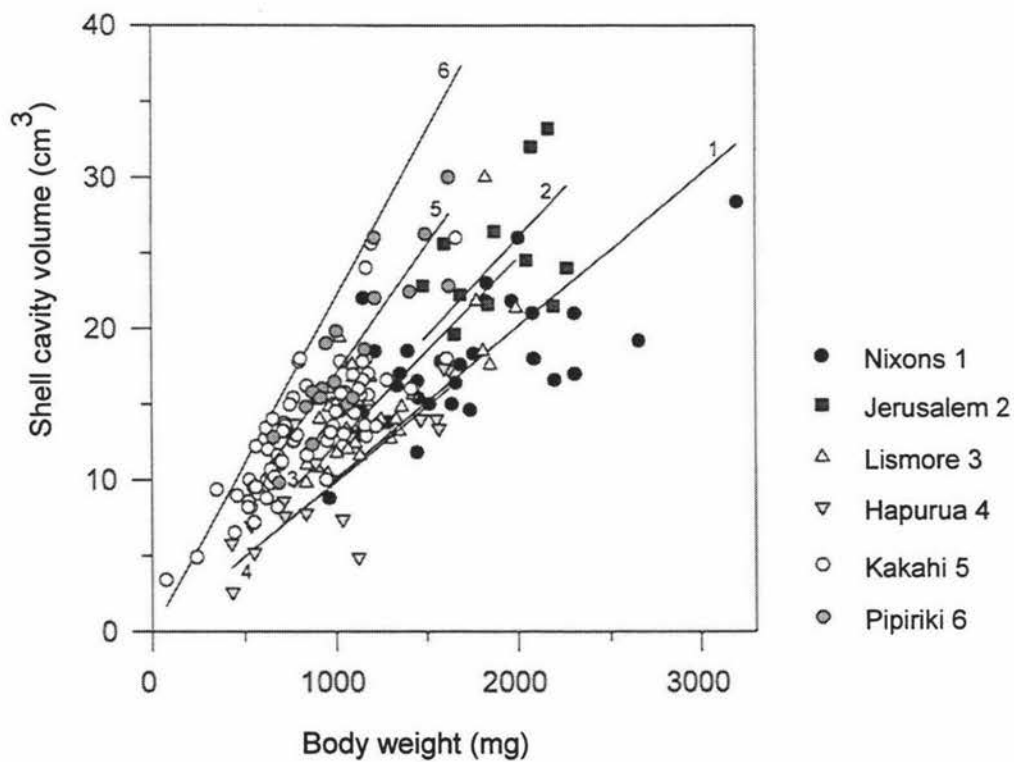


Figure 4.6. Shell cavity volume vs. body weight for mussels from six sites in the Whanganui River catchment. Regression lines are shown.

