

Copyright is owned by the Author of the thesis. Permission is given for a copy to be downloaded by an individual for the purpose of research and private study only. The thesis may not be reproduced elsewhere without the permission of the Author.

Twisted Intercalating Nucleic Acids (TINA) in Guanosine-rich Oligonucleotides

A thesis submitted in the partial fulfilment
of the requirements for the degree of

Doctor of Philosophy in Chemistry

Massey University,

Palmerston North, New Zealand



MASSEY UNIVERSITY

Osman Doluca

2013

Abstract

The main role in the structural diversity of DNA molecules belongs to guanosines due to their array of hydrogen bond donors and acceptors, large aromatic surface and ability to adopt *syn* or *anti* conformations. These properties lead to the formation of various DNA topologies such as triplexes or G-quadruplexes by guanosine-rich oligonucleotides. For a long time these secondary structures were mainly considered to be a fascinating phenomenon with little practical use; it was subsequently realised that these structures are likely to be formed under physiological conditions and therefore might be involved in many important biological processes, including genome recombination, telomere stability and regulation of gene expression. Thus, there is a growing interest in development and control of these non-traditional nucleic acid structures.

Although the secondary structures of nucleic acids can be controlled to a certain extent by the careful design of oligonucleotide sequence this strategy alone is not always sufficient. In this thesis we investigated how to control the assemblies of guanosine-rich oligonucleotides using a novel tool, twisted intercalating nucleic acids (TINAs). The incorporation of pyrene-containing TINA monomers into guanosine-rich oligonucleotides led to the formation of stable triplexes or G-quadruplexes depending on the position of TINA monomers. In the light of our results, we have established a set of rules that helps to create a desired structure of guanosine-rich oligonucleotides using TINA molecules.

In the second half of the thesis we focused on expanding the functionality of TINA conjugated oligonucleotides. In terms of fluorescence, we synthesised several fluorescently-silent triplex-forming oligonucleotides (TFOs) equipped with a dye at different positions in the DNA. Fluorescence properties were strongly dependent on the position of the dye. These fluorescently silent TFOs showed up to an 18-fold increase in fluorescent intensity upon triplex formation.

These findings lay the foundation for the future design of artificial DNA sequences for expanding the repertoire of DNA secondary structures and function.

Acknowledgements

I would like to thank my supervisors Professor Geoff Jameson and Dr Vyacheslav V. Filichev for their support and assistance throughout this project. Especially Vyacheslav for the enthusiasm he showed towards the project and the time taken to show me various techniques of DNA chemistry. I would like to thank Professor Alexandre Boutorine for his support to this project and hospitality.

Thanks to all former and present colleagues in IFS for their assistance over the years, especially Jamie Withers and Emad Al-imarrah for their friendship and Pat Edwards for his help with NMR. I am also grateful for assistance of departmental staff over the years I have been at Massey University.

I would like to acknowledge important financial support from the Marsden grant administered by the Royal Society of New Zealand, the Dumont d'Urville grant from NZ/France Science and Technology Support Programme and a doctoral completion bursary from Massey University.

Many thanks to my dear parents, Saliha and Nevzat Doluca, for their infinite support reaching all the way to the other side of the world.

Above all I would like to thank my dear wife, Rose, for her patience and support during completion of this PhD and my kids, Leila and Musa, for making my life cOLourfuL at every step.

Table of Contents

Abstract.....	i
Acknowledgements.....	ii
Table of Contents.....	iii
List of Figures.....	vii
List of Tables.....	xiv
Abbreviations.....	xvi
Chapter 1. Introduction.....	1
1.1 Aim.....	1
1.2 Properties of DNA.....	3
1.2.1 Triple Helical DNA.....	7
1.2.1.1 Properties of Parallel Triplexes.....	9
1.2.1.2 Properties of Antiparallel Triplexes.....	10
1.2.2 G-quadruplexes.....	10
1.2.3 Using Synthetic Chemistry for Studying DNA Structures.....	13
1.2.3.1 Automated DNA Synthesis.....	13
1.3 Post-Synthetic Treatments and Purification.....	16
1.3.1 Backbone Modifications.....	17
1.3.2 Base Modifications.....	18
1.3.3 Intercalators.....	20
1.3.3.1 Twisted Intercalating Nucleic Acid (TINA).....	21
1.4 Challenges.....	23
1.5 Thesis Outline.....	24
Chapter 2. Methods for Studying DNA Structures.....	26
2.1 Introduction.....	26
2.2 Gel Electrophoresis.....	26
2.2.1 Denaturing Gel Electrophoresis.....	27
2.2.2 Non-denaturing Gel Electrophoresis.....	27

