Copyright is owned by the Author of the thesis. Permission is given for a copy to be downloaded by an individual for the purpose of research and private study only. The thesis may not be reproduced elsewhere without the permission of the Author.

Characterization of *Arabidopsis thaliana* CPR5 via the Elucidation of Interacting Protein Partners

A thesis presented in partial fulfilment of the requirements for the degree of

Masters of Science in Biochemistry

At Massey University, Palmerston North, New Zealand



Fiona (Shane) Chiem 2015

Abstract

The *Arabidopsis thaliana* Constitutive expresser of pathogenesis related genes5 (*CPR5*) has previously been suggested to play a role in the regulation of disease resistance, plant and cell proliferation, development and death. Analysis of *cpr5* mutant alterations to hormone and hormone-like signalling mechanisms have provided evidence that abolishment of *CPR5* involvement within these hormone signalling pathways, results in many of the stunted growth, early senescence and constitutive expression of pathogen defense phenotypes observed. Despite the pleiotropic effect that *cpr5* mutants have on the plant system, it is unclear whether *CPR5*-dependent pathways are due to a direct interaction with *CPR5* or due to a more indirect association. *CPR5* has been proposed to be a regulator of a multitude of different pathways, including reactive oxygen species (ROS), cell wall biosynthesis, and transcription but evidence of these proposals are limited to the effects that *cpr5* mutants have on downstream targets.

In an attempt to address the involvement of CPR5 in *Arabidopsis* plant processes, a series of studies were conducted to determine the protein interacting partners of CPR5. Proteins were identified via 2 independent yeast 2 hybrid (Y2H) screening of an *Arabidopsis* transcriptome library. Ten proteins of interest were identified via two independent screenings using two truncated forms of CPR5. Functional involvement of CPR5 with the identified proteins was further explored using the Y2H pairwise interaction system. CPR5 was found to interact with 3 full length proteins identified.

To explore the possibility that CPR5 interacts with multiple protein partners in different locations within the cell, Bifluorescence molecular complementation assays were performed to determine the localization and interaction of CPR5 with the ten identified genes as well as 3 previously identified genes. Several novel interactions were identified that occur within the nucleus and outside of the nucleus. Not only was CPR5 confirmed to have an interaction with KRP2 within the nucleus, CPR5 exhibited interaction with FSD1, CRK4, PATL3, PATL5, and PATL6, outside of the nucleus.

In the final set of experiments, several double mutant lines were produced that did not yield any observable phenotypes that differ from *cpr5-2* single mutant plants. In order to determine the effects these double mutants have on various plant processes affected by *cpr5-2* single mutant; qRT-PCR was performed to determine the expression pattern of pathogen related genes (*PR1* and *PDF1.2*) known to be significantly upregulated in *cpr5-2* plants. qRT-PCR analysis revealed that *cpr5-2 fsd1* exhibits a down-regulation of *PDF1.2*.

PR1 regulation was found to be down-regulation in *cpr5-2 bzip61* and up-regulated in *cpr5-2 patl3* compared to *cpr5-2*.

Sugar and dark treatment of the *cpr5-2* double mutant lines yielded several alterations to hypocotyl length, root length, and apical hook curvature by several of the double mutant lines, indicating a connection between CPR5 and the knocked out gene of interest. None of the double mutants were able to completely rescue the sugar-induced morphological phenotypes exhibited by *cpr5-2*, and some double mutant lines exhibited more pronounced effects indicating an additive effect by sugar treatment.

Together this data suggests that CPR5 interacts with various proteins involved in different plant processes in various locations throughout the cell. Further research of these proteins and a more direct analysis of the interaction that may occur between CPR5 and these proteins will be required to provide a foundation for more direct characterization the *CPR5* molecular function; and ultimately to determine the role that CPR5 plays within the hormone and hormone like signalling pathway and their effects on major plant processes.