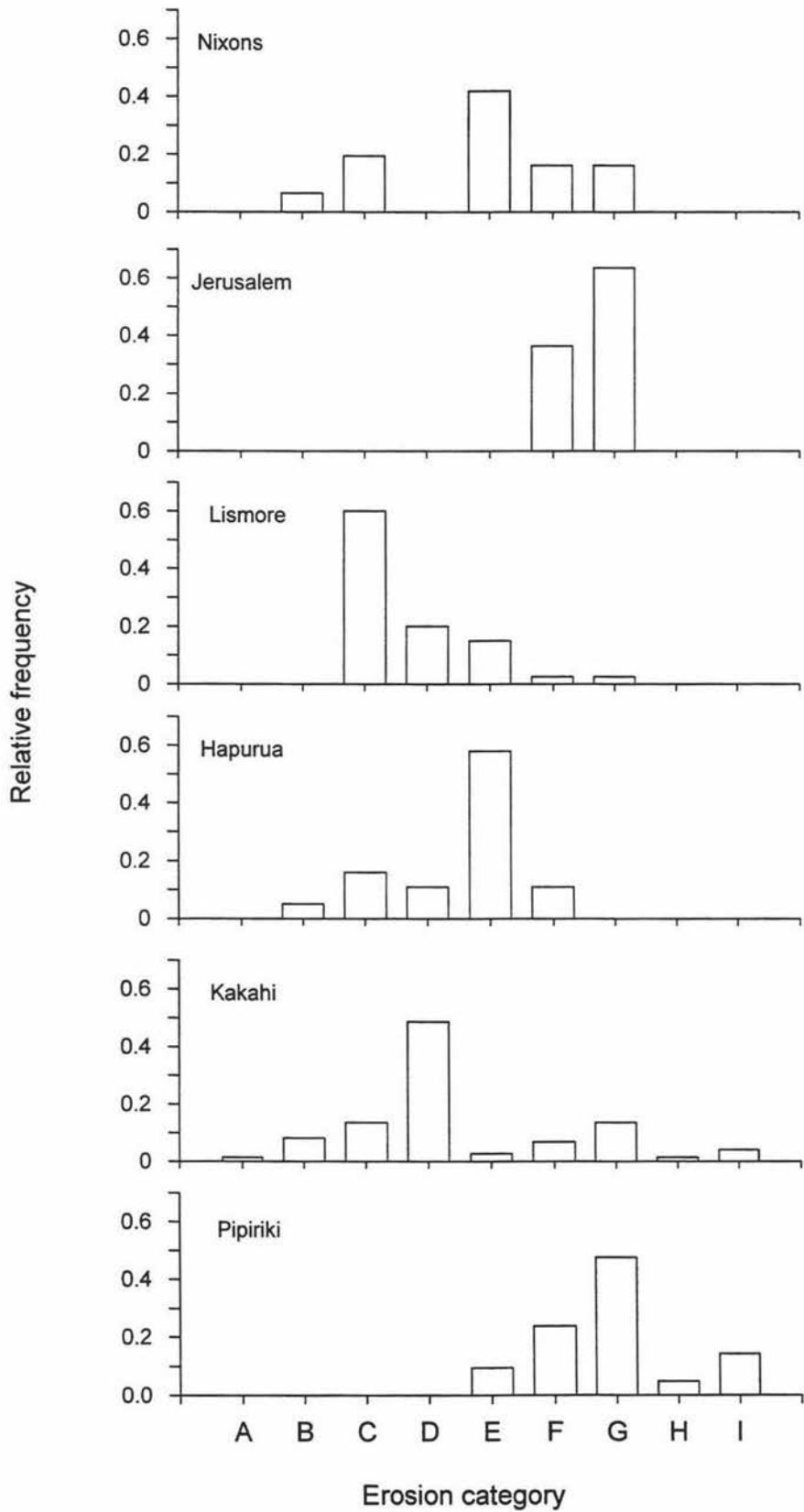


Figure 4.7. Relative frequencies of shell erosion from six sites in the Whanganui River catchment. Letters A to I represent increasing erosion levels. A < 1%; B 1-2%; C 2-5%; D 5-12%; E 12-20%; F 20-30%; G 30-40%; H 40-50%; I > 50%.

Table 4.2. Physicochemical and biotic measures of water quality from the six sample sites in the Whanganui River catchment taken between 1996 and 1997.

Site	Width (m)	Depth (cm)	pH	Conductivity (μScm^{-2})	MCI
Nixons	1.9	9-30	7.3	610	85
Jerusalem	26	150	7.3	140	108
Lismore	1	38	7.8	359	85
Hapurua	3.1	15-30	7.5	137	102
Kakahi	6	30-60	7.3	86	121
Pipiriki	27	230	7.2	222	100

