Copyright is owned by the Author of the thesis. Permission is given for a copy to be downloaded by an individual for the purpose of research and private study only. The thesis may not be reproduced elsewhere without the permission of the Author.

Effects of cutting, nitrogen, closing date and water on herbage and seed production in Ruzi grass (*Brachiaria ruziziensis* Germain and Everard)

A thesis presented in partial fulfilment of the requirements for the degree of Doctor of Philosophy (Ph.D) in Plant Science (Seed Technology) at Massey University, New Zealand

> NARONGRIT WONGSUWAN March, 1999.

ABSTRACT

The successful production of Ruzi grass (*Brachiaria ruziziensis*) for both herbage and seed production requires a knowledge of vegetative and reproductive development and their reaction to management. Several management aspects were investigated in the present study, i.e. defoliation, closing date, nitrogen fertilizer application and water stress effects during reproductive development. In addition, an attempt was made to describe more fully phenotypic variation in a Ruzi grass population, an aspect which may provide information useful for a future plant breeding programme in this species.

Phenotypic variation in Ruzi grass was investigated under plastic house conditions where minimum and maximum temperatures were set at 20 C and 30 C respectively. The seed used was from a commercial seedlot obtained from Thailand. Single plant measurements were made during the vegetative stage (herbage production and tiller development, growth form, plant height, canopy width, leaf width, leaf hairiness, ligule colour, and stem colour) and during the reproductive stage (date of first flower initiation, tiller numbers at harvest, and development of inflorescences).

Erect and semi-erect plants had significantly higher herbage production than prostrate plants but failed to show significant differences in tiller numbers. Seed yield was unaffected by plant growth form. Other parameters such as ligule colour, stem colour, leaf width and hairiness also showed considerable variation from plant to plant. However there was little evidence they were linked to any particular plant growth form.

Management practices for herbage and seed production, particularly in terms of defoliation, time of closing and nitrogen fertilizer application were studied in miniature swards in a plastic house at 20 C and 30 C (night and day temperature). These swards were established in wooden bins each with 49 seedlings at a 15 cm square spacing.

In a complete randomised block design, the treatments comprised a combination of cutting frequencies viz. at 20, 35, 50 and 65 cm height and cutting intensities *viz*. 4 and 12 cm

stubble height, with 3 replications. This experiment was designed to determine the most appropriate defoliation management practice for Ruzi grass "pasture", and to provide a reasoned justification for the defoliation strategy adopted in subsequent studies.

Differences in cutting intensity between 4 cm (hard) and 12 cm (lax) had no significant effect on total plant dry matter, but under lax cutting plants produced significantly greater leaf dry matter and a higher leaf:stem ratio. Despite this, herbage quality was not significantly different mainly due to the longer cutting interval under hard cutting conditions resulting in plants producing bigger leaves and a significantly higher LAI compared with lax cutting. Although plants in the longer cutting frequency treatment had a significantly higher LAI than with more frequent cutting, the greater proportion of stem had a major effect in lowering herbage quality.

The overall assessment of the data from the defoliation treatments concluded that the most appropriate defoliation management for a Ruzi sward was 12-35 cm i.e. cutting when the canopy reached a height of 35 cm down to 12 cm. This was the defoliation strategy subsequently employed in studies on the management of Ruzi swards for herbage and seed production.

The effects of nitrogen and "closing date" (cessation of cutting) on herbage and seed production of Ruzi grass, were examined under three nitrogen levels viz. 50, 150 and 250 kg N/ha and three closing dates viz. early (24th March 1997), medium (7th April 1997) and late (21st April 1997) with 3 replications in a complete randomised block design. The experiment was conducted in "miniature swards" as previously described.

Herbage dry matter production prior to closing increased progressively with increasing levels of nitrogen supply. This nitrogen effect continued to produce significantly higher herbage production (250 kg N/ha) even after seed harvest, but there were no significant differences between 50 and 150 kg N/ha. Different closing dates did not cause similar effects to nitrogen application, simply because plant growth rate declined with the approach of the reproductive stage. This was particularly evident in medium and late closing treatments.