2.2.3	Determination of Dissociation Constant (K_d).....	27
2.3	UV-Vis Spectroscopy	28
2.3.1	Determination of Concentration.....	28
2.3.2	Determination of Melting Temperature	29
2.3.3	Thermal Difference Spectra (TDS).....	30
2.4	Fluorescence Spectroscopy	31
2.5	Circular Dichroism (CD) Spectroscopy	32
2.6	Workflow for Studying Nucleic Acid Structures.....	33
Chapter 3. Synthesis and Biological Studies of TINA-TFOs		34
3.1	Introduction.....	34
3.2	Chapter Outline.....	36
3.3	Synthesis of TINA Monomer.....	36
3.4	Design and Synthesis of Triplex-forming Oligonucleotides (TFOs).....	37
3.5	Properties of TINA-Incorporated Antiparallel Triplexes.....	40
3.5.1	Determination of Dissociation Constants of Triplexes (K_d).....	40
3.5.2	TINA Insertion as a Bulge in the TFO is Essential for Triplex Formation and Stability, Especially at Physiological pH	41
3.5.3	TINA-Conjugated Triplexes Preserve Sequence Selectivity	41
3.5.4	Antiparallel Triplexes are Significantly More Stable than Parallel Triplexes	41
3.5.5	Triplexes Formed by GT Sequences are More Stable than GA sequences.....	43
3.5.6	TINA Incorporation at the Junction Points of Different Nucleotide Tracts Should be Avoided.....	44
3.5.7	Insertions of TINA should be Located at Least 3 Bases Apart.....	44
3.5.8	TINA Insertion into the G-tract Helps to Disrupt the G-quadruplex Formation.....	44
3.5.9	TINA Oligonucleotides Show Tendency to Self-aggregate.....	47
3.5.10	Antiparallel Triplex Formation by TINA-TFOs is a Slower than Formation of Parallel Triplexes	48
3.5.11	Thymidine Insertion Instead of TINA can be Used to Simulate TINA's Effect of G-quadruplex Disruption	49
3.6	Demonstration of Applicability of the Rules for TINA-TFO Design.....	50
3.7	Discussion	52
Chapter 4. TINA Incorporated G-quadruplexes		56
4.1	Introduction.....	56

4.2	Design of G-quadruplexes	57
4.3	Properties of TINA Incorporated G-quadruplexes.....	58
4.3.1	TINA Located Outside the G-tract.....	58
4.3.2	TINA in the Middle of the G-tract.....	67
4.3.3	TINA Accelerates Association of Tetramolecular G-quadruplexes	76
4.3.4	Effect of 3' Fluorescein Modification	77
4.4	Discussion.....	80
4.5	Summary	82
Chapter 5. Incorporation of Minor Groove Binders to TINA-Conjugated TFOs.....		84
5.1	Introduction.....	84
5.1.1	MGB-TFO Conjugates.....	86
5.1.2	Aim	86
5.2	Probe Design.....	87
5.2.1	Solid-phase Synthesis of Minor-Groove Binders	88
5.3	Synthesis of Probes	88
5.3.1	Conjugation with TFOs.....	89
5.4	Dissociation Constants (K_d) of MGB-TFO Conjugates from Target I.....	92
5.5	Discussion	92
5.6	Summary	93
Chapter 6. Fluorescently Silent TFO probes.....		94
6.1	Introduction.....	94
6.1.1	Aim	95
6.2	Synthesis of Fluorescently Silent DNA Probes	95
6.2.1	Synthesis of Fluorescent Moiety.....	97
6.2.2	Synthesis of Oligonucleotides.....	98
6.2.3	Purification of Modified TFOs	102
6.3	Properties of Fluorescently Silent Probes	102
6.3.1	Parallel TFOs	103
6.3.1.1	Properties of Parallel Triplexes.....	103
6.3.1.2	Properties of DNA/RNA and DNA/DNA Duplexes.....	108
6.3.2	Antiparallel TFOs	109
6.3.2.1	Properties of Antiparallel Triplexes.....	109

