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THE CELLULAR AND CLINICAL PATHOLOGY OF CLOSTRIDIUM PERFRINGENS TYPE D ENTEROTOXAEMIA

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at

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The Cellular and Clinical Pathology of Cl. perfringens type D Enterotoxaemia.

Abstract.

Enterotoxaemia of sheep is caused by the absorption of the epsilon toxin of <u>Cl. perfringens</u> type D. In preliminary experiments it was found that the sex and nutritional status of mice could influence the outcome of experiments in which this toxin was used. Female mice tended to survive longer than males and food deprivation caused the accumulation of free lipid in the proximal tubules of the kidneys of control, as well as intoxicated, mice.

It appears that there may be species differences in the morphological changes which epsilon toxin induces in the intestinal mucosa. Severe inflammatory necrosis occurred in the mucosa of toxin-containing loops of rabbit intestine but was absent from similar loops of lamb intestine from cases of experimentally induced enterotoxaemia.

While the toxin itself is fairly stable in vitro it may deteriorate during storage unless held at low temperatures. The addition of chloroform, as a preservative for the texin in intestinal contents, does not increase the persistence of the toxin to a significant extent.

The pattern of absorption of epsilon toxin from the intestine into the bloodstream during the course of experimental enterotoxaemia was followed by using a radioactive tracer (I_{125} Polyvinylpyrroldine). It was found that, as with the protein tracers used by other workers, the values of I_{125} PVP in studies of this nature is limited. However, it did reveal that there is a loss of high molecular weight substances from the bloodstream into the extravascular tissues during this intoxication.

Studies of the ultrastructural changes which occur in the tissues of intoxicated animals have revealed that there is a severe generalised vascular endothelial damage, both when epsilon toxin is administered intravenously and when it is absorbed from the intestine. There is no histochemical or histological evidence to suggest that epsilon toxin produces primary morphological changes in tissues, other than endothelium,

although secondary effects are detectable in other organs. These secondary changes include the development of protein-containing effusions in serous cavities, and oedena in a number of tissues.

In the brain, fluid accumulation is primarily intracellular and is confined to the protoplasmic astrocytes. This results in swelling of astrocyte processes in the grey matter so that the increase in fluid content can be detected by electron microscopy as well as quantitatively. The changes in the astrocytes form the basis for early brain lesions which are detectable by light microscopy e.g. vacuolation in the corebellar granular layer. Associated with the intracellular fluid accumulation there is an extracation of protein into the extracellular spaces of the brain and this is detectable in mice by using exogenous peroxidase as a tracer.

Fluid loss from the bloodstream is also prominent in the heart and the lungs. In the former tissue there is severe myocardial oedema and fluid accumulation within the cardiac muscle cells, which may explain the electrocardiographic abnormalities that develop during the course of intoxication. Although lung oedema is not a consistent feature of the disease, and may be a reflection of damage caused by high concentrations of epsilon toxin entering the pulmonary circulation, it can be extremely severe with accumulation of fluid in the alveoli and interstitium.

Although it is possible to relate these changes to the vascular damage, no possible mechanism for the action of the toxin on the endothelium has been established. The use of red cell stroma and fluorescent antibody tests has failed to provide any evidence that the toxin is bound to cell membranes either <u>in vitro</u> or <u>in vivo</u>. Warburg respirometry did not reveal any reduction in the metabolic efficiency of tissue slices that were under the influence of epsilon toxin.

The overall loss of fluid into the tissues of intoxicated animals results in a severehaemoconcentration which, because of the severity of the endothelial damage, is not associated with any increase in levels of plasma proteins or inorganic ions such as sodium, potassium or chloride.

One of the most prominent of the biochemical changes that occur in enterotoxaemia is a severe progressive hyperglycaemia which appears to result from the rapid mobilisation of hepatic glycogen. The reduction in the glycogen content of the hepatic tissue can be detected histochemically. A further finding which suggests that hepatic glycogenolysis forms the basis for the blood sugar changes is that the hyperglycaemic response was suppressed in animals in which hepatic glycogen stores had been depleted prior to the administration of epsilon toxin.

Associated with the rise in blood glucose there are also increased amounts of lactate, pyruvate and alphaketoglutarate in the blood. These changes are considered to be a reflection of increased metabolic activity due to the increased amounts of available glucose. The build-up of intermediate substances reflects normal rate-limiting steps in aerobic glycolysis rather than any direct interference with a particular biochemical pathway by the toxin.

