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EXPRESSION, PURIFICATION AND MUTAGENESIS OF RECOMBINANT CLASS 1 ALDEHYDE DEHYDROGENASE

A thesis presented in partial fulfilment of the requirements for the degree of Master of Science in Molecular Biology at Massey University

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1997

ABSTRACT

Aldehyde dehydrogenase (AlDH) catalyses the conversion of aldehydes, for example acetaldehyde and retinal, to carboxylic acids in an NAD⁺-dependent reaction involved in the detoxification of aldehydes and alcohols. There are several isoenzymes, class 1 being the cytosolic form.

Three over-expression and purification systems have been tested, in order to gain a high yield of active, pure class 1 aldehyde dehydrogenase. The traditional method, using T7 polymerase-driven expression in E. coli p-hydroxyacetophenone-affinity ion-exchange and followed by chromatography, gave 3 mg/L human aldehyde dehydrogenase with a high specific activity of 1.2 units/mg. Human Class 1 AlDH has been overexpressed and purified using the GST Gene Fusion System (Pharmacia Biotech), avoiding the need for AlDH-affinity chromatography and therefore allowing straight-forward purification of mutated enzymes. The GST fusion system produced 2.6 mg/L pure AIDH with a specific activity of 0.39 units/mg. The methylotrophic yeast Pichia pastoris was chosen for its high yields, in the region of grams/litre. Preparatory work was carried out with the construction of the expression plasmids and screening of the *Pichia* transformants.

A highly conserved lysine residue (Lys-272) may be involved in acid-base catalysis of aldehyde oxidation, as well as of the esterase reaction also catalysed by AIDH. Preliminary work has been carried out on the generation of the K272A, K272R, K272H and K272L altered enzymes. The resultant activity level, kinetic behaviour and active site structure of these modified enzymes should help to elucidate further the mechanism of action of AIDH.

ACKNOWLEDGEMENTS:

I would like to offer a very large thank you to my supervisors, Dr Mike Hardman and Dr Mark Patchett. Thank you for your time and patience, and for allowing me my independence in order to make my own mistakes. In addition, the writing of this thesis has been greatly helped by Mike, who would read chapter drafts in a day (the headings did renumber themselves!), and my time in the lab would have been far more stressful without Mark's extensive knowledge of the best way to do things and his ability to think around a problem to find solutions.

My thanks are due to two of our Twilight Zone postdocs, Dr Shaun Lott and Dr Catherine Day. Shaun coped well with a never-ending barrage of questions and often managed to make me see the overall picture when I had been incapable of seeing the wood for the trees. Mrs Carole Flyger deserves a special mention for her perseverance in keeping the lab running when we all needed everything yesterday. My thanks also to everyone who has worked in the Twilight Zone over the last two years, for making the time an enjoyable one.

I would like to thank my friends and flatmates over the last two years, especially Kathryn Frith who has put up with my stresses and moans and then offered me a place to stay for my last three months of this thesis - thank you. Nachos will never be the same again.

My parents have always supported me in everything that I have strived to do. Your love, encouragement and friendship have meant so much to me. Thank you for believing in me; this thesis is dedicated to you.

Finally my thanks to Andrew Milne. We have survived both my thesis and you in Gisborne; Christchurch here we come!

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