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The Role of Ethylene and Auxin in Responses of Roots to Phosphate Supply in White Clover (Trifolium Repens L.)

Phuong Dinh Thi Yen 2009

The Role of Ethylene and Auxin in Responses of Roots to Phosphate Supply in White Clover (Trifolium Repens L.)

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ABSTRACT

Phosphate (P) supply is one of the major determining factors to plant productivity, since the element affects the growth and the development of plants. In response to P-deficiency treatment, plants display alterations in root system architecture caused by changes in primary root (PR) and lateral root (LR) length and LR density. In this thesis, the root growth of the agronomically important legume, white clover (*Trifolium repens* L.) was found to be slightly stimulated in terms of PR length, LR number and total LR length when plants were grown in a P-deficient media (0.01 mM orthophosphate; Pi) when compared with plants grown in a P-sufficient media (1.00 mM Pi) when using a hydroponic growth system.

When plants are grown in a P-sufficient media, treatment with 100 nM exogenous 1-aminocyclopropane-1-carboxylic acid (ACC) and exogenous auxin (5 nM 1-naphthylacetic acid, NAA) resulted in significant increases in white clover PR length, LR number and LR length. However, when ethylene action or auxin transport were inhibited using 300 ppm 1-methylcyclopropene (1-MCP) and 100 nM 1-N-naphthylphthalamic acid (NPA), respectively, root growth was significantly reduced which suggests roles for ethylene and auxin in mediating white clover root growth.

To examine the effects of these hormones on plants grown in P-deficient media, 100 nM ACC treatment significantly enhanced the stimulatory effects of growth on P-deficient media only, while exposure of plants to P-deficiency alone was sufficient to significantly neutralise the inhibitory effects of 1-MCP on root growth. Hence, exposure to P-deficiency is proposed to increase either ethylene biosynthesis or ethylene sensitivity in white clover roots. In contrast, for plants grown in P-deficient media, treatment with 5 nM NAA significantly abolished the stimulation of white clover root growth observed with P-deficiency so it is proposed that exposure to P-deficiency increases either auxin biosynthesis or auxin sensitivity, but the 5nM NAA concentration used was too high to stimulate root growth. Using DR5p::GUS transgenic white clover, auxin activity was found in the root tips and root primordia. Using these plants, it is suggested that P-deficient treatment and ACC treatment influenced white clover root growth through an increase in auxin sensitivity.

Overall, ethylene and auxin are found to be essential in mediating white clover root growth in P-sufficiency, and also in mediating root responses to P-deficiency through changes in terms of the biosynthesis and the sensitivity of these two hormones.

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LIST OF ABBREVIATIONS

1-MCP 1-methylcyclopropene

6-BAP 6-Benzylaminopurine

ACC 1-aminocyclopropane-1-carboxylate

ACO 1-aminocyclopropane-1-carboxylate oxidase
ACS 1-aminocyclopropane-1-carboxylate synthase

AVG Aminoethoxyvinylglycine

BCIP 5 bromo-4-chloro-3-indolyl phosphate

BFA Brefeldin A

bp base pair

ca. approximately

Cef³⁰⁰ Cefotaxime (300 µg.mL⁻¹)

cm centimetre

CTR CONSTITUTE TRIPLE RESPONSE

DAT day after treatment

DMF Dimethylformamide

DMSO Dimethyl sulphoxide

DNA Deoxyribonucleic acid

EDTA Ethylenediaminetetraacetic acid

EIN ETHYLENE INSENSITIVE

ERF ETHYLENE RESPONSE FACTOR

ERS ETHYLENE RESPONSE SENSOR

ETR ETHYLENE TRIPLE RESPONSE

FAA Formalin acetic acid

g acceleration due to gravity (9.8 m.s^{-2})

g gram

GMO Genetically Modified Organism

GUS β -glucuronidase

HCl Hydrochloric acid

IAA Indole-3-acetic acid

Kan¹⁵⁰ Kanamycin sulfate (150 μg.mL⁻¹)

°C Degrees in celsius

Kb kilo base pair

kPa kilo Pascal

L Litre

LR(s) lateral root(s)

M moles L⁻¹

mg milligram

MgSO₄ Magnesium sulfate

min minute mL millilitre

mM millimoles L⁻¹

MQ water water purified by a Milli-purification system

MS Murashige and Skoog base media

NAA 1-naphthyleneacetic acid

NaCl Sodium chloride NaOH Sodium hydroxide

NBT Nitro-blue tetrazolium chloride

ng nanogram

nM nanomoles L⁻¹

NPA 1-N-naphthylphthalamic acid

P P value from *t*-test

P Phosphate

 P_{adi} adjusted P value from Tukey's test

PAT polar auxin transport

PC2 Physical Containment 2

PCR Polymerase Chain Reaction

PGU Plant Growth Unit

pH $-\log [H^{\dagger}]$

Pi⁻ P-deficient treatment

Pi inorganic phosphate, orthophosphate

Pi⁺ P-sufficient treatment

ppm parts per million PR(s) primary root(s)

QC quiescent centre

RAM root apical meristem

RH Relative humidity

RNA ribonucleic acid

Rnase Ribonuclease

RO reverse osmosis

rpm round per minute

RSA root system architecture

RT room temperature

SAM S-adenosyl methionine

SDS Sodium dodecyl sulfate

SE standard error of the mean

SSC Sodium citrate – sodium chloride buffer

TAE Tris-Acetate-EDTA

TFs transcription factors

TIBA 2,3,5-triiodobenzoic acid

Tris Tris(hydroxymethyl)aminomethylamine

Trp Tryptophan

TY Tryptone-yeast extract

UV ultra violet light

V volts

v/v volume per volume

w/v weight per volume

w/w weight per weight

WT wild type

X-GLUC 5-Bromo-4-chloro-3-indolyl β-D-glucuronide

μg microgram

μL microlitre

μm micrometre

μM micromoles L⁻¹

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