Acknowledgements

I would like to thank my supervisor Dr. Paul Dijkwel for giving me this opportunity and for helping me complete my Masters degree. Paul has been an exemplary supervisor providing me with the guidance I required when help was asked and for challenging me. I thank you for the patience you have shown me and for providing me with the independence I required to succeed. Thank you especially for your confidence in my abilities to successfully accomplish the goals we set forth and for allowing me to carry out these goals in my own unorthodox fashion.

I would like to thank Elizabeth Jennens for always believing in me and for giving me the love, space, and time to complete my degree. Without you I would not have started my Masters and I would not be where I am in my life without you.

I would like to thank Prof. Michael McManus and all of my C5.19 lab mates for their critiques and insights and for listening when I needed help with my research. In particular, I would like to thank my family away from home, Jay Jayaraman and Srishti Joshi, for their continual help academically and personally. I cannot imagine my time at Massey without your unwavering friendships, and willingness and dedication to helping me in all aspects of my life.

To my parents, family, and friends, I am forever grateful for your unending encouragement as I continued my studies around the world, and for your understanding as I concentrated on finishing my thesis. Knowing that you are all waiting for me to come home gave me the motivation I needed to return to you having successfully completed another chapter of my life.

I would most especially like to thank my mother, Sandy Chiem, for allowing me to be my independent self despite how much it hurt to let me travel in distance and on my own path of life. You have shown me such unrelenting selfless love and support regardless the decisions I make, and it is for you and because of you that I am inspired to be the best version of myself.

Abbreviations

minutes
seconds
Ade Adenine
aa amino acids
amp Ampicillin

BiFC BiFluorescence Molecular Complementation

BLAST Basic logical alignment search tool

bp Base-pair

cDNA DNA synthesized from an mRNA template

C-terminus (at the) carboxy-terminal end of a polypeptide chain

(terminal)

CPR5 CPR5 wild-type gene
 CPR5 CPR5 wild-type protein
 cpr5 CPR5 mutant gene
 cpr5 CPR5 mutant protein

cpr5-2 cpr5 mutant line with mutation at aa420 (W->stop)

DAPI A DNA binding fluorescent stain ((4',6-diamidino-2-phenylindole)

DNA Deoxyribonucleic acid DNase Deoxyribonuclease

dNTP 2'-deoxynucleotide 5' triphosphate

dH2O distilled water

ddH2O double distilled water

E. coli Escherichia coli

EDTA Ethylenediaminetetraacetic acid

FW Fresh weight

g Gram

gDNA Genomic DNA Gen Gentamycin

h Hour His Histidine

IPTG Isopropyl-β-D-thiogalactopyranoside

kankanamycinkbKilo base-pairkD(a)Kilo daltons

L Litre

LB Luria-Bertani (media or broth)

Leu Leucine

M Molarity (moles per litre)
MCS Multiple cloning site

mg Miligram

Milli-Q-water Water purified by Milli-Q-ion exchange chromatography

ml Milliters

mol Mole (Avagadro's number)

mRNA Messenger RNA

MS Murashige & Skoog Media

NCBI National Centre for Biotechnology Information

ng Nanogram

OD600 optical density at 600nm (measured in a spectrophotometer)

°C Degree celsius

PAGE Polyacrylamide gel electrophoresis

PBS Phosphate buffer saline
PCR Polymerase chain reaction

pH -Log (H+)

psi a unit of pressure (pounds per square inch)
qRT-PCR Reverse transcriptase-polymerase chain reaction

RE Restriction Enzyme
Rnase Riboxynuclease
RO Reverse osmosis

rpm revolutions per minute

SALK Arabidopsis T-DNA insertion lines from the SALK Institute, a non-

profit research organization

SD Synthetic Defined (media)
SDS Sodium Dodecyl Sulfate
SEM Standard error mean

TAE Tris base, acetic acid, and EDTA buffer
TAIR The Arabidopsis Information Resource

TE Tris base, EDTA buffer

Tet Tetracycline

Tm Melting temperature at which DNA strands separate prior to annealing

Tris (hydroxymethyl) aminomethane

Trp Trpytophan

Tween-20 Polyoxyethylenesorbitan monolaurate

U Unit (based on enzyme activity)