However, as expected, the earlier the closing date the higher the amount of herbage dry matter yield obtained after seed harvest. This was mainly contributed by both new vegetative and old reproductive tillers.

Nitrogen application up to 150 kg/ha increased seed yield mainly by increasing total and harvested ripe inflorescence density, total floret numbers, and seed numbers. However nitrogen had no effect on percentage seed set or seed weight. Early and medium closing dates produced significantly greater seed yields than the late closing date mainly through an effect on total and harvested ripe inflorescence density, seed numbers, seed weight. Closing date had no effect on percentage seed set, but early closing resulted in greater inflorescence size (floret numbers/inflorescence). The interaction of nitrogen level and closing date suggested that higher nitrogen supply (150 kg N/ha) and early closing increased the percentage of pure germinating seed, suggesting that this is the most appropriate management for enhancing seed quality and yield.

The final experiment in this study was established to determine the effects of different levels of water stress applied at different stages of reproductive development on seed yield, yield components and seed quality in Ruzi grass. Individual plants were grown in 10 litre pots filled with potting mixture. Three levels of water stress were imposed (control (nil), mild and severe) at three different reproductive development stages (floral initiation, ear emergence and full flowering).

The response of plants to different levels of water application were clearly shown in terms of physiological and morphological changes particularly when these applications were continued throughout the entire reproductive development stage. Although the higher the amount of water applied the greater the dry matter produced, in terms of reproductive development, there was relatively little difference between non stressed and mild stressed plants. Under severe stress, however, although plants developed inflorescences, they were unable to exsert to full ear emergence. Generally, the stage of plant development and level of water stress applied had a bearing on plant dry matter, seed yield and seed yield components.

ACKNOWLEDGEMENTS

I wish to express my deepest gratitude to many people who have each contributed in their own way for their patience, enthusiastic encouragement, guidance and supervision throughout the completion of my study.

I gratefully acknowledge the help and support received from my chief supervisor, Associate Professor Alex Chu for his guidance and suggestions. Although his fully involvement just exited at the final period of my studies, but it was greatly useful to ensure my studies were appropriately finished.

I am greatly indebted to Professor M.J. Hill the Director of New Zealand Seed Technology Institue, Lincoln University (the former Director of Seed Technology Centre at Massey University and also my previous chief supervisor), who helpfully constructed the foundation of my works and continued his support in many useful suggestions and read the manuscript carefully. Thanks are extended to Associate Professor J.G. Hampton, Lincoln University (the former senior lecturer at Massey University and also my co-supervisor) for his constructive guidance and sharing his experiences since the studies commenced.

My sincerest thanks to Professor B.R. Watkin (Professor Emeritus) for his willing helpfulness and highly encouragement provided me with an another opportunity to this further study in New Zealand, and for his patience and wise supervision through the conducting of this research and writing of the manuscript. Without his support my studies would never have been succeeded. I owe him a great debt.

My thanks are also due to Dr Ian Gordon who provided some answers on statistical problems promptly, as did Dr Cory Matthew also commented on some sections and in particular, Mr Craig McGill who always assisted for making computer programmes available. This has been highly appreciated.

I would like to thank Mr Ray Johnston, the manager of Plant Growth Unit (PGU) at Massey

University and his staff, for making available to me the glasshouse facilities and also providing technical assistance. Importantly, the big and very comfortable study room provided at PGU during the writing process is much appreciated. Thanks are extended to Ms Karen Hill, Mr Robert Southward, Ms Collette Gwynne for assisting in different ways.

The friendly environment provided by my Thai friends and fellow graduate students at the Seed Technology Centre, is much appreciated, especially the friendship of five Thai students at STC (Winai Chompukeaw, Jumpa Padrit, Varenya Singkanipa, Uraiwan Supradith and Wasu Amaritsut) will never be forgotten.

I would like to express my sincere thanks to the New Zealand Ministry of Foreign Affairs and Trade for financial assistance during my studies in New Zealand, and also the Dairy Farming Promotion Organisation of Thailand (DPO) for allowing me to undertake this study opportunity especially Mr Sanchai Prasertsuwan the director of DPO and also Mr Pichet Sukpituksakul my boss.