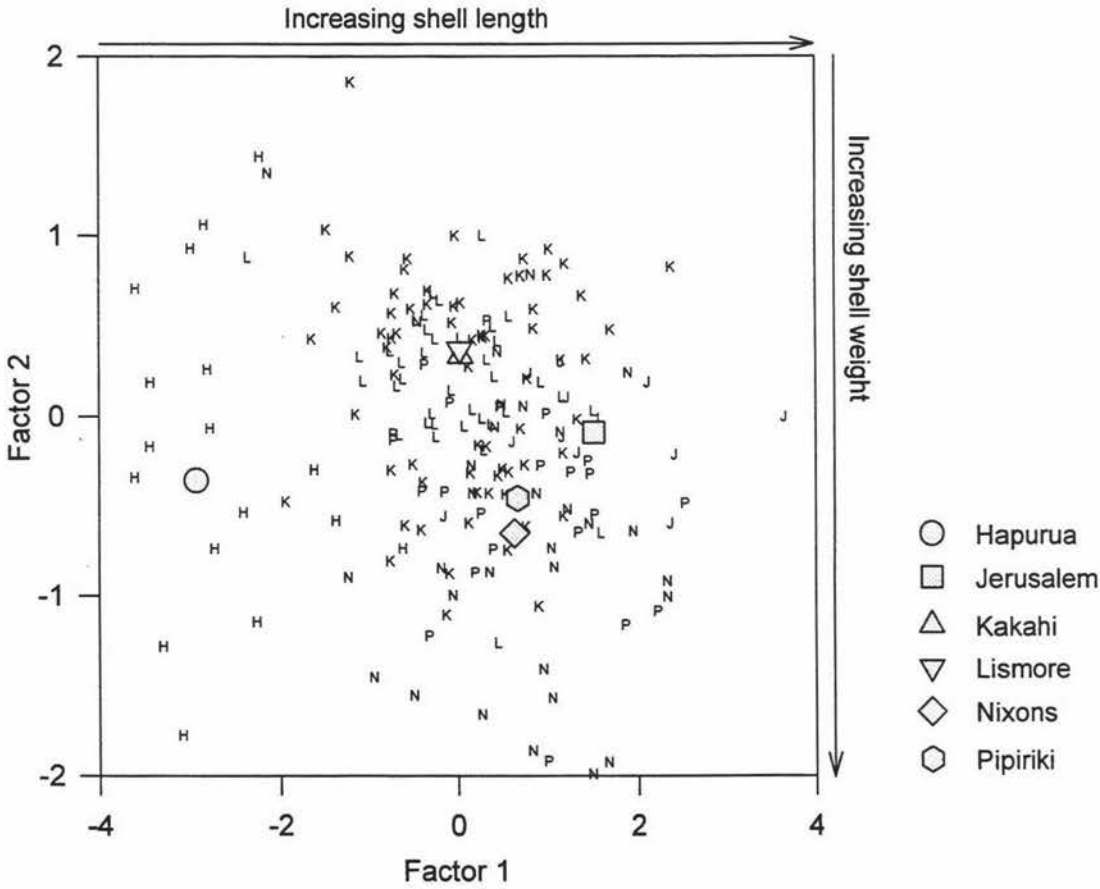


Figure 4.8. Plot of Factor 1 and factor 2 of the canonical discriminant analysis based on length, width, depth and shell weight. Means are shown by symbols and individual mussels are indicated by the first letter of their respective site.

DISCUSSION

The density and spatial patterns of mussels varied widely between sites but overall shape and structure was similar (excluding scale differences) for all but the Hapurua population. Amounts of shell erosion and levels of condition varied significantly among populations. However, morphology was not related to any chemical or biological measures of water quality.

Population density appears to be regulated primarily by temporal and spatial habitat stability (e.g., Cvancara, 1970) combined with the presence of fine substrate. A lack of resilience compared to other stream invertebrates means they are reliant on suitably sheltered or stable habitat (Cvancara, 1970) such as areas of lower velocity (Salmon and Green, 1983). From the anecdotal information collected (much of which dated back 50 years), it appears that populations of *Hyridella menziesi* are becoming scarcer in the Whanganui River. Possible reasons for this include increased silt loading, agrochemical pollution and lower water quality. I suggest that reduced habitat stability over the history of agricultural development in the Whanganui River catchment may have lead to reduced abundances of *Hyridella menziesi*. The effects of pastoral agriculture on aquatic ecosystems are discussed further in Chapter Two.

