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MODE OF ACTION OF DOTHISTROMIN

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ABSTRACT

Dothistronin is a bright orange-red pignent produced by the pine-needle pathogen <u>Dothistrona pini</u> Hulbary, the causal agent of a necrotic disease known as dothistronal blight. This compound, implicated as a fungal toxin, has been isolated both from laboratory cultures of <u>D. pini</u> and from infected <u>Pinus radiata</u> foliage.

Detailed chemical investigation by Gallagher (1971) showed that dothistronin is a tri -a - hydroxyanthraquinone fused to a substituted tetrahydrobifuran ring system. The bifuran ring moiety is incorporated in other fungal metabolites including the extremely toxic and carcinogenic aflatoxin compounds. There is an increasing body of evidence to suggest that these fungal metabolites share a common biosynthetic origin.

Bioassay has demonstrated the toxicity of dothistronin to the unicellular green alga, <u>Chlorella pyrenoidosa</u>, and to tissue cultures of <u>P.attermata</u>. The very low level of solubility of the compound in aqueous solutions has precluded bioassay using pine seedlings. This thesis reports an investigation of the biochemical changes induced by dothistronin in microbiological systems.

In the course of this investigation dothistronin has been shown to be toxic to a range of microorganisms in addition to its known toxicity to Chlorella pyrenoidosa. These studies have suggested possible ways of increasing the sensitivity of bioassays for dothistronin.

It was found that the addition of dothistronin to liquid cultures of <u>Chlorella</u> as an ethyl acetate solution caused reproducible levels of inhibition, provided that the ethyl acetate concentration was less than 0.5%. Batch culture techniques were used to establish the levels of dothistronin required for inhibition of growth of <u>Chlorella</u>. The ratio of dothistronin concentration to cell number was found to be an important factor in the inhibition response.

Utilization of synchronous culture techniques permitted the study of biochemical changes induced by dothistronin throughout the cell cycle of <u>Chlorella</u>. Results showed a marked inhibition of the rate of increase of total protein and RNA over the cell cycle with no significant alteration of the rate of DNA increase.

A dose-response curve for dothistronin inhibition of growth of <u>Chlorella</u> was established and a more detailed investigation of the action of dothistronin in inhibiting growth was undertaken using radioactive isotopes. By this means it was shown that ³H-uridine and ¹⁴C-phenylalanine incorporation into cell naterial

is inhibited within 30 mins of exposure to the toxin. Difficulties encountered in attempts to obtain satisfactory incorporation of label into <u>Chlorella DNA-fractions</u> prevented further investigation of the effect of dethistronin on DNA synthesis in this organism. This led to the investigation of other microorganisms as more suitable experimental systems for this study.

Bacillus negaterium KM, proved to be very sensitive to dothistronin and showed rapid incorporation of radioactive isotopes into protein and nucleic acid fractions. Growth curves established that the inhibitory ratios of dothistromin concentration to cell numbers for this organism were in the order of 0.25 $\mu g/\text{cell} \times 10^8$ (as compared to 2.0 mg/cell X 10⁸ for Chlorella). At this concentration, over the 30 min time course studied, dothistromin had no effect on the incorporation of 3H-thypidine into the DNA fraction. Inhibition of ³H-uridine incorporation was evident at 6 min and very marked by 10 min while the inhibition of 14 C-phenylalanine incorporation into protein was not evident until considerably later. The effects of dothistronin in this system were compared with those of antibiotics with known sites of action. Dothistronin inhibition of ³H-uridine incorporation has a similar time course to that shown by actinonycin D. although marked inhibition by Actinonycin D is evident at 3 min. whereas dothistromin inhibition is not noticeable until 6 min.

On the basis of these results it is suggested that dothistromin interferes with RNA synthesis and that the observed inhibition of protein synthesis is a secondary effect of this impairment.

Confirmation of dothistromin action 'in situ' by administration of the compound to pine seedlings is necessary before any definitive statement can be made concerning its role in dothistromal blight.

However these results indicate the possible importance of dothistromin in pathology of dothistromal needle blight of pines.

Impairment of the RNA synthetic capacity in pine needle tissue by the toxin could rapidly lead to cell death and to the necrosis of needle tissue observed in diseased foliage.

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