Special thanks to Ms Gay Eustace for providing a wonderful home stay and making my stay in New Zealand a great pleasant throughout the completion of my studies.

Finally, I would like to express my gratitude to my parents, brothers, sisters and also to my girlfriend for their love, understanding and encouragement. To them, I dedicate this thesis.

Narongrit Wongsuwan March, 1999.

TABLE OF CONTENTS

Pa	age

ABSTRACT	· · · · · ·		· · · · · ·	•	v . vii . xiv xviii
TABLE OF CONTENTS. .	· ·		••••	• • •	. vii . xiv xviii
LIST OF TABLES	· ·		 	•	. xiv xviii
	 	•	 	•	xviii
LIST OF FIGURES		•		•	
					. xx
LIST OF PLATES		•			
LIST OF APPENDICES			•••	•	. xx
Chapter 1. INTRODUCTION AND OBJECTIVES	•••	•			1
Chapter 2. LITERATURE REVIEW		•			4
2.1 INTRODUCTION	• •			•	4
2.2 GENERAL DESCRIPTION OF THE SPECIES	• •			•	5
2.3 RUZI GRASS SEED PRODUCTION IN THAILAND				•	8
2.4 EFFECT OF DEFOLIATION MANAGEMENT			•••	•	9
2.4.1 Influence of cutting frequency and intensity on herbage					
production	•••			•	. 10
2.4.2 The effects of defoliation on sward dynamics				• •	. 11
2.4.3 The effects of defoliation on physiological responses of	pla	ant	s.	• •	. 12
2.4.3.1 residual leaf area, light interception and photos	syn	nthe	esis	5	. 12
2.4.3.2 meristematic tissue	••	•			. 13
2.4.4 Processes leading to recovery from defoliation	•	•			13
2.4.5 The effects of defoliation on pasture seed production	•	• •	•	• •	14
2.4.5.1 reproductive apices	•	• •	•	• •	15
2.4.5.2 tillering and tiller development	•	• •	•		16
2.4.5.3 closing date	•	• •	•		17
2.4.5.4 assimilate supply	•	• •	•		18
2.5 EFFECT OF NITROGEN FERTILIZER APPLICATION	•		•		18
2.5.1 On herbage production.	•		•		18

	2.5.1.1 dry matter yield	18
	2.5.1.2 crude protein in herbage	19
	2.5.1.3 time of nitrogen application	19
	2.5.2 On seed production	19
	2.5.2.1 seed yield and yield components	19
	2.5.2.2 species and cultivar	21
	2.5.2.3 seed quality	22
	2.5.2.4 crop matuality	22
2.6 EF	FECT OF WATER STRESS ON SEED PRODUCTION	22
	2.6.1 Physiological and morphological response	22
	2.6.2 Reproductive stages and their sensitivity to water stress	25
	2.6.3 Irrigation practice and benefits for seed crops	27
Chapter 3.	PHENOTYPIC VARIATION IN RUZI GRASS	28
3.1 IN	TRODUCTION	28
3.2 M	ATERIALS AND METHODS	30
	3.2.1 Environmental Conditions And Establishment Of The Trial Plots	30
	3.2.2 Measurements	31
	3.2.2.1 Vegetative stage	31
	(a) herbage production and tiller development.	31
	(b) growth form	31
	(c) plant height and canopy width	33
	(d) leaf width	33
	(e) leaf hairiness	33
	(f) ligule colour.	33
	(g) stem colour	34
	3.2.2.2 Reproductive stage	34
	(a) date of first flower initiation	34
	(b) tiller numbers at harvest	34
	(c) reproductive development of inflorescences	34
	3.2.3 Statistical Analysis.	35