μg Microgram μl Microlitre μM Micromolar

V Volt

v/v Volume per volume w/v Weight per volume w/w Weight per weight

X- α -Gal X- α -Gal is a chromogenic substrate used to detect α -galactosidase

activity

Y2H Yeast-2-hybrid

YFP Yellow fluorescent protein

YPDA yeast peptone dextrose adenine (media/agar)

List of Figures

Figure 1.1: Predicted CPR5 Involvement in Plant Processes4
Figure 1.2: Diagram of CPR5 putative protein structure and location of several
identified cpr mutant alleles 13
Figure 2.1: Cassette Set Up For Western Blotting Adapted From Mini Trans-Blot
Electrophoretic Transfer Cell, Instruction Manual (Bio-Rad Laboratories, Hercules,
CA, USA)24
Figure 2.2: Bait Protein Plasmid Constructions25
Figure 2.3: Plasmid Construction for BiFC Assay 33
Figure 3.1: The Theory of Y2H assay43
Figure 3.2: Western Blot Analysis of BD-CPR5TM0 and BD-CPR5TM145
Figure 3.3: Representative Plate of Yeast Grown on SD/-His/-Trp/-Leu/-Ade/X-A-Gal
and Subsequent Screening Plate48
Table 3.1: Genes Identified in CPR5TM0/CPR5TM1 Y2H Library Screenings49
Figure 3.4: Protein Pair Y2H Transformation Assays of Constructs Rescued from Y2H
Screening
Figure 3.5: Protein Pair Y2H Transformation Assay on Medium Stringency Media . 52
Figure 3.6: Protein Pair Y2H Transformation Assay on High Stringency Media 53
Figure 3.7: Heat Map Summary of Interactions Identified via Y2H and BiFC 54
Figure 3.8: The Theory of BiFC assay57
Figure 3.9: EDS1-YFP Fluorescence in the cytoplasm and nucleus59
Figure 3.10: BiFC Signal form N. benthamiana Expressing N-terminal tagged, C-
terminal-YFP-CPR5 and N-terminal tagged, N-terminal-YFP-GOI
Figure 3.11: BiFC Assay was carried out in <i>N. benthamiana</i> Expressing C-terminal
tagged, C-terminal-YFP-CPR5 and N-terminal tagged, N-terminal-YFP-GOI
Figure 3.12: Representative Arabidopsis Plants Grown for 21 Days under Normal
Short Day Conditions
Figure 3.13: Representative Drought Tolerance Phenotypes of 5-Week Old
Arabidopsis Mutant and Wild Type Seedlings
Figure 3.14: Arabidopsis crp5-2 and Col-0 Seedlings under Dark and Sugar
Treatment
Figure 3.15: Representative Arabidopsis Seedlings from Dark and Sugar Treatment
for 3 and 5 Days
Figure 3.16: Hypocotyl Length of 3 and 5 Day Old Dark and Sugar Treated Seedlings
Figure 3.17: Root Length of 3 and 5 Day Old Dark and Sugar Treated Seedlings 78
Figure 3.18: Apical Hook Curvature of 3 and 5 Day Old Dark and Sugar Treated
Seedlings79
Figure 3.19: Heat Map Summary of Arabidopsis Seedlings Grown under Dark and
Sugar Treatment for 3 and 5 Days80
Figure 3.20: Expression of Housekeeping Genes, At2G31270, AtTUB5 and AtUBC9
Across All Lines Investigated81
Figure 3.21: Transcriptional Changes of Defense-Related Genes PR1 and PDF1.2 in
Arabidopsis Mutant Plant Lines83
Figure 4.1: CPR5 predicted phosphorylation sites108
Figure 4.2: Proposed Model of CPR5 Dependent Regulation of Plant Processes via
Putative Direct Interaction with Identified111

