Copyright is owned by the Author of the thesis. Permission is given for a copy to be downloaded by an individual for the purpose of research and private study only. The thesis may not be reproduced elsewhere without the permission of the Author.

Genetic studies of *Phytophthora* on *Theobroma cacao* from East New Britain and Bougainville (Papua New Guinea)

A thesis presented in partial fulfilment of the degree of

Master of AgriScience (Horticulture)

at Massey University,
Palmerston North, New Zealand



James Butubu 2016

ABSTRACT

DNA was extracted from 14 Phytophthora isolates from Theobroma cacao plants (SG2 hybrids, hybrid derived clones and Trinitario varieties) collected from New Britain and Bougainville in Papua New Guinea (PNG). A fragment of the mitochondrial genome cytochrome b (cytb) region was amplified from these DNAs using the polymerase chain reaction (PCR) and compared to cytochrome b sequences from Phytophthora palmivora and other *Phytophthora* (two isolates previously isolated from cocoa lesions and two *P*. palmivora isolates obtained from culture collection-Australia). All isolates were identical in their cytb gene sequence and similar to P. palmivora. Additionally, we sequenced the mitochondrial genomes of four isolates from PNG. The syntenic arrangement of genes in one complete assembly was compared with other published mitochondrial genomes. The sequences of four mitochondrial genes (COII, nad2, rps10 and SecY) from the four PNG isolates were aligned with orthologues from accessions of P. palmivora and other Phytophthora species available in the NCBI Genbank reference database. A concatenated data matrix was produced with 2,295 homologous sequence positions. 34 accessions of Phytophthora (including 14 P. palmivora) were used to construct a maximum likelihood tree of phylogenetic relationships. This reconstruction recovered all 10 major clades of *Phytophthora* previously reported. In this phylogenetic reconstruction, the four PNG isolates were clearly identifiable as P. palmivora and these were closely related to the Clade 4 Phytophthora species P. megakarya and P. quercetora. Of the genes analysed, COII showed greatest variability, resolving P. palmivora into three sub groups. COII was sequenced in all P. palmivora isolates from PNG and used to reconstruct an ML tree. The phylogenetic analyses suggested a potential origin for the PNG strain of P. palmivora in Samoa. Syntenic comparisons of P. palmivora and other clade 4 species identified a potential target for developing a Loop Mediated Amplification (LAMP) assay for P. palmivora near the atpH gene region. DNA amplification primers were designed for this region using PrimerExplorer, V4, Eiken Chemical CO.Ltd) and validated against available DNAs for Clade 4 and other Phytophthora species.

TABLE OF CONTENTS

Pag	ţе	No.
CHAPTER 1		
1.1 Cocoa history and uses		1
1.1.1 Cocoa and PNG economy	•	3
1.1.2 Cocoa diseases pose a major constraint to increasing yield and income		5
1.2 A history of research on <i>Phytophthora</i>	,	6
1.2.1 New species discovery prior to 2000.		7
1.2.2 New species discovery post 2000.	•	7
1.3 Phytophthora palmivora		9
1.3.1 Life cycle of <i>P. palmivora</i>		10
1.3.2 Morphology of <i>P. palmivora</i>		10
1.3.3 Epidemiology of <i>P. palmivora</i>		11
1.4 Disease Management for <i>P. palmivora</i>		12
1.4.1 Improved tolerance to <i>P. palmivora</i>		12
1.4.2 Management and sanitation of trees	••	13
1.4.3 Screening for <i>P. palmivora</i> resistance.		14
1.4.4 Pod Inoculation screening method.		15
1.5 Diagnostics for <i>P. palmivora</i>		15
1.5.1 The Polymerase Chain Reaction.	· • •	17
1.5.2 Dideoxy DNA Sequencing.		19
1.5.3 High Throughput (NGS) DNA sequencing		22
1.5.4 DNA sequence analysis		23
1.5.5 Targeting the nuclear genome		24
1.5.6 Targeting the mitochondrial genome	•	24
1.5.7 Isothermal DNA amplification tests		25
1.6 Aims and objectives of the present study		26

2.1 Introduction	27
2.2 Phytophthora palmivora collections	28
2.3 Culturing of <i>Phytophthora palmivora</i> isolates	28
2.3.1 Culturing of <i>P. palmivora</i> isolates	28
2.3.2 Microscopic examination of cultured <i>P. palmivora</i>	30
2.3.3 Harvesting and storage of <i>P. palmivora</i> mycelium for molecular analysis	31
2.3.4 Microscopic examination of cultured <i>P. palmivora</i>	31
2.3.5 Harvesting and storage of <i>P. palmivora</i> mycelium for molecular analysis	31
2.4 DNA extraction	31
2.4.1 Sample homogenisation	31
2.4.2 Genomic DNA extraction	32
2.4.3 Evaluation of DNA quality and quantity	33
2.5 PCR amplification of mitochondrial loci	33
2.6 Enzymatic purification of PCR products	34
2.7 Sanger sequencing of PCR products	34
2.8 Processing of Sanger sequence data	34
2.9 Next generation sequencing and assembly of whole mitochondrial genomes	34
2.9.1 High through-put sequencing of PNG <i>P. palmivora</i> isolates	35
2.9.2 Mitochondrial genome assembly	35
2.9.3 Mitochondrial genome annotation	36
2.10 Relationships of PNG <i>P. palmivora</i> based on mitochondrial genes	36
2.11 Designing primers for LAMP amplification of <i>P. palmivora</i> specific targets	38
2.12 Testing of the LAMP assay	39
2.12.1 Initial testing of LAMP primers using PCR	39
2.12.2 Optimisation of the LAMP assay	39