viii

3.3 RESULTS
3.3.1 Vegetative Stage
3.3.1.1 Herbage production and tiller development
3.3.1.2 Growth form
3.3.1.3 Plant height and canopy width
3.3.1.4 Leaf width, Leaf hair, Ligule colour and Stem colour 41
3.3.1.5 Relationship between plant parameters and some
phenotypic variation recorded during the
vegetative growth
3.3.2 Reproductive Stage
3.3.2.1 Date of first flower initiation
3.3.2.2 Tiller numbers at harvest
3.3.2.3 Inflorescence development
3.3.2.4 Inflorescence development and growth form
3.3.2.5 Relationship between plant parameters and some
phenotypic variation recorded during the
vegetative and reproductive growth
3.4 DISCUSSION
3.5 CONCLUSION
Chapter 4. THE EFFECTS OF DEFOLIATION ON HERBAGE AND SEED
PRODUCTION IN RUZI GRASS.
4.1 INTRODUCTION
4.2 MATERIALS AND METHODS
4.2.1 Site and Study Period
4.2.2 Design and Treatments
4.2.3 Measurements
4.2.3.1 Herbage production
4.2.3.2 Seed production
4.2.4 Statistical Analysis
4.3 RESULTS

.

ix

4.3.1 Number of Cut, Cutting Interval and Total Herbage Production 71
4.3.2 Growth and Growth Components
4.3.2.1 Growth rate of sward (total plant)
4.3.2.2 Growth rate of leaf
4.3.2.3 Growth rate of stem
4.3.2.4 Leaf:Stem ratio
4.3.2.5 Leaf area index
4.3.3 Tiller Development
4.3.4 Crude Protein
4.3.5 Seed Production
4.4 DISCUSSION
4.5 CONCLUSION AND RECOMMENDATION

Chapter 5.	EFFECTS OF DIFFERENT NITROGEN APPLICATION LEVELS AND
	CLOSING DATES DURING A DECREASING PHOTOPERIOD ON
	SEED PRODUCTION OF RUZI GRASS
5.1 IN	TRODUCTION
5.2 M	IATERIALS AND METHODS
	5.2.1 Plant Preparation and Study Period
	5.2.2 Design and Treatments
	5.2.3 Measurements
	5.2.3.1 Herbage production
	5.2.3.2 Seed production
	5.2.3.3 Seed quality
	5.2.3.4 Tiller development
	5.2.3.5 Crop index and Harvest index
	5.2.4 Statistical Analysis
5.3 R	ESULTS
	5.3.1 Herbage Production
	5.3.1.1 Until closing
	5.3.1.2 After seed harvesting

	5.3.2 Seed Yield and Yield Components
	5.3.2.1 Actual seed yield
	5.3.2.2 Seed yield components
	(a) inflorescence development
	(b) Thousand seed dry weight (TSDW)
	(c) floret numbers from harvested ripe inflorescences110
	(d) seed number
	(e) percentage seed set
	(f) seed quality
	5.3.3 Tiller Development
	5.3.4 Crop index and Harvest index
5.4 DI	SCUSSION
5.5 CC	DNCLUSION
Chapter 6.	EFFECTS OF WATER STRESS ON SEED PRODUCTION IN RUZI
	GRASS
6.1 IN	TRODUCTION
6.2 M.	ATERIALS AND METHODS
	6.2.1 Plants and Experimental Site
	6.2.2 Design and Treatments
	6.2.3 Measurements
	6.2.3.1 Leaf extension rate (LER)
	6.2.3.2 Relative leaf water content (RWC)
	6.2.3.3 Reproductive development
	6.2.3.4 Seed harvesting and data collection
	6.2.3.5 Percentage of Pure Germinating Seed (%PGS) and PGS
	yield
	6.2.3.6 Plant dry weight and dry weight partitioning
	6.2.3.7 Tiller development
	6.2.3.8 Harvest index

xi

	6.3.1 LER and RWC
	6.3.2 Reproductive Development
	6.3.3 Seed Yield and Yield Components
	6.3.3.1 Seed production at harvest
	(a) calculated seed yield
	(b) harvested ripe inflorescence numbers
	(c) total inflorescence numbers produced per plant146
	(d) the percentage of harvested ripe inflorescences
	and partitioning of total inflorescence numbers 147
	(e) harvested seed numbers
	(f) Thousand seed dry weight (TSDW)
	(g) floret numbers at seed harvest
	(h) percentage seed set
	(i) deformed inflorescence numbers
	6.3.3.2 Percentage of Pure Germinating Seed (%PGS) and PGS
	yield
	6.3.4 Plant Dry weight and Dry Weight Partitioning
	6.3.4.1 Total plant dry weight
	6.3.4.2 Leaf dry weight
	6.3.4.3 Stem dry weight
	6.3.4.4 Leaf area
	6.3.4.5 Leaf : Stem ratio
	6.3.5 Tiller Development
	6.3.6 Harvest Index
6.4 DI	SCUSSION
6.5 CC	ONCLUSION
Chapter 7.	GENERAL DISCUSSION AND CONCLUSIONS
7.1 IN	TRODUCTION
7.2 PH	ENOTYPIC OF PLANT POPULATION
	7.2.1 Comparison of plant population