6.3.2.2	Properties of DNA/RNA and DNA/DNA Duplexes.....	115
6.4	Discussion.....	116
6.5	Summary.....	118
Chapter 7. Thesis Conclusion and Future Directions.....		119
7.1	Thesis Conclusion.....	119
7.2	Future Directions.....	120
Chapter 8. Experimental Methods		122
8.1	General Remarks.....	122
8.2	Synthesis of TINA phosphoramidite.....	122
8.3	Synthesis of Cyanine Dye.....	126
8.4	Synthesis of Minor Groove Binders (MGBs).....	128
8.5	Other Modifications.....	130
8.6	Oligonucleotide Synthesis.....	130
8.7	Post-synthetic CuAAC Reaction for Fluorescent TFOs.....	130
8.8	Purification of Oligonucleotides.....	131
8.9	Post-synthetic Conjugation of Minor Groove Binders.....	132
8.10	Polyacrylamide Gel Electrophoresis.....	133
8.10.1	Dissociation Constant (K_d) Measurements.....	133
8.10.2	Kinetics experiments using PAGE.....	134
8.11	UV Spectroscopy.....	135
8.11.1	Determination of Concentration.....	135
8.11.2	Thermal Difference Spectra.....	135
8.11.3	Determination of Melting and Annealing Temperatures.....	136
8.12	Association Rate Constant (k_{on}) Measurements.....	136
8.13	Circular Dichroism (CD) Spectroscopy.....	137
8.14	Fluorescence Spectroscopy.....	137
8.14.1	Quantum Yield (Φ_F) Measurements.....	137
8.15	NMR Spectroscopy of Oligonucleotides.....	138
8.16	Mass Spectrometry of Oligonucleotides.....	138
Appendix.....		140
References.....		143

List of Figures

<i>Number</i>	<i>Page</i>
Figure 1.1 Twisted Intercalating Nucleic Acid (TINA).....	2
Figure 1.2 A double-stranded DNA (dsDNA) formed by two single-stranded DNA (ssDNA) chains.	3
Figure 1.3 B-type DNA model.....	4
Figure 1.4 Top, an AT base-pair with two hydrogen bonds. Bottom, a GC base-pair with three hydrogen bonds. Hydrogen bonds between the pairs are shown as dashed lines.	5
Figure 1.5 <i>Anti</i> and <i>syn</i> confirmations of adenosine and cytidine.	6
Figure 1.6 A) Models for T•A-T, C ⁺ •G-C, G•G-C, A•A-T base triplets within a triple helix motif. Directionality of the backbones is indicated with ⊗ (3') and ⊙ (5'). B) Orientations of parallel and antiparallel TFOs relative to the duplex strands. Shaded boxes represent pyrimidines, empty boxes represent purine bases, boxes with dashed borders represent bases of the third strand (Sun, Garestier <i>et al.</i> 1996).....	8
Figure 1.7 A) G-tetrad formation. 'R' refers to backbones of strands that form G-quadruplex. B) Various G-quadruplex structures. Arrows indicate 5' to 3' polarity. Squares indicate guanine arrangement inside a G-tetrad.	11
Figure 1.8 Design of nucleoside phosphoramidites (A) and protected nucleic bases (B) for DNA synthesis.....	14
Figure 1.9 Synthetic cycle for preparation of oligonucleotides by phosphoramidite method.....	15
Figure 1.10 Unmodified deoxyribonucleic acid (DNA) and backbone modifications: locked nucleic acid (LNA), peptide nucleic acid (PNA), phosphothiate (PS), N,N-Dimethylaminopropylamine (DMAP), N,N-diethylethylenediamine (DEED) and methoxyethylamine (MeOEt).	18
Figure 1.11 Base modifications (A) and intercalators (B) for triplex-forming oligonucleotides: 6-thioguanine (6SG), 7-deazaguanine (^{7da} G) and 6-thio-7-deazaguanine (^{7da} 6SG), 7-Chloro-7-deazaguanine (^{7Cl} daG), 7-deazaxanthines (^{7d} X), psoralen (Ps), thiazole orange (TO1), pyrene (P) and intercalating nucleic acid (INA).	19