The high levels of lactate in the blood cause a severe metabolic acidosis in intoxicated animals which may sometimes be masked by alterations in blood pH associated with deficient respiratory exchange and elevated values of blood pCO₂ caused by the pulmonary cedema.

The importance of the morphological, biochemical, haematological and physiological changes, which have been found in the present investigation, in increasing our understanding of the pathogenesis of enterotoxacmia are discussed. In addition the relevance of these findings to the broader fields of experimental pathology and to the inter-relationships which exist between the cellular and clinical pathology of disease states are described.

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INDEX

CHAPTER:		PAGE
1	INTRODUCTION	1
2	A REVIEW OF THE LITERATURE ON CLOSTRIDIUM PERFRINGENS TYPE D ENTEROTOXAEMIA.	5
3	RATIONALE OF THE EXPERIMENTAL METHODS USED IN THE PRESENT INVESTIGATION. Materials and Methods Employed to Produce	15
	Intexication. Information Required on the Pathogenesis of	15
	Enterotoxaemia.	17
. 4	CHARACTERISTICS OF THE EPSILON TOXIN USED IN THE EXPERIMENTAL PROCEDURES AND THE STABILITY OF EPSILON	
	TOXIN IN VITRO. Determination of Lethal Levels of Epsilon Toxin	20
	Preparations for Mice. Confirmation of the Necrotising Action of Epsilon	21
	Toxin on Guinea Pig Skin. Neutralisation of the Toxicity of Epsilon Toxin	22
	with Type Specific Antisera. i. Lethal Action. Neutralisation of the Toxicity of Epsilon Toxin with Type Specific Antisera. ii. Necrotising	23
	Action. The Stability of the Samples of Epsilon Toxin	25
	Used in the Experimental Procedures. The Stability of Epsilon Toxin in Intestinal	25
	Contents Under Different in vitro Storage Conditions.	26
	Discussion.	27
	Conclusion.	30

CHAPTER		PAGE
5	FACTORS AFFECTING THE ACTION OF EPSILON TOXIN ON	
	LABORATORY ANIMALS.	32
	General Description of the Mice Used in the	
	Experimental Work.	33
	The Effect of Toxin Dose on the Survival Time	
	of Intoxicated Mice.	34
	The Effect of Sex on the Survival Time of	
	Intoxicated Mice.	34
	An Investigation of the Possible Hormonal Basis	
	for the Sex Difference in Survival Time Pattern.	35
	The Effect of Nutritional Factors on the Action	
	of Epsilon Toxin in Mice.	36
	Discussion.	38
	Conclusion.	40
6	FACTORS INFLUENCING THE ACTION OF EPSILON TOXIN ON	
	LAMBS.	41
	The Influence of Different Rates and Routes of	
	Administration of Epsilon Toxin on the Patholog-	
	ical Changes Produced in Lambs.	43
	The Effect of the Infusion of Cl.perfringens type	8
	$\underline{\mathtt{D}}$ plus Carbohydrate into the Duodenum of Lambs.	44
	The Pattern of Absorption of Radiodinated	
	Polyvinylpyrrolidone from the Intestine of Lambs.	. 46
	Discussion.	47
	Conclusion.	49
7	THE CLINICAL SIGNS AND GROSS PATHOLOGICAL CHANGES	
	ASSOCIATED WITH CL.PERFRINGENS TYPE D EPSILON TOXIN	
	INTOXICATION.	51
	The Clinical Signs and Gross Pathology of Epsilon	l
	Tóxin Intoxication in Mice.	52
	The Clinical Signs and Gross Pathology of Epsilon	l
	Toxin Intoxication in Lambs.	52

CHAPTER		PAGE
	The Electrocardiographic Changes which Occur	
	in Lambs Receiving Epsilon Toxin.	54
	Discussion.	56
	Conclusion.	59
8	THE MORPHOLOGICAL CHANGES PRODUCED IN THE CENTRAL NERVOUS SYSTEM DURING <u>CL.PERFRINGENS</u> TYPE D EPSILON TOXIN INTOXICATION.	60
	Quantitative Aspects of Alterations in Fluid Balance in the Brains of Intoxicated Mice. Quantitative Aspects of Alterations in Fluid	62
	Balance in the Brains of Intoxicated Lambs. Morphological Changes in the Brains of Intoxicat	64 -
	ed Animals seen by Electron Microscopy. The Use of Horse Radish Peroxidase as an Indicator of Altered Capillary Permeability in	65
	Intoxicated Animals. Morphological and Histochemical Changes Detectable by Light Microscopy in the Brains of	67
	Intoxicated Animals. The Effect of Critical Hypoxia on the Morphology	68
	and Fluid Balance in Mouse Brain.	70
	Discussion.	71
	Conclusion.	76
9	THE MORPHOLOGICAL ASPECTS OF THE PULMONARY AND MYOCARDIAL OEDEMA WHICH OCCURS IN EXPERIMENTAL	
	ENTEROTOXAEMIA. The Histopathological Changes in the Lungs of	78
	Epsilon Toxin Intexicated Lambs. Quantitative and Morphological Evidence of the Presence of Myocardial Oedema in Intoxicated	79
	Lambs.	80
	Discussion.	81
	Conclusion.	84