Overall, shell proportions were similar between sites but differed in mean size. This suggests mean age differences between sites deduced from shell length can relate well to accurate annuli counts (James, 1985) but the possibility of morphological clines (Dell, 1952) between sites hinders a comparison of this kind. It is difficult to decide what constitutes a 'population' as all sites occur in the same catchment and are not isolated except by distance. However, the mussels from Hapurua stream proved to be distinctive in that they were relatively deep in comparison to their other dimensions. Water quality was not correlated with any measures of mussel morphology and a MCI score of 2 assigned to *Hyridella menziesi* indicates its tolerance of poor water quality. Therefore it is unlikely that water quality, chemistry or habitat are responsible for the distinctive nature of mussels from Hapurua.

Hapurua stream is situated at the north western point of the catchment where streams flow along low gradients much closer to the coast north of Taranaki than to the outlet of the Whanganui River, well to the south. A geological map of the area (Grange, 1927)

shows a fault line running between the headwaters of the Mokau and Whanganui Rivers, 8 km from where mussels were collected. Land on the Mokau side of the fault has been elevated simultaneously as the Whanganui side has dropped, potentially cutting off the Mokau headwaters and diverting them into the Whanganui River (Neal pers. comm.). Hence mussels from Hapurua may have originated from the Mokau catchment explaining their dissimilarity to other mussels in the Whanganui. Clearly, examination of more mussel populations in both the Mokau and Whanganui catchments needs to be undertaken to test this hypothesis.

Body mass and condition varied widely between mussel populations. Starvation for a period of 31 days has been shown to produce 23% to 34% losses in body mass of *Hyridella menziesi* (Roper and Hickey, 1995). This loss can presumably be regained in favourable conditions suggesting that body mass is in a continual state of flux, particularly in highly variable environments. Condition indices then are only representative of a single point in time being potentially confounded by the spatial and temporal factors of the sampling regime. Certainly, applying indices calculated for one population to another is risky due to the likelihood of morphological clines (Dell, 1952; Cvancara, 1970; Green, 1972). Mussels from Pipiriki and Kakahi were sampled much earlier in the study (spring, 1996) whereas others were sampled later (summer/autumn 1997). Low condition for Pipiriki and Kakahi mussels could be the result of less favourable growth conditions over winter.

In older mussels erosion may often be more pronounced (Hinch and Green, 1987). However, no significant correlations were found between mussel size (i.e., length/width/depth/shell weight) and erosion. It has been suggested that chemical causes can influence shell etching (Åberg *et al.*, 1995). Despite the relationship between pH and factor 1 of the canonical discriminant analysis, the lack of a significant direct correlation between erosion and pH, and the small pH range among sites suggests water chemistry is not important. If larger streams and rivers are considered to have greater levels of turbulence, particularly in spate, turbulence would appear to be affecting shell erosion (e.g., Hinch and Green, 1987; Green *et al.*, 1989).

Attempts were made in this study to count shell annuli but estimates were not accurate enough for statistical comparison. Difficulties in determining true annuli from false have been noted elsewhere (e.g., Coon *et al.*, 1977; Carell *et al.*, 1987; Green *et al.*,