Figure 1.12 Different ways of intercalation: a) Intercalator is placed between adjacent base pairs, and b) intercalator is stacking at the ends of the duplex.	20
Figure 1.13 Parallel triple helical DNA complex formed by TINA incorporated TFO.	21
Figure 1.14 Representation of the TINA molecule in the design of G-quadruplexes (path a) and antiparallel DNA triplexes (path b).....	22
Figure 2.1 DNA melting curve examples. Solid line and dashed line represent melting curves with a single and double transition, respectively. Circle represents the point of midtransition. ..	29
Figure 2.2 Workflow for studies of nucleic acid structures.....	33
Figure 3.1 CD spectra of G-quadruplexes formed by 5'-dAGGGGGGGTTTTGTTTT sequence (10 μ M) at 20 $^{\circ}$ C (solid line) and 90 $^{\circ}$ C (dashed line) in the presence of 10 mM Na cacodylate buffer, 100 mM NaCl and 10 mM MgCl ₂ at pH 7.2.....	35
Figure 3.2 Synthesis of TINA monomer. Reagents and conditions: a) KOH, toluene, reflux. b) Amberlite (H ⁺) resin, H ₂ O, 65 $^{\circ}$ C. c) Pd(PPh ₃) ₄ , CuI, DMF/Et ₃ N, argon at 20 $^{\circ}$ C. d) DCM/Et ₃ N, argon at 20 $^{\circ}$ C. e) DCM, diisopropylammonium tetrazolide, argon at 20 $^{\circ}$ C.	37
Figure 3.3 Non-denaturing gel analysis of TFO 7 with Target I. Duplex concentration: 60 nM, probe concentrations: 5, 1, 0.5, 0.1, 0.05, 0 μ M (lanes 1 to 6, respectively) in 0.05 M HEPES buffer, 50 mM NaCl, 5 mM MgCl ₂ at pH 7.2, 37 $^{\circ}$ C.	40
Figure 3.4 Non-denaturing 20 % PAGE analysis of TFOs in HEPES buffer (50 mM), KCl (150 mM), MgCl ₂ (5 mM), at pH 7.2, 37 $^{\circ}$ C. Lanes 1 - 11, 19 - 22 refer to TFOs 1 - 11, 19 - 22, respectively. M: Marker. Oligonucleotide concentration was set to 100 μ M. Gel was stained by Stains-All® and destained in water.	43
Figure 3.5 Native PAGE (20 %) analysis of TFOs 1-10 and 16 in HEPES buffer (50 mM), NaCl (50 mM), MgCl ₂ (5 mM), at pH 7.2, 37 $^{\circ}$ C. Lanes 1 - 10 refers to TFOs 1 - 10, respectively. Lane 10.1 refers to 1M urea and LiOH treated TFO 10 in order to destabilise G quadruplexes, while lane 16 refers to TFO 16. Oligonucleotide concentration was set to 100 μ M. Gel was initially observed under UV light (A) and then stained by Stains-All® and destained in water (B).	45
Figure 3.6 CD spectra of A) TFO 3 and B) TFO 7 in comparison with TFO 10. CD profiles of target duplex, target I, in the absence and presence of C) TFO 7 and D) TFO 10 in Na cacodylate buffer (20 mM) with NaCl (50 mM) and MgCl ₂ (5mM) at pH 7.2, 20 $^{\circ}$ C. Concentrations of oligonucleotides are 1.0 μ M.	46