CHAPTER		PAGE
10	THE DEVELOPMENT OF RENAL LESIONS IN ENTEROTOXAEMIA.	86
	The Lesions which Occur in the Kidneys of	
	Intexicated Animals at the Time of Death.	88
	The Influence of Epsilon Toxin upon the	
	Histochemical Activity of the Kidney.	89
	The Influence of Starvation on the Renal Lesions	
	which Occur during Epsilon Toxin Intoxication in	
	Mice,	90
	The Influence of Autolysis on the Development of	
	Renal Lesions in Epsilon Toxin Intoxication and	- 0
	Enterotoxaemia.	92
	Discussion.	98
	Conclusion.	101
11	THE MORPHOLOGICAL CHANGES PRODUCED IN THE INTESTINE	
	BY EPSILON TOXIN.	103
	The Effect of Epsilon Toxin on Ligated Intestinal	10)
	Loops in the Rabbit.	104
	The Effect of Epsilon Toxin on Ligated Intestinal	
	Loops in the Lamb.	105
	The Morphological Changes Produced in the Intest-	
	ine of Lambs by Experimental Enterctoxaemia.	107
	Discussion.	108
	Conclusion.	110
12	THE MORPHOLOGICAL CHANGES IN SKIN, PANCREAS, THYROID,	
	ADRENAL, MUSCLE, PITUITARY AND LIVER PRODUCED BY	
	EPSILON TOXIN.	112
	Changes in Liver, Pancreas, Thyroid, Adrenal,	!
	Pituitary and Skeletal Muscle in Mice and Lambs.	114
	The Morphological Features of the Cutaneous	
	Damage Produced by the Intradermal Administration	
	of Epsilon Toxin.	113
	Discussion.	116
	Conclusion.	118

CHAPTER		PAG?
13	THE MOLPHOLOGICAL FEATURES OF FIELD CASES OF	
	CL.PERFRINGENS TYPE D EN PEROTOXAEMIA IN LAMBS.	11:
	A Case of Acute Enterotoxaemia.	120
	A Case of Focal Symmetrical Encephalomalacia.	122
	Discussion.	124
	Conclusion.	126
14	THE EFFECT OF EPSILON TOXIN ON TISSUE FUNCTION.	127
	Possible Binding of Epsilon Toxin to Cell	
	Membranes - Erythrocyte Stroma in vitro.	128
	Possible Binding of Epsilon Toxin to Cell	
	Membranes - Indirect Fluorescent Antibody Methods	
	in viva.	130
	Oxygen Uptake of Control and Intoxicated Tissue	471.
	Slices - Warburg Respirometry.	134
	Discussion.	137
	Conclusion.	138
15	THE NORMAL VALUES OF A NUMBER OF BLOOD CONSTITUENTS	
	OF LAMBS AND THE TECHNIQUES EMPLOYED FOR THEIR	
	ESTIMATION.	140
	The Estimation of Biochemical and Haematological	
	Parameters in Lambs.	141
	Discussion.	1½6
	Conclusion.	156
16	THE EFFECT OF CL.PERFRINGENS TYPE D EPSILON TOXIN ON	
	RESPIRATORY EN HANGE AND ACID-BASE BALANCE IN LAMBS.	159
	Discussion	163
	Conclusion.	165