1989; Kesler and Downing, 1997). The technique used was that of Grimmond (1968) and has been applied successfully in other studies of *Hyridella menziesi* (e.g., Grimmond, 1968; James, 1985; James, 1989; Roper and Hickey, 1994). However these involved mussels in lentic habitats (except for the study by Roper and Hickey (1994) in impounded reaches of the Waikato River). Errors occur in annuli counts when difficulty arises in distinguishing dark winter lines from growth disturbance lines (Carell *et al.*, 1987). Variation in physical factors such as rapid temperature fluctuations, storm disturbance and environmental stress, all of which are more pronounced in lotic environments (Salmon and Green, 1983), make for more ambiguous annuli records (Shuster, 1951; Stansbery, 1967; Imlay, 1982; Green *et al.*, 1989). For example, mussels can migrate vertically through the substrate in response to adverse environmental conditions (Amyot and Downing, 1997). In this study Lismore mussels became endobenthic in response to summer drought. This response will lead to growth disturbance annuli unrelated to winter growth interruption. As well as the timing of variation, the magnitude of variation may be important. Finally, other lentic studies involving the assessment of winter growth annuli have been conducted predominantly in temperate to subarctic regions of the Northern Hemisphere (e.g., Bailey and Green, 1987; Carell *et al.*, 1987) where seasonal changes are more pronounced leading to greater definition in shell annuli (McCuaig and Green, 1983).

In this study shell proportions were similar among all sites except for Hapurua. While habitat stability is likely to be responsible for the degree of shell erosion and population densities neither physical or chemical factors appear responsible for the distinctive morphology of Hapurua shells. Further work needs to evaluate the possibility of biogeographical changes in this part of the catchment. Environmental variability within most stream ecosystems means condition indices may be misleading as single spot measures of body mass used to calculate these are in constant flux. This variability is also apparent in shell annuli making ageing techniques using this method unreliable in this study.

REFERENCES

- Åberg, G., Wickman, T. and Mutvei, H. 1995. Strontium isotope ratios in mussel shells as indicators of acidification. Ambio **24(5)**: 265-268.
- Amyot, J.P. and Downing, J.A. 1997. Seasonal variation in vertical and horizontal movement of the freshwater bivalve *Elliptio complanata* (Mollusca: Unionidae). Freshwater Biology **37**: 345-354.
- Bailey, R.C. and Green, R.H. 1987. Spatial and temporal variation in a population of freshwater mussels in shell lake, N.W.T. Canadian Journal of Fisheries and Aquatic Sciences **46**: 1392-1395.
- Carell, B., Forberg, S., Grundelius, E., Henrikson, L., Johnels, A., Lindh, U., Mutvei, H., Svärdström, K. and Westermarck, T. 1987. Can mussel shells reveal environmental history? Ambio **16(1)**: 2-10.
- Chapman, A. and Lewis, M. 1976. An introduction to the freshwater Crustacea of New Zealand. Auckland: Collins.
- Coon, T.G., Eckblad, J.W. and Trygstad, P.M. 1977. Relative abundance and growth of mussels (Mollusca: Eulamellibranchia) in pools 8, 9 and 10 of the Mississippi River. Freshwater Biology **7**: 279-285.
- Cowley, D.R. 1978. Studies on the larvae of New Zealand Trichoptera. New Zealand Journal of Zoology **5**: 639-750.
- Cvancara, A.M. 1970. Mussels (Unionidae) of the Red River Valley in North Dakota and Minnesota, U.S.A. Malacology **10(1)**: 57-92.
- Dell, R.K. 1952. The freshwater Mollusca of New Zealand Part 1- The genus *Hyridella*. Transactions of the Royal Society of New Zealand **81(2)**: 221-237.