Figure 3.7 Triplex conversion versus time (min) by TFO 3 and 7 at 100 μM strand concentration in the presence of 0.06 μM Target I in HEPES buffer (50 mM), NaCl (50 mM), MgCl_2 (5 mM), pH 7.2 at 20 $^\circ\text{C}$.	48
Figure 3.8 Fluorescence spectra of TFO 3 (A), 4 (D), 7 (B) and 9 (C) in the presence and in the absence of the target duplex, Target I, λ_{ex} :373 nm, λ_{em} :380 nm – 600 nm, in Na cacodylate buffer (20 mM), NaCl (50 mM), MgCl_2 (5 mM), at pH 7.2, 20 $^\circ\text{C}$. Concentration of each strand is set to 1.0 μM .	49
Figure 3.9 Non-denaturing PAGE (20 %) analysis of ABL sequences listed in Table 3.4 in HEPES buffer (50 mM), NaCl (50 mM), MgCl_2 (5 mM) at pH 7.2, 37 $^\circ\text{C}$ with DNA ladders consisting of 50-, 40-, 30-, 20- and 15-mer oligothymidylates. Oligonucleotide concentration is 100 μM . Gel was stained with Stains-All [®] and destained in water.	52
Figure 4.1 Thermal difference spectrum of TG4T at 10 μM strand concentration after annealing in the presence of 110 mM KCl in 10 mM Li cacodylate buffer at pH 7.2, 20 $^\circ\text{C}$.	58
Figure 4.2 Thermal difference spectra of XTG at 10 μM strand concentration after annealing in the presence of 110 mM KCl (dashed line) or NaCl (grey line) in 10 mM Li cacodylate buffer at pH 7.2, 20 $^\circ\text{C}$.	59
Figure 4.3 Thermal difference spectra of TXG at 10 μM strand concentration after annealing in the presence of 110 mM KCl (dashed line) or NaCl (grey line) in 10 mM Li cacodylate buffer at pH 7.2, 20 $^\circ\text{C}$.	59
Figure 4.4 Thermal difference spectrum of the TINA duplex at 10 μM strand concentration after annealing in the presence of NaCl in 10 mM Li cacodylate buffer at pH 7.2, 20 $^\circ\text{C}$.	60
Figure 4.5 Melting (dashed lines) and annealing (solid lines) profiles of XTG (A) and TXG (B) after incubation in 110 mM NaCl, 10 mM Li cacodylate buffer at pH 7.2. Oligonucleotide concentration was 10 μM . The profiles are based on absorbance data recorded at 373 nm with 0.18 $^\circ\text{C}/\text{min}$ temperature ramp.	61
Figure 4.6 Native PAGE (20 %) analysis of oligonucleotides (100 μM) in the presence of 110 mM KCl, 10 mM Li cacodylate at pH 7.2, 4 $^\circ\text{C}$. Ladder contains 10-, 15-, 20-, 25-, 40- and 50-mer oligothymidylates. See Appendix Figure 2 for oligonucleotides in the presence of 110 mM NaCl instead of 110 mM KCl.	62
Figure 4.7 Fluorescence emission spectra of XTG, TXG and TINA duplex at 10 μM strand concentration in the presence of 110 mM NaCl in 10 μM Li cacodylate buffer at pH 7.2, 20 $^\circ\text{C}$.	