7.2.2 Plant performance under spaced plant and miniature swards
conditions
7.2.3 Plant selection and recommendations
73 THE MANAGEMENT PRACTICES FOR HERBAGE AND SEED
PRODUCTION
7.3.1 Effect of defoliation
7.3.2 Effect of closing pasture
7.3.3 Nitrogen fertilizer
7.4 THE EFFECTS OF WATER STRESS ON SUBSEQUENT
REPRODUCTIVE DEVELOPMENT AND SEED YIELD
7.4.1 The sensitivity of vegetative growth to plant water stress
7.4.2 The sensitivity of seed yield and yield components to plant
water stress
7.5 CONCLUSIONS
APPENDICES
REFERENCES

xiii

LIST OF TABLES

CHAPTER 3.

Table 3.1	Herbage production (plant DW, leaf DW, stem DW), leaf:stem ratio, leaf
	area and tiller numbers over 4 cuts during vegetative growth
Table 3.2	The range, mean and coefficients of variation (%CV) of the 72 Ruzi
	plants after 4 cuts
Table 3.3	Number and percentage of plants with different growth form in the
	population
Table 3.4	Herbage production parameters in different each growth form group 40
Table 3.5	Mean plant height and canopy width (cm) in different plant growth form
	group
Table 3.6	Leaf width, leaf hair, ligule colour and stem colour characteristics 41
Table 3.7	Leaf width and hairiness of plants in each plant growth form group 42
Table 3.8	Ligule colour and stem colour in each plant growth form group 43
Table 3.9	Simple correlation coefficients (r ²) between plant parameters and
	some phenotypic variation recorded during the vegetative growth 44
Table 3.10	Number of plants and percentage showing first flower initiation 45
Table 3.11	Reproductive (old, new and total) and vegetative tillers and percentage
	at harvest
Table 3.12	Mean inflorescence numbers and components
Table 3.13	The range, mean and coefficients of variation (%CV) of 48 Ruzi
	plants
Table 3.14	Inflorescence development in each plant growth form group
Table 3.15	Simple correlation coefficients (r^2) between plant parameters and
	some phenotypic variation recorded during the vegetative and
	reproductive growth

CHAPTER 4.

Table 4.1	The management programme in this study
Table 4.2	Number of cuts, cutting interval and total herbage production in
	different defoliation treatments
Table 4.3	Mean growth rate of sward (kgDM/ha/day)
Table 4.4	Mean growth rate of leaf DM (kg/ha/day)
Table 4.5	Mean growth rate of stem DM (kg/ha/day)
Table 4.6	Mean DM leaf:stem ratio
Table 4.7	Mean leaf area index
Table 4.8	Early, medium and late tiller development in each treatment during
	herbage production
Table 4.9	Mean crude protein percentage
Table 4.10	Mean calculated seed yield (kg/ha)
Table 4.11	Ranking of different parameters $(1 = highest; 8 = lowest)$ in each
	cutting treatment

CHAPTER 5.