- Forsyth, D.J. and McCallum, I.D. 1978. *Xenochironomus canterburyensis* (Diptera: Chironomidae), a commensal of *Hyridella menziesi* (Lamellibranchia) in lake Taupo; features of pre-adult life history. New Zealand Journal of Zoology **5**: 795-800.
- Grange, L.I. 1927. The geology of the Tongaporutu-Ohura subdivision, Taranaki division. New Zealand Geological Survey Bulletin no. 31.
- Green, R.H. 1972. Distribution and morphological variation of *Lampsilis radiata* (Pelecypoda, Unionidae) in some central Canadian lakes: a multivariate statistical approach. Journal of the Fisheries Research Board Canada **29**: 1565-1570.
- Green, R.H., Bailey, R.C., Hinch, S.G., Metcalfe, J.L. and Young, V.H. 1989. Use of freshwater mussels (Bivalvia: Unionidae) to monitor the nearshore environment of lakes. Journal of Great Lakes Research **15**(4): 635-644.
- Grimmond, N.M. 1968. Observations on growth and age in *Hyridella menziesi* (Mollusca, Bivalvia) in a freshwater tidal lake [M.Sc. thesis]. Dunedin, New Zealand: University of Otago.
- Harmen, W.N. 1972. Benthic substrates: their effect on freshwater Mollusca. Ecology **53**(2): 271-277.
- Hinch, S.G., Bailey, R.C. and Green, R.H. 1986. Growth of *Lampsilis radiata* (Bivalvia: Unionidae) in sand and mud: a reciprocal transplant experiment. Canadian Journal of Fisheries and Aquatic Sciences **43**: 548-552.
- Hinch, S.G. and Green, R.H. 1987. Shell etching on clams from low-alkalinity Ontario lakes: a physical or chemical process? Canadian Journal of Fisheries and Aquatic Sciences **45**: 2110-2113.
- Imlay, M.J. 1982. Use of shells of freshwater mussels in monitoring heavy metals and environmental stresses: a review. Malacological Review **15**: 1-14.

- James, M.R. 1985. Distribution, biomass and production of the freshwater mussel, *Hyridella menziesi* (Gray), in lake Taupo, New Zealand. Freshwater Biology **15**: 307-314.
- James, M.R. 1989. Ecology of the Freshwater mussel *Hyridella menziesi* (Gray) in a small oligotrophic lake. Archive für Hydrobiologia **108(3)**: 337-348.
- Kauss, P.B. and Hamdy, Y.S. 1985. Biological monitoring of organochlorine contaminants in the St. Clair and Detroit Rivers using introduced clams, *Elliptio complanatus*. Journal of Great Lakes Research **11(3)**: 247-263.
- Kesler, D.H. and Downing, J.A. 1997. Internal shell annuli yield inaccurate growth estimates in the freshwater mussels *Elliptio complanata* and *Lamprolaima radiata*. Freshwater Biology **37**: 325-332.
- McCuaig, J.M. and Green, R.J. 1983. Unionid growth curves derived from annual rings: a baseline model for Long Point Bay, Lake Erie. Canadian Journal of Fisheries and Aquatic Sciences **40**:436-442.
- McFarlane, A.G. 1951. Caddisfly larvae (Trichoptera) of the family Rhyacophilidae. Records of the Canterbury Museum **5**: 267-289.
- Millington, P.J. and Walker, K.F. 1983. Australian Freshwater mussel *Velesunio ambiguus* (Philippi) as a biological monitor for zinc, iron and manganese. Australian Journal of Marine and Freshwater Research **34**: 873-892.
- Müller, D. and Patzner, R.A. 1996. Growth and age structure of the swan mussel *Anodonta cygnea* (L.) at different depths in lake Mattsee (Salzburg, Austria). Hydrobiologia **341**: 65-70.
- Nobes, R.G. 1980. Energetics of the freshwater mussel *Hyridella menziesi* (Gray) [M.Sc. thesis]. Hamilton, New Zealand: University of Waikato.