$\lambda_{\text{ex}} = 373 \text{ nm}$. The fluorescence profiles in the presence of 110 mM KCl instead of 110 mM NaCl are very similar (see Appendix Figure 3).	64
Figure 4.8 CD spectrum of TG4T at 10 μM strand concentration in the presence of 110 mM KCl in 10 mM Li cacodylate buffer, pH 7.2, 20 $^{\circ}\text{C}$	64
Figure 4.9 CD spectra of XTG at 10 μM strand concentration in the presence of 110 mM KCl (dashed line) or NaCl (grey line) in 10 mM Li cacodylate buffer, pH 7.2, 20 $^{\circ}\text{C}$	65
Figure 4.10 CD spectra of TXG at 10 μM strand concentration in the presence of 110 mM KCl (dashed line) or NaCl (grey line) in 10 mM Li cacodylate buffer, pH 7.2, 20 $^{\circ}\text{C}$	65
Figure 4.11 A) NMR spectra of TXG at 100 μM strand concentration in 10 mM Na^+ phosphate buffer, pH 7.0. B) NMR spectra of TXG at 500 μM strand concentration in 10 mM Na^+ phosphate buffer and 10 mM KCl, pH 7.0.	66
Figure 4.12 Thermal difference spectra of GXG at 10 μM strand concentration after annealing in the presence of 110 mM KCl (dashed line) or NaCl (grey line) in 10 mM Li cacodylate buffer at pH 7.2, 20 $^{\circ}\text{C}$	67
Figure 4.13 Thermal difference spectra of GXX at 10 μM strand concentration after annealing in the presence of 110 mM KCl (dashed line) or NaCl (grey line) in 10 mM Li cacodylate buffer at pH 7.2, 20 $^{\circ}\text{C}$	68
Figure 4.14 Thermal difference spectra of G3X at 10 μM strand concentration after annealing in the presence of 110 mM KCl (dashed line) or NaCl (grey line) in 10 mM Li cacodylate buffer at pH 7.2, 20 $^{\circ}\text{C}$	68
Figure 4.15 Native PAGE (20 %) analysis of oligonucleotides (100 μM) in the presence of 110 mM NaCl (A) or 110 mM KCl (B) in 10 mM Li cacodylate at pH 7.2, 37 $^{\circ}\text{C}$. Ladder contains 10-, 15-, 20-, 25-, 40- and 50-mer oligothymidines.....	69
Figure 4.16 Melting and annealing profiles of GXG (A) and G3X (B) at 10 μM strand concentration after incubation in 110 mM NaCl, 10 mM Li cacodylate buffer at pH 7.2. The profiles are based on absorbance data recorded at 373 nm with a 0.18 $^{\circ}\text{C}/\text{min}$ rate.	70
Figure 4.17 CD spectra of GXG at 10 μM strand concentration in the presence of 110 mM KCl (dashed line) or NaCl (grey line) in 10 mM Li cacodylate buffer at pH 7.2, 20 $^{\circ}\text{C}$	72
Figure 4.18 CD spectra of GXX at 10 μM strand concentration in the presence of 110 mM KCl (dashed line) or NaCl (grey line) in 10 mM Li cacodylate buffer at pH 7.2, 20 $^{\circ}\text{C}$	72

Figure 4.19 CD spectra of G3X at 10 μ M strand concentration in the presence of 110 mM KCl (dashed line) or NaCl (grey line) in 10 mM Li cacodylate buffer at pH 7.2, 20 $^{\circ}$ C.	73
Figure 4.20 Fluorescence emission spectra of GXG, G3X, GXX and TINA duplex at 10 μ M strand concentration in the presence of 110 mM NaCl in 10 mM Li cacodylate buffer at pH 7.2, $\lambda_{\text{ex}} = 373$ nm.	74
Figure 4.21 NMR spectra of GXG at 1.0 mM strand concentration in 10 mM Na^+ phosphate buffer, pH 7.0.	75
Figure 4.22 Isothermal renaturation processes of GXG (20 μ M, A) and G3X (10 μ M, B) recorded at 275 and 280 nm, respectively, in the presence of 110 mM NaCl in 10 mM Li cacodylate buffer at pH 7.2, 20 $^{\circ}$ C. For experiment details see Chapter 8.12.	77
Figure 4.23 CD spectra of GXGF at 10 μ M strand concentration in the presence of 110 mM KCl (dashed line) or NaCl (grey line) in 10 mM Li cacodylate buffer, pH 7.2.	78
Figure 4.24 CD spectra of TXGF at 10 μ M strand concentration in the presence of 110 mM KCl (dashed line) or NaCl (grey line) in 10 mM Li cacodylate buffer at pH 7.2, 20 $^{\circ}$ C.	79
Figure 4.25 Fluorescence emission spectra of GXGF, TXGF at 10 μ M concentration in the presence of 110 mM NaCl or KCl in 10 mM Li cacodylate buffer at pH 7.2, 20 $^{\circ}$ C. $\lambda_{\text{ex}} = 373$ nm.	79
Figure 4.26 The intensity of fluorescence emission at 520 nm during thermal melting of GXGF (10 μ M) with 0.2 $^{\circ}$ C/min rate in the presence of 110 mM NaCl in 10 mM Li cacodylate buffer at pH 7.2, 20 $^{\circ}$ C. $\lambda_{\text{ex}} = 465$ nm.	80
Figure 5.1 Structures of <i>N</i> -methylpyrrole (Py), <i>N</i> -methylimidazole (Im), β -alanine (β) and γ -aminobutyric acid (γ).	84
Figure 5.2 Structure of the polyamide hairpin minor groove binder, Py ₂ Im- γ -Py ₃ - β -Dp (A) and its interaction with double-stranded DNA sequence (B). Dp is (<i>N,N</i> -dimethylamino)propylamide; β is β -alanine; γ is γ -aminobutyric acid.	85
Figure 5.3 The recognition sites of TFO and MGB on the duplex, Target I.	87
Figure 5.4 Denaturing PAGE (20 %) of a coupling reaction between MGB and p(EG) ₆ -TFO after 2 and 16 hours.	89
Figure 5.5 Activation and coupling of MGBs with TFO-p(EG) ₆ ; compound 1, mono-conjugated TFO-p(EG) ₆ -MGB; compound 2, bis-conjugated TFO-p(EG) ₆ -MGB ₂	90