2.12.3 Specificity of the LAMP assay	40
2.12.4 Sensitivity of the LAMP assay	40
CHAPTER 3	
3.1 Introduction	42
3.2 Genetic diversity of <i>P. palmivora</i> isolates	42
3.3 Whole mitochondria genome sequencing and comparison	43
3.3.1 Sequencing statistics	43
3.3.2 Genome assemblies	43
3.3.3 Annotation of the <i>P. palmivora</i> genome	45
3.4 Relationships of PNG <i>P. palmivora</i> based on mitochondrial genes	45
3.4.1 DNA sequences and data matrices.	45
3.4.2 Nucleotide substitution models	45
3.4.3 Relationships of PNG <i>P. palmivora</i> based on mitochondrial genes	46
3.5 Characterising PNG <i>P. palmivora</i> isolates for cox1-cox2 sequence variation	48
3.6 Designing primers for LAMP amplification of <i>P. palmivora</i> specific targets	49
3.7 Testing of the LAMP assay	52
3.7.1 Initial testing of LAMP primers using PCR	52
3.7.2 Optimisation of the LAMP assay	52
3.7.3 Specificity of the LAMP assay	52
3.7.4. Sensitivity of the LAMP assay	52
CHAPTER 4	
4.1 Introduction.	56
4.2 Cytochrome b gene analyses of PNG isolates	56

	Page	No
4.3 The mitochondria genome of <i>P. palmivora</i>		57
4.4 Phylogenetic relationship of <i>P. palmivora</i> based on four genes		58
4.5 Cox2 gene sequence variation.		59
4.6 The Design of LAMP primers	•	60
4.7 Testing and implementation of LAMP in diagnostics		61
4.8 Concluding Statement		61
4.9 Future Work	•	62

LIST OF FIGURES

Figure 1.1	Chocolate and cosmetic products made from cocoa bean	2
Figure 1.2	Proportion (%) of total merchandised export (K19484	
	million or. NZ\$ 9917 million) in 2013	4
Figure 1.3	Relative export value of agricultural and marine produce in 2013	4
Figure 1.4	Types of cocoa diseases caused by P. palmivora A) Black pod,	
	B) Canker and C) seedling blight	6
Figure 1.5	Number of described Phytopththora species over time,	
	1876-1995. Source: Braiser (2009)	7
Figure 1. 6	6. Graph depicting number of <i>Phytophthora</i> taxa discovered	
	pre and post 2000. Source: Braiser (2009)	8
Figure 1.7	Different stages of the life cycle of the <i>P. palmivora</i>	10
Figure 1.8	Sporagia types from Isolates collected in West Africa,	
	Note typically thin non-occluded pedicels and variation in	
	sporangial size, e.g 25-27 (Nigeria Isolate), 33-34	
	(Cameroon Isolate). Source: Braiser (1979)	11
Figure 1.9	A cocoa tree trunk affect by canker caused by P. palmivora	
	and treatment with chemical fungicide (metalaxyl	14
Figure 1.1	O Screening resistance to black pod disease (P. palmivora),	
	A) detached pod inoculation test in the lab,	
	B)-leave discs inoculation test in the lab	15
Figure 1.1	1 Morphological forms of sporagispores of <i>P. palmivora</i> ,	
	A) Non-papilate, B) semi-papilate and C) papillated sporangia.	
	Source: Drenth (2001)	16

Figure 1.12 Lupin baiting of <i>Phytophthora</i> from soil samples.	
Source: Drenth (2001)	17
Figure 1.13 Principle of the Polymerase Chain Reaction	18
Figure 1.14 Molecular structure of 2',3-dideoxyadenosine triphosphate (ddATP)	20
Figure 1.15 Form of dideoxynucleotide triphosphate and dideoxynucleotide	
triphosphate used to terminate sequencing reactions	20
Figure 1.16 Illustration of the Chain Termination sequencing method	21
Figure 1.17 Sequencing, contig formation and alignment of contigs	22
CHAPTER 2	
Figure 2.1 Regional Context of current study in New Britain and Bougainville.	
The location of Samoa (Apia) has also been indicated	27
Figure 2.2 Map of PNG showing <i>P. palmivora</i> collection sites (base map from	
d-maps.com)	28
Figure 2.3 Photographs of <i>Theobroma cacao</i> pods from Bougainville (A-C)	
and East New Britain (d-F) with dark discolouration consistent with	
P. palmivora infection. Phytophthora isolated from these pods is	
among that analysed in the present study. Isolate codes (see Table 2.1)	
for pods are – A, Buin3; B, Buin4; C, Buka2; D, Kerv; E, Raul; F, Tav	
(office)	29
Figure 2.4 The process of isolating and culturing <i>P. palmivora</i> from diseased	
Theobroma cacao pods collected in the field. The media pods are	
surface sterilised by first spraying with an insecticide (A) and then	
washing in a NaOCl bath (B). Sterilised pods (C) then have a hole	
drilled into them; surface scrapings are taken from a diseased pod	
(D) and packed into this hole (E). The inoculated media pods are	
covered and incubated in the dark at 24°C to allow the <i>P. palmivora</i>	
infection to develop (F)	30
Figure 2.5 Flowchart for Nextera DNA library preparation	35