CHAPTER		PAG
17	HAEMATOLOGICAL ALTERATIONS AND CHANGES IN LEVELS OF PLASMA PROTEINS IN LAMBS AFFECTED BY EPSILON TOXIN.	165
		10.5
	Alterations in Haematological Parameters and Plasma Protein Levels Following the Parenteral	
	Administration of Epsilon Toxin.	167
	lterations in Haematological Parameters and	
	Plasma Protein Levels in Lambs with Experimental	
	Enterotoxaemia.	169
	Discussion.	170
		174
	Conclusion.	1/4
18	THE EFFECT OF EPSILON TOXIN ON THE CONCENTRATIONS OF	
	SOME OF THE MAJOR IGNIC CONSTITUENTS OF THE PLASMA	
	OF LAMBS.	176
	The Effect of the Parenteral Administration of	
	Epsilon Toxin on the Concentration of Sodium,	
	Potassium, Chloride and Phosphate Ions in Lamb	
	Blood.	177
	The Effect of Experimental Enterotoxaemia on	
	Blood Levels of $Na^+, K^+, Cl^- \& PC_4^{}$ in Lambs.	178
	Discussion.	179
	Conclusion.	181
19	THE PATTERN OF CHANGES IN BLOOD GLUCOSE, PYRUVATE,	
	LACTATE, ALPHAKETO=GLUT.RATE AND KETONE BODIES IN	
	INTOXICATED LAMBS.	183
	Alterations in Glucose and Lactate in Lamb Blood	
	Following the Intravenous Administration of	
	Epsilon Toxin.	184
	Alterations in Blood Levels of Glucose, Lactate,	
	Pyruvate, Alphaketoglutarate and Ketone Bodies	
	During Experimental Enterotoxaemia.	185

CHAPTER		PAGE
	Glucose a: llactate Values in Fost Mortem Blood	
	Samples from a Field Case of Enterotoxaemia.	189
	Discussion.	189
	Conclusion.	193
20	THE INFLUENCE OF INSULIN AND STARVATION UPON THE HYPERGLYCAEMIA INDUCED BY <u>CL.PERFRINGENS</u> TYPE D EPSILON TOXIN.	195
	The Influence of Insulin and Starvation upon the Blood Glucose Response of Lambs Following the Parenteral Administration of Epsilon Toxin.	196
	The Effect of Insulin upon the Hyperglycaemia which Occurs in Experimental Enterotoxaemia.	199
	Discussion.	200
	Conclusion.	201
21	THE EFFECT OF EPSILON TOXIN ON THE LEVELS OF SEVERAL	
	SERUM ENZYMES IN SHEEP.	203
	Discussion.	206
	Conclusion.	207
22	ALTERATIONS IN URINE CONSTITUENTS OF LAMBS INDUCED BY EPSILON TOXIN.	208
	The Urinary Constituents of 2-3 week-old Lambs and the Effect of Epsilon Toxin upon their Excretion.	209
	The Urine Constituents of Lambs During the Course of Experimental Enterotoxaemia.	212
	Discussion.	214
	Conclusion.	216
23	GENERAL DISCUSSION.	2 i 8

APPENDIX .		Pi.GE
1	Surgical and Manipulative Procedures - Mice.	01
2	Sex: Survival Time Data - Mice.	03
3	Hormonal Status: Survival Time Data for Mice.	04
4	Surgical and Manipulative Procedures - Sheep.	05
5	Total Body, Brain and Lung Weights, Epsilon Antitoxin Levels and Dosage of Epsilon Toxin for the 2-3 week-old Experimental Lambs.	08
6	Levels of I ₁₂₅ Polyvinylpyrrolidone in Plasma Following Infusion of this Compound into the Duodenum of Normal Sheep and Sheep with Experimental Enterotoxaemia.	09
7	Survival Time: Brain water Content Data - Mice	011
8	Brain Water Content Data - Lambs.	013
9	Fluorescent Antibody Methods.	015
10	The Oxygen Uptake of Control and Intoxicated Tissue Slices.	018
11	Automated Biochemical Estimations.	020
а	Glucose, Fyruvate and Lactate	022
Ъ	Glutamic-oxalcacetic and Glutamic-pyruvic Transaminases plus Total Protein	029
С	Lactic and Isocitric Dehydrogenases	02!
d	Acid and Alkaline Phosphatases	039
е	Serum and Urine Sodium, Potassium and Chloride	- 1 1
	(and Bicarbonate)	044
f	Inorganic Phosphate	049
12	Control Haematological and Biochemical Data 2-3 Week-old Lambs.	052
13	Post-Inoculation Haematological and Biochemical Data 2-3 week-old Lambs.	056