- Nyström, J., Dunca, E., Mutvei, H. and Lindh, U. 1996. Environmental history as reflected by freshwater pearl mussels in the River Vramsån, Southern Sweden. Ambio **25(5)**: 350-355.
- Roper, D.S. and Hickey, C.W. 1994. Population structure, shell morphology, age and condition of the freshwater mussel *Hyridella menziesi* (Unionacea: Hyriidae) from seven lake and river sites in the Waikato River system. Hydrobiologia **284**: 205-217.
- Roper, D.S. and Hickey, C.W. 1995. Effects of food and silt on filtration, respiration and condition of the freshwater mussel *Hyridella menziesi* (Unionacea, Hyriidae): Implications for bioaccumulation. Hydrobiologia **312(1)**: 17-25.
- Salmon, A. and Green, R.H. 1983. Environmental determinants of unionid clam distribution in the middle Thames river, Ontario. Canadian Journal of Zoology **61**: 832-837.
- Shuster, C.N. 1951. On the formation of mid-season checks in the shell of *Mya arenaria*. Anatomical Record **111**: 127.
- Stansbery, D.H. 1967. Growth and longevity of najades from Fishery Bay in western Lake Erie. Annual Report of the American Malacological Union. p 10-11.
- Stark, J.D. 1985. A macroinvertebrate community index of water quality for stony streams. Water and Soil Miscellaneous Publication **87**: 53p.
- Storey, A.W. and Edward, H.D. 1989. The freshwater mussel, *Westralunio carteri* Iredale, as a biological monitor of organochlorine pesticides. Australian Journal of Marine and Freshwater Research **40**: 587-593.
- SYSTAT. 1996. SYSTAT for Windows: Statistics, Version 6 Edition. Evanston, Illinois: SYSTAT Inc.

- Tevesz, M.J.S. and Carter, J.G. 1980. Environmental relationships of shell form and structure of Unionacean bi-valves. In: Rhodes, D.C. and Lutz, R.A., editors. Skeletal growth of aquatic organisms. New York: Plenum. p 295-322.
- Towns, D.R. and Peters, W.L. 1996. Leptophlebiidae (Insecta: Ephemeroptera). Fauna of New Zealand **36**: 37-50.
- Winterbourn, M.J. 1973. A guide to the freshwater Mollusca of New Zealand. Tuatara **20**: 141-159.
- Winterbourn, M.J. and Gregson, K.L.W. 1989. Guide to the Aquatic Insects of New Zealand. Bulletin of the Entomological Society of New Zealand 9, 96 p.

ACKNOWLEDGEMENTS

ACKNOWLEDGEMENTS

This thesis would not have been possible without the combined assistance of a large number of people. Except for gratitude there was no material incentive for these people to contribute their valuable time and energy and all those who did have my greatest appreciation and respect.

At Massey University I would like to extend a big thank you to my supervisors and stream ecology mentors Russell Death and Ian Henderson for sparking my interest in stream ecology. Also for all their guidance and good advice from set-up to write-up of this project. The collective efforts of fellow students has been a huge help with the day to day thesis learning curve and I would like to thank the following people. Andrew Taylor for introducing and helping me in the wonderful world of computers and invertebrate identification. Graeme Franklyn for assistance with identification and general advice. Kimberly Dunning for Lismore raw data. Also to James Bower, Peter Ritchie, Reece Fowler, Karen Eggers, Brent Stephenson and Jason Gibson for general assistance. Cheers to Jens Jorgensen for help with artificial substrates and the manufacture of other miscellaneous pieces of equipment. Also to Vince Neal for advice on geological matters.

Thank you to the Department of Conservation for generous financial/logistical support, and competent staff. These include Eric Pyle for the initial set-up of the study. Rosemary Miller for her patience and help with draft material. Dennis, Milo, and Eddie at Pipiriki for accommodation and transport up the River (Paua's in the mail), and Steve McGill for accommodation at Taumaranui.

Thank you to Chris Fowles, John Stark and the Manawatu Wanganui Regional Council for miscellaneous Whanganui River and catchment data. I haven't used it but thanks for providing it so freely. Also to my Holden that despite its age never broke down on any of those remote dirt roads, and both the Massey University Climbing Wall and the Tararua Ranges for keeping me sane during my time in Palmerston North.

Finally, thanks to my family for their unwavering support and continual encouragement.