Figure 3.1	A visual presentation of the completed mitochondria gene map	
	of P. palmivora isolate from Raulavat, East New Britain. The	
	genes and rRNA, coding regions (green) area also shown. Arrows	
	indicate transcriptional orientation	31
Figure 3.2	Maximum likelihood phylogeny from the combined analysis of	
	atp9-nad9, cox1-cox2, rps10 and secY regions. Numbered nodes	
	indicate clades recognised by Blair et al. (2008), numbers associated	
	with branches are bootstrap support values obtained for 500 replicates	48
Figure 3.3	The regions coloured show where each primers sequence is	
	located as identified by the primer explorer software. Primer	
	sequences are given by the reference sequence (P. palmivora1).	
	Where there is dots (no letters) the nucleotide are identical to the	
	reference genome	51
Figure 3.4	Electrophoresis of PCR products, Ladder, 1=negative control,	
	2=isolate 3, 4=isolate 6 and isolate 11 amplified by F3/B3	
	primers, 5=negative control, 6, 7, and 8 are same isolate but	
	amplified with FIB/BIP primers	53
Figure 3.5	Electrophoresis of LAMP products. Ladder, 1=negative control,	
	2=ratio of 1: 4, 3=ratio of 1:6 and 4=ratio of 1:8 F3/B3 to FIB/BIP	
	primer without Loop primer, 5=Negative control, 6= ratio of	
	1: 4, 7=ratio of 1:6, 8=ratio of 1:8 F3/B3 to FIB/BIP primer with	
	loop primer, isolate but amplified with FIB/BIP primers. Only one	
	isolate of <i>P. palmivora</i> was used	53
Figure 3.6	Different species of <i>Phytophthora</i> tested for DNA amplification	
	using LAMP primers, Ladder, 1=negative control,	
	2= P. palmiyora (PNG-Raul) 3=P. palmiyora	

	(Chest nut-ICMP 1770), 4= P. palmivora (Papaya-ICMP14517),	
	5=P. capitosa, clade 9, ICMP17567), 6=P. agathicidae,	
	(NZFS3128, clade 5), 7=P. cryptogea NZFZ 4156-clade 8)	
	and 8=P. cinnamoni (ICMP20276-clade 7)	54
Figure 3.7	PCR amplification of the specified region of <i>P. palmivora</i> ,	
	DNA at different concentration (high -low), Ladder,	
	1=negative control, $2=1 ng/\mu L$ 3=100pg/ μL , 4= 10pg/ μL	
	5= 1pg/ μ L, 6=100fg/ μ L 7=10fg/ μ L , 8=1fg/ μ L	54
Figure 3.8	LAMP reaction of the specified region of P. palmivora, DNA	
	at different concentration (high -low), Ladder, 1=negative	
	control, $2=1$ ng/ μ L $3=100$ pg/ μ L , $4=10$ pg/ μ L $5=1$ pg/ μ L,	
	$6{=}100 fg/\mu L~7{=}10 fg/\mu L~,~8{=}1 fg/\mu L$	55
Figure 3.9	LAMP reaction detecting P.palmivora isolates (haplotypes)	
	from DNA loaded in different Lanes, lane1=Negative control,	
	lane 2)=Nari 1, lane 3=Nari 2, lane 4)= Raulavat, lane 5= (Buka 4),	
	lane 6= (Buka2), lane 7=Buin3 and lane 8=Tav.Office	55

X

LIST OF TABLES

Table 2.1 Collection details for P. palmivora samples sent to	
Massey University	32
Table 2.2 Details of publically available DNA sequences used	
for phylogenetic analyses	37
CHAPTER 3	
Table 3.1 Statistics for matrices and phylogenetic analyses	46
Table 3.2 Statistics for matrices and phylogenetic analyses	49

ACKNOWLEDGEMENTS

I wish to thank my supervisors Nick Roskruge, Richard Winkworth and Peter Lockhart for their excellent supervision. A special mention of thanks also goes to Trish McLenachan for her assistance with lab work at Massey. This work would not have been possible without support from the Ministry of Agriculture (PNG), New Zealand Aid, the New Zealand BioProtection CoRE, and most of all my family and friends! Special acknowledgement also goes to my colleagues Mrs Olisha Wesley and Mary Paul Marfu for their technical assistance in culturing the fungal pathogen at PNGCCI.