APPENDIX		PAGE
14	Haematological and Biochemical Data from Lambs	
	which received Intraduodenal Infusions of	
	Cl.perfringens type D plus Carbohydrate	060
15	Blood Glucose: Survival Time Data for Starved and	
	Unstarved Lambs which received Insulin and/or	
	Epsilon Toxin	062
16	Control and Post-Inoculation Biochemical Data -	
	Urine - 2-3 week-old Lambs.	064

LIST OF ILLUSTRATIONS

Figure	:	Following Page
4.1	The Necrotising Action of Cl.perfringens type D	
	Epsilon Toxin and its Neutralisation by Type-	
	Specific Antitoxin.	23
4.2	The Persistence of Epsilon Toxin in Intestinal	
	Contents under Different Storage Conditions.	27
5.1	The Influence of the Administration of One or Two	
	Lethal Doses of Epsilon Toxin on the Pattern of	
	Survival Time in Mice.	34
5.2	The Influence of Sex on Survival Time in Mice.	35
5.3	The Effect of Hormonal Status on Survival Time in	
	Mice.	36
6.1	The Influence of Circulating Epsilon Antitoxin on	
	the Dose of Epsilon Toxin required to Produce	
	Intoxication in Lambs.	44
6.2a	The Effect of Mode of Administration of Epsilon	
	Toxin on the Development of Pulmonary Oedema in	
	Lambs.	44
6.2b	The Effect of Mode of Administration of Epsilon	
	Toxin on the Development of Plaural Effusions in	
	Lambs.	44
6.3	The Pattern of Development of Epsilon Toxin in the	
	Intestine of a Lamb during Infusion of Cl.	
	perfringens type D Culture plus Starch into the Duodenum.	1.6
		46
6.4	The Pattern of Absorption of I ₁₂₅ PVP from the Intestine into the Plasma of Control and	
	Intoxicated Lambs.	47
7 4		47
7.1	Alterations in the Electrocardiograms of Lambs which had received Parenterally Administered	
	Epsilon Toxin.	55
	•	//

Figure:	Following Page
7.2 Alterations in the Electrocardiograms of Lamb which had Absorbed Enteric-Origin Epsilon Tox	
8.1 The Effect of Epsilon Toxin Intoxication on t Water Content of Mouse Brain.	he 63
8.2 Swelling of Astrocyte Foot Processes around Capillaries in the Thalamic Region of the Brain Intoxicated Mice.	in 67
8.3 Swelling of Astrocyte Processes in the Thalam Neuropil of Intoxicated Mice.	
8.4 Differential Swelling of Protoplasmic Astrocy in the Cerebellar Granular Layer of Intoxicat Mice.	
8.5 Swelling of Astrocyte Processes around Neuron in the Thalamic Neuropil of Intoxicated Mice.	
8.6 Vascular Endothelial Damage in the Thalamic R of the Brains of Intoxicated Mice.	egion 67
8.7a The Development of Brain Oedema in Intoxicate Lambs.	67
8.7b Vascular Endothelial Damage in the Brain of Intoxicated Lambs - Parenterally Administered Toxin.	67
8.8 The Distribution of Horse Radish Peroxidase i Brains of Control and Intoxicated Mice.	
8.9 The Distribution of Horse Radish Peroxidase i Cerebellar Granular Layer of Intoxicated Mice	
8.10 The Sequence of Changes seen by Light Microsc in the Cerebellum of Intoxicated Mice.	юр у 69
8.11 Neuronal Damage in Mice Resulting from Prolon Critical Hypoxia.	ged 71
8.12 The Differential Swelling of Cells Resembling Bergman Glial Cells in the Cerebellum of Into Mice.	

Figure	:	Following Page
9.1	Pulmonary Oedema in Intoxicated Lambs - Light Microscopic Appearance.	79
9.2	Vascular Damage in the Lungs of Lambs after the Parenteral Administration of Epsilon Toxin.	79
9.3	The Development of Pulmonary Oedema in Lambs after the Parenteral Administration of Epsilon Toxin.	79
9.4	Vascular Endothelial Damage in Lamb Heart following the Parenteral Administration of Epsilon Toxin.	81
9.5	Myocardial Oedema in Intoxicated Lambs.	81
9.6	Myocardial Gedema in Epsilon Toxin Intoxication and Experimental Enterotoxaemia.	81
9•7	Swelling of Mitochondrial Cristae in the Myocardium of Intoxicated Lambs.	81
9.8	Myocardial Oedema in Intoxicated Lambs - Light Microscopic Appearance.	81
10.1	The Light Microscopic Appearance of the Renal Change seen in Field Cases of <u>Cl.perfringens</u> type D Enterotoxaemia.	e s 86
10.2	Vascular Endothelial Damage in the Renal Intertubula Capillaries of Intoxicated Lambs.	
10.3	Accumulation of Demonstrable Lipid in the Kidneys of Starved and Intoxicated Mice.	f 92
10.4a	The Sequence of Autolytic Changes in the Kidneys of Normal Mice after Death - Electron Microscopy.	95
10.4b	The Sequence of Autolytic Changes in the Kidneys of Intoxicated Mice after Death - Electron Microscopy.	95
10.5	The Sequence of Autolytic Changes in the Kidneys of Intoxicated Lambs after Death - Light Microscopy.	96
10.6	The Sequence of Autolytic Changes in the Kidneys of Normal Lambs after Death - Light Microscopy.	97