List of Tables

	٠.
Table 2.2: List of Primers used for colony PCR and sequencing of genes cloned in	ω
Y2H AD- and BD- plasmid	26
Table 2.3: Genes and sequences of primers used for amplifying full length gen	ne
coding regions for cloning into Y2H GAL4-AD vector pGADT7	30
Table 2.4: Summary of Arabidopsis Plant Lines and Parent Lines	35
Table 2.5: Primer Sequences used for q-RT-PCR (A. thaliana)	37
Table 2.6: Primer Sequences for Genotyping of Arabidopsis T-DNA SALK Lines	39
Table 3.1: Genes Identified in CPR5TM0/CPR5TM1 Y2H Library Screenings4	19

Table of Contents

Abstract	i
Acknowledgements	iii
Abbreviations	iv
List of Figures	vi
List of Tables	vii
Chapter 1: Introduction	1
1.1 General Introduction to CPR5	1
1.2 Phytohormones Signalling	3
1.3 Plant Germination and Development	5
1.4 Plant Senescence	7
1.5 Resistance and Hypersensitive Response-Mediated PCD	8
1.6 Reactive Oxygen Species (ROS)	11
1.7 CPR5 Molecular Function: Protein Localization and Structural Analysis	12
1.7 CPR5 and Cell Cycle Involvement	14
1.8 Concluding Remarks	15
Chapter 2: Materials and Methods	17
Chemicals used	17
2.1 General Use Protocols:	17
2.1.1 Bacterial Propagation	17
2.1.2 Preparation of Plasmid DNA (Alkaline Lysis Miniprep)	18
2.1.3 Agarose Gel Electrophoresis	18
2.1.4 Preparation of Chemically Competent Bacterial Strains	19
2.2 Cloning 19	
2.2.1 PCR Amplification of cDNA	19
2.2.2 Restriction Digestion and DNA Ligation	20
2.2.3 Bacterial Chemical Transformation	20
2.2.4 Bacterial Colony PCR	21
2.2.5 DNA Sequencing	21
2.3 SDS PAGE and Western Blot Analysis	22
2.3.1 Protein Extraction and Sodium Dodecyl Sulfate (SDS) Polyacrylamide Gel	
Electrophoresis (PAGE)	22
2.3.2 Transfer of Protein onto PVDF Membrane	24
2.4 Yeast-Two-Hybrid (Y2H)	24
2.4.1 Generating Bait Plasmids for Y2H "Mate and Plate" Library Screening	25

2.4.2 Yeast Transformation	26
2.4.3 Y2H Transcriptome Library Mating Assay	26
2.4.4 Yeast Colony PCR Analysis	27
2.4.5 Yeast Plasmid Extraction and Rescue	28
2.4.6 Generating "Prey" Plasmids for Y2H Protein Pair Assays	28
2.4.7 Y2H Protein Pair Transformation Assays	31
2.5 Bimolecular Fluorescence Complementation (BiFC)	32
2.5.1 Generating Plasmids for BiFC and Transformation into Agrobacterium	
tumefaciens	32
2.5.2 Agrobacterium tumefaciens Transformation and Infiltration	33
2.5.3 Confocal Microscopy	34
2.6 Quantitative Real-Time PCR (qRT-PCR)	34
2.6.1 Isolation of total RNA and Quantification	34
2.6.2 cDNA Synthesis	36
2.6.3 qRT-PCR Amplification	36
2.6.4 qRT-PCR Statistical Analysis	37
2.7 Plant Propagation and Harvesting Methods	38
2.7.1 Plant Genetic Crosses	38
2.7.2 Plant Dark and Sugar Treatment	40
2.7.3 Plant Propagation for Morphological Studies	40
2.7.4 Plant Drought Tolerance	40
Chapter 3: Results	41
3.1 Yeast-Two-Hybrid Identification of Protein-Protein Interactions	41
3.1.1 Introduction to Yeast Two Hybrid	41
3.1.2 Cloning of CPR5 into Y2H GAL4-BD Vector	43
3.1.3 Expression of GAL4-BD Fusion Proteins	44
3.1.4 Establishment of Protein Pair Y2H Mating Assays	46
3.1.5 Identification of Novel CPR5 Proteins Interactions	47
3.1.6 Examination of Physical Interactions Of CPR5TM0/CPR5TM1	51
3.1.7 Summary of Y2H Results	54
3.2 BiFC Assay	55
3.2.1 Introduction to Bimolecular Fluorescence Complementation	55
3.2.2 Examination of Physical Protein Interactions via BiFC	57
3.2.3 Summary of BiFC Results	61
3.3 Changes in <i>cpr5-2</i> Morphological Phenotypes	
3.3.1 Trichomes and Lesions	63