Table 5.1	The number of cuts and cutting interval for the different closing
	treatments
Table 5.2	Range of harvest dates in different nitrogen and closing date treatments . 97
Table 5.3	Total dry matter herbage production from first cut to closing date in
	different nitrogen and closing date treatments (kg/ha)
Table 5.4	Mean herbage dry matter production from closing date to final harvest
	in different nitrogen and closing date treatments (kg/ha)
Table 5.5	Relationship between nitrogen application and closing date on tiller dry
	matter (kg/ha) originating from old and new vegetative and
	reproductive tillers
Table 5.6	Mean leaf:stem ratio after seed harvest in different nitrogen and closing
	date treatments

Table 5.7	Mean leaf area index after seed harvest in different nitrogen and
	closing date treatments
Table 5.8	Mean total seed yield obtained from different nitrogen and closing
	date treatments (kg/ha)
Table 5.9	Total inflorescence numbers from different nitrogen and closing date
	treatments (m ²) \ldots
Table 5.10	Mean harvested inflorescence numbers from different nitrogen and
	closing date treatments (m^2)
Table 5.11	Mean unharvested inflorescence numbers from different nitrogen and
	closing date treatments (m^2)
Table 5.12	Comparison of mean inflorescence numbers produced by old and new
	tillers in different nitrogen and closing date treatments
Table 5.13	Mean TSDW at harvest from different nitrogen and closing date
	treatments (g)
Table 5.14	Mean total floret numbers at harvest from different nitrogen and
	closing date treatments (no/m^2)
Table 5.15	Mean total numbers of seeds harvested from different nitrogen and
	closing date treatments (no./m ²)
Table 5.16	Mean percentage seed set from different nitrogen and closing date
	treatments (%)
Table 5.17	Mean percentage pure germinating seed from different nitrogen and
	closing date treatments (%)
Table 5.18	Mean yield of pure germinating seed from different nitrogen and
	closing date treatments (kg/ha)
Table 5.19	Tiller numbers from different nitrogen and closing date treatments at
	the time of the first cut, at the early closing date and at the final cut116
Table 5.20	Tiller partitioning between old and new vegetative and reproductive
	tillers at the final cut
Table 5.21	Crop index in different nitrogen and closing date treatments (%)118
Table 5.22	Harvest index in different nitrogen and closing date treatments (%)118

.

xvi

CHAPTER 6.

Table 6.1	Water stress treatments used in this study
Table 6.2	The number of days from the commencement of water stress (FI) to
	the completion of seed harvest (HI) or final cut, and for the transition
	periods FI-EE, EE-FF, and FF-SH
Table 6.3	Effect of water stress on seed yield (g/plant)
Table 6.4	Effect of water stress on harvested ripe inflorescences per plant and
	percentage of total inflorescences harvested
Table 6.5	Effect of water stress on total inflorescence numbers produced per plant.147
Table 6.6	Effect of water stress on total inflorescence numbers partitioned
	into inflorescences borne on old basal tillers (OBT), old aerial tillers
	(OAT), new basal tillers (NBT) and new aerial tillers (NAT)
Table 6.7	Effect of water stress on harvested seed numbers
Table 6.8	Effect of water stress on TSDW (g)
Table 6.9	Effect of water stress on harvested floret numbers
Table 6.10	Effect of water stress on seed set per plant (%)
Table 6.11	Effect of water stress on the number of deformed inflorescences per
	plant
Table 6.12	Effect of water stress on pure germinating seed (%PGS) and PGS yield
	(g/plant)
Table 6.13	Effect of water stress on total plant dry weight (g/plant)
Table 6.14	Effect of water stress on leaf dry weight (g/plant)
Table 6.15	Effect of water stress on stem dry weight (g/plant)
Table 6.16	Effect of water stress on leaf area (cm ² /plant)
Table 6.17	Effect of water stress on leaf:stem ratio
Table 6.18	Effect of water stress on tiller numbers per plant
Table 6.19	Effect of water stress on harvest index (HI)

LIST OF FIGURES

CHAPTER 3.

Figure 3.1	A comparison of individual total plant dry weights, including leaf and
	stem dry weight after four successive cuts
Figure 3.2	A comparison of individual plants development after four successive
	cuts in terms of (a) Total Leaf Area, (b) Mean Leaf:Stem ratio and (c)
	Mean Tillers
Figure 3.3	Date of first flower initiation in the Ruzi plant population
Figure 3.4	The pattern of inflorescence development per plant per day based on
	first flower initiation
Figure 3.5	A comparison of inflorescence development per plant per day from
	different plant growth forms and means of each growth form 48
Figure 3.6	A comparison of tiller numbers at harvest and mean tiller numbers
	produced over the vegetative stage
Figure 3.7	A comparison of mean inflorescence numbers per plant from
	different reproductive tillers
Figure 3.8	A comparison of mean floret numbers per inflorescence per plant
	from different reproductive tillers
Figure 3.9	The comparison of mean raceme numbers per inflorescence per plant
	from different reproductive tillers
Figure 3.10	A comparison of mean floret numbers per plant from different
	reproductive tillers

CHAPTER 4.