Figure:	1	Following Page
10.7	The Sequence of Autolytic Changes in the Kidneys of Intoxicated Lambs after Death - Electron Microscopy.	98
10.8	Vascular Changes which Developed 4 Hours after Death in a Case of Experimental Enterotoxaemia in a Lamb.	98
10.9	The Sequence of Autolytic Changes in the Kidneys of Normal Sheep after Death - Electron Microscopy.	98
11.1	The Effect of Epsilon Toxin on the Intestine of the Rabbit.	104
11.2	The Effect of Epsilon Toxin on the Intestine of the Rabbit - Light Microscopy.	105
11.3	Epithelial and Endothelial Damage Produced by Epsilos Toxin in Ligated Loops of Rabbit Intestine.	n 105
11.4	The Effect of Epsilon Toxin on the Mucosa of Lamb Intestine.	107
12.1	The Loss of Glycogen from the Livers of Intoxicated Lambs - Light Microscopy.	115
12.2	The Loss of Glycogen from the Livers of Intoxicated Lambs - Electron Microscopy.	115
13.1	Vascular Endothelial Damage in the Cerebral Cortex of a Field Case of Acute Enterotoxaemia.	122
13.2	Autolytic Changes and Vascular Damage in Kidney from a Field Case of Acute Enterotoxaemia.	122
13.3	Thalamic Lesions in a Field Case of Focal Symmetrica Encephalomalacia	1 123
14.1	Effect of <u>Cl.perfringens</u> type D Epsilon Toxin on the Respiration of Tissue Slices.	136
15.1	Differences in the Electrophoretic Patterns of the Plasma Proteins of 2 - 3 week old and 8 month old	
	Lambs.	156
16.1	The Effect of Intravenously Administered Epsilon Toxin on Blood pH and pCO2 in Lambs.	161

Figure:	<u>F</u>	ollowing Page
16.2	The Influence of Pulmonary Oedema on the Changes which occur in Blood pH and pCO ₂ in Intoxicated Lambs.	162
16.3	The Pattern of Change in Blood pH and pCO ₂ in a Lamb with Experimental <u>Cl.perfringens</u> type D Enterotoxaemia	. 162
17.1a	The Effect of the Parenteral Administration of Epsilon Toxin on the Haematocrit and Total Plasma Protein of Lambs.	169
17.1b	The Effect of the Development of Pulmonary Oedema on the Changes in Haematocrit which occur in Epsilon Toxi Intoxicated Lambs.	.n 169
17.2	The Effect of Enterotoxaemia, Produced by the Intraduodenal Infusion of <u>Cl.perfringens</u> type D Culture plu Carbohydrate on the Haematocrit and Total Plasma Protected Levels of Lambs.	
18.1	The Effect of Experimental Enterotoxaemia on Blood Lev of Inorganic Phosphate.	rels 179
19.1a	Alterations in the Levels of Blood Glucose and Lactate following Intravenous Administration of Epsilon Toxin.	
19.1b	Alterations in the Level of Blood Glucose and Lactate during Experimental Enterotoxaemia in Lambs.	185
19.2	The Effect of Enterotoxaemia on Blood Levels of Glucos Pyruvate, Lactate and Alphaketoglutarate.	se, 188
20.1	The Influence of Insulin and Starvation upon the Blood Glucose Response of Lambs to Intravenously Administers	ed
22.1	Epsilon Toxin. Alterations in Blood and Urine Glucose during Cl.perfringens type D Enterotoxaemia.	199 2 13
Append.	'Autoanalyser' Flow Diagrams and Calibration Peaks Employed for the Automated Biochemical Analyses.	025