3.3.2 Drought Tolerance	65
3.3.3 Dark and Sugar Treatment	67
3.4 qRT-PCR	81
3.4.1 Identification of Stable Housekeeping Genes	81
3.4.2 qRT-PCR of PDF1.2 and PR1 in Arabidopsis Lines	82
Discussion	84
4.1 Interaction Studies Identified and Confirmed Protein Interactions	85
4.1.1 Y2H Identification of CPR5 Potential Protein Partners and Interacting	
Domains	85
4.1.2 BiFC Identified 6 different Protein-Protein Interactions in planta	89
4.2 Effect of mutation of potential interacting proteins on cpr5-2 hypersen	sivity to
sucrose	91
4.2.1 Effects of Dark Treatment on cpr5-2 Double Mutant Seedlings	91
4.2.2 CPR5 and BZIP61 Regulation of Root Elongation in Response to Exogen	ous
Application of Sugar	91
4.2.3 CPR5 Regulation of AKIN10 Sugar Starvation-dependent Activation	92
4.2.4 Uncoupling CPR5 Hypersensitivity to Sugar –Future Outlooks	94
4.3 Identification of Plant Processes Relating to CPR5 Interaction with the Id	dentified
Proteins of Interest	96
4.3.1 CPR5 may Confer Drought Tolerance through Interaction with Heatsho	ck
Protein DNAJ	96
4.3.2 CPR5 as a Putative Mediator of ROS via Interaction with a Superdismut	ase97
4.3.3 CPR5 may Modify PI Signalling through Interaction with SAC9 and Pate	ellins99
4.3.4 CPR5 Involvement in Transcriptional Regulation.	101
4.4 CPR5-The Grand Scheme of Things	103
4.4.1 CPR5 Upstream Regulation of Downstream Targets	103
4.4.2 CPR5 Functionality in and Outside of the Nucleus	104
4.4.3 CPR5 Post-Translational Modification	106
4.5 Summary of Discussion	109
Appendices	112
Appendix 1. CPR5 Coding Sequences	112
CPR5:112	
CPR5TM0:	113
CPR5TM1:	113
Appendix 2. Plasmid Vector maps for Y2H Cloning	114

B) pGADT7-RecAB	114
C) pGADT7	115
Appendix 3. Restriction Sites used for Y2H	116
Appendix 4. YFP Coding Sequences	117
nYFP 117	
cYFP 117	
Appendix 5 pGreenII 0029 62 SK plasmid vector map for BiFC cloning and p	pSOUP
helper plasmid	118
Appendix 6 Primers and Cloning Restriction Sites for BiFC Cloning	119
6A: Primers for BiFC Cloning of p(HA:;cYFP), p(cYFP::HA) and p(nYFP::HA)	119
6B: Primers for BiFC Cloning of plasmids p(cYFP::HA::GOI-X), p(GOI-X::HA::cYI	FP),
and (p(nYFP::HA::GOI-Y)	120
6C: Restriction enzymes for BiFC Cloning of p(HA:;cYFP), p(cYFP::HA) and	
p(nYFP::HA)	121
6D: Restriction enzymes for BiFC Cloning of p(cYFP::HA::GOI-X), p(GOI-	
X::HA::cYFP), and (p(nYFP::HA::GOI-Y)	
Appendix 7 TAIR Ascension and obtained SALK lines	
References	124