Figure 4.1	Plant layout per bin
Figure 4.2	The layout of treatment defoliation study
Figure 4.3	Effect of cutting intensity (a) and frequency (b) on numbers of cuts and
	cutting interval (days). Vertical bars show standard error of the mean 72

Figure 4.4	Tiller development at each of the cutting intensities and frequencies used	
	in the defoliation study. Vertical bars show standard error of the mean .	77
Figure 4.5	A comparison of tiller number development at the date of closing and	
	final tiller number at seed harvest. Vertical bars show standard error	
	of the mean	78
Figure 4.6	Relationship between seed yield and tiller number at seed harvest.	
	$Y = -104.767 + 0.66615*x, r^2 = 70.64$	82

CHAPTER 5.

Figure 5.1	Effect of nitrogen fertilizer and closing date on total dry matter
	(DM) production from the first cut to closing (DM until closing) and
	after seed harvest (DM after seed harvest). Vertical lines indicate
	the standard error (SEM) of the mean (total dry matter)
Figure 5.2	The relationship between seed yield and ripe inflorescence number at
	seed harvest. $Y = 79.9523 + 0.8501*x$, $r^2 = 87.16$
Figure 5.3	Effect of nitrogen fertilizer and closing date on percentage of harvested
	and immature inflorescences
Figure 5.4	Effect of nitrogen fertilizer and closing date on floret number per
	inflorescence (harvested ripe inflorescence) Vertical lines indicate the
	standard error of the mean. $\ldots \ldots \ldots$
Figure 5.5	Effect of nitrogen fertilizer and closing date on nitrogen efficiency (kg
	pure germinating seed per kg nitrogen applied). Vertical lines indicate
	the standard error of the mean

CHAPTER 6.

•

Figure 6.1	The effect of water stress on (a) Leaf Extension Rate and (b) Leaf
	Relative Water Content. Vertical bars show standard error of the mean .142
Figure 6.2	Effect of water stress on tiller development viz. old main tillers (OMT),
	new main tillers (NMT) and vegetative tillers (VT). Vertical bars
	show standard error of the mean

LIST OF PLATES

CHAPTER 3.

Plate 3.1 Plant growth form categories (a) erect, (b) semi-erect, and (c) prostrate . 32

CHAPTER 4.

Plate 4.1Miniature 'swards' showing effect of removal of guard rows before
cutting (a); plants after cutting to 12 cm (b) or 4 cm (c). Plate (d)
shows the method of plant support to prevent lodging 70

CHAPTER 6.

Plate 6.1	General view of plants used in the water deficit study (a); initiated leaf
	used for leaf extension measurement (b); and the plant support system
	used to prevent lodging (c)
Plate 6.2	Examples of deformed (non) fully exserted inflorescences in severe
	water stress treatment (a, b, c); rewatering recovery of inflorescences
	(d, e)
Plate 6.3	Examples of differences in dry matter production according to the level
	of water supply. Full water throughout (WWW); mild stress $(D_1D_1D_1)$
	and severe stress $(D_2D_2D_2)$

LIST OF APPENDICES

Appendix 4.1	Method of calculating herbage production, leaf:stem ratio and leaf
	area index

xx

Appendix 5.1	Mean tiller dry matter production (kg/ha) of new reproductive	
	tillers in different nitrogen and closing date	183a
Appendix 5.2	Mean inflorescence numbers (numbers/m ²) produced by new tillers	
	in different nitrogen and closing date	183a
Appendix 5.3	Mean tiller numbers at final cutting (tiller numbers/m ²) in different	
	nitrogen and closing date	183b
Appendix 5.4	Mean new reproductive tiller numbers (tiller numbers/ m^2) in	
	different nitrogen and closing date	183b