Figure 5.6 A) TFO-p(EG) ₆ -Py ₃ -γ-Py ₃ -β-Dp, mono conjugate; B) TFO-p(EG) ₆ -(Py ₃ -γ-Py ₃ -β-Dp) ₂ , bis conjugate.	91
Figure 6.1 Copper-catalysed azide-alkyne cycloaddition reaction (CuAAC).	96
Figure 6.2 Structure of the cyanine-based dye (compound 20) to be incorporated into DNA.	96
Figure 6.3 The structures of the linkers after DNA synthesis prior to CuAAC reaction.	97
Figure 6.4 Synthesis of the azide-containing fluorescent moiety for incorporation into TFOs. ..	98
Figure 6.5 Deprotection of ethynyl-containing nucleotides and the CuACC reaction with fluorescent dye (compound 20).	101
Figure 6.6 CD spectra of duplex D1 and a parallel triplex T1+D1 in Li cacodylate buffer (10 mM), NaCl (100 mM) and MgCl ₂ (10 mM) at pH 5.0, 20 °C. Strand concentration was set to 1.0 μM.	103
Figure 6.7 CD spectra of TFO T2, duplex D1 and a parallel triplex T2+D1 in Li cacodylate buffer (10 mM), NaCl (100 mM) and MgCl ₂ (10 mM) at pH 5.0, 20 °C. Strand concentration was set to 1.0 μM.	104
Figure 6.8 CD spectra of TFO T3, duplex D1 and a parallel triplex T3+D1 in Li cacodylate buffer (10 mM), NaCl (100 mM) and MgCl ₂ (10 mM) at pH 5.0, 20 °C. Strand concentration was set to 1.0 μM.	104
Figure 6.9 CD spectra of TFO T4, duplex D1 and a parallel triplex T4+D1 in Li cacodylate buffer (10 mM), NaCl (100 mM) and MgCl ₂ (10 mM) at pH 5.0, 20 °C. Strand concentration was set to 1.0 μM.	105
Figure 6.10 Fluorescence emission spectra (λ _{ex} = 464 nm) of TFO T2, duplexes T2+C1 and T2+R1, parallel triplex T2+D1 and mismatched triplex T2+D2 in Li cacodylate buffer (10 mM), NaCl (100 mM) and MgCl ₂ (10 mM) at pH 5.0, 20 °C. Strand concentration was set to 1.0 μM.	107
Figure 6.11 Fluorescence emission spectra (λ _{ex} = 464 nm) of TFO T3, duplexes T3+C1 and T3+R1, parallel triplex T3+D1 and mismatched triplex T3+D2 in Li cacodylate buffer (10 mM), NaCl (100 mM) and MgCl ₂ (10 mM) at pH 5.0, 20 °C. Strand concentration was set to 1.0 μM.	107
Figure 6.12 Fluorescence emission spectra (λ _{ex} = 464 nm) of TFO T4, duplexes T4+C1 and T4+R1, parallel triplex T4+D1 and mismatched triplex T4+D2 in Li cacodylate buffer (10 mM),	

NaCl (100 mM) and MgCl ₂ (10 mM) at pH 5.0, 20 °C. Strand concentration was set to 1.0 μM.	108
Figure 6.13 Fluorescence emission spectra ($\lambda_{\text{ex}} = 464 \text{ nm}$) of TFO T5, duplexes T5+C2 and T5+R2, parallel triplex T5+D2 and mismatched triplex T5+D2m in Li cacodylate buffer (10 mM), NaCl (100 mM) and MgCl ₂ (10 mM) at pH 7.2, 20 °C. Strand concentration was set to 1.0 μM.....	111
Figure 6.14 Fluorescence emission spectra ($\lambda_{\text{ex}} = 464 \text{ nm}$) of TFO T6, duplexes T6+C2 and T6+R2, parallel triplex T6+D2 and mismatched triplex T6+D2m in Li cacodylate buffer (10 mM), NaCl (100 mM) and MgCl ₂ (10 mM) at pH 7.2, 20 °C. Strand concentration was set to 1.0 μM.....	111
Figure 6.15 Fluorescence emission spectra ($\lambda_{\text{ex}} = 464 \text{ nm}$) of TFO T8, duplexes T8+C3 and T8+R3, parallel triplex T8+D3 and mismatched triplex T8+D3m in Li cacodylate buffer (10 mM), NaCl (100 mM) and MgCl ₂ (10 mM) at pH 7.2, 20 °C. Strand concentration was set to 1.0 μM.....	112
Figure 6.16 Fluorescence emission spectra ($\lambda_{\text{ex}} = 464 \text{ nm}$) of TFO T9, duplexes T9+C3 and T9+R3, parallel triplex T9+D3 and mismatched triplex T9+D3m in Li cacodylate buffer (10 mM), NaCl (100 mM) and MgCl ₂ (10 mM) at pH 7.2, 20 °C. Strand concentration was set to 1.0 μM.....	113
Figure 6.17 CD spectra of TFO T7 and triplex T7+D3 in Li cacodylate buffer (10 mM), NaCl (100 mM) and MgCl ₂ (10 mM) at pH 7.2, 20 °C. Strand concentration was set to 1.0 μM.....	113
Figure 6.18 CD spectra of TFO T8 and triplex T8+D3 in Li cacodylate buffer (10 mM), NaCl (100 mM) and MgCl ₂ (10 mM) at pH 7.2, 20 °C. Strand concentration was set to 1.0 μM.....	114
Figure 6.19 CD spectra of TFO T9 and triplex T9+D3 in Li cacodylate buffer (10 mM), NaCl (100 mM) and MgCl ₂ (10 mM) at pH 7.2, 20 °C. Strand concentration was set to 1.0 μM.....	114
Figure 6.20 Dyes used in the synthesis of fluorescently silent parallel TFOs.....	117

List of Tables

<i>Number</i>	<i>Page</i>
Table 2.1 Characteristic wavelengths of TDS and CD spectra for different nucleic acid structures.....	30
Table 3.1 Sequences of TFOs synthesised or purchased.....	38
Table 3.2 Sequences of target duplexes I, II and III. Target II consists of an additional A-T base-pair, was designed as target for unmodified TFO 11. Target III was designed as a mismatching target.....	39
Table 3.3 Modified and unmodified TFOs targeting HIV-1 proviral DNA and their dissociation constants [μM] from triplexes with corresponding duplex, target I (60 nM), in HEPES buffer (50 mM) containing MgCl_2 (5 mM), at pH 7.2, 37 °C, supplemented with 50 mM NaCl, 150 mM NaCl or 150 mM KCl.	42
Table 3.4 Modified and unmodified TFOs targeting the <i>ABL</i> region and their dissociation constants [μM] from triplexes with corresponding duplexes (Table 3.5, 60 nM) in HEPES (50 mM), NaCl (50 mM) and MgCl_2 (5 mM) at pH 7.2, 37 °C.....	51
Table 3.5 Sequences of target duplexes IV, V. Target IV and V are designed for probes based on ABL 1 and ABL 6, respectively.	51
Table 4.1 Sequences used in this chapter and their abbreviations.....	57
Table 4.2 $T_{1/2}$ values of G-quadruplexes at 10 μM strand concentration obtained during melting and annealing processes using 0.18 °C/min temperature ramp in the presence of 110 mM NaCl or KCl in 10 mM Li cacodylate at pH 7.2.	62
Table 4.3 Retardation of the G-quadruplexes formed by TINA-TG ₄ T sequences in native PAGE (20 %) and molecularity in the presence of 110 mM NaCl or KCl in 10 mM Li cacodylate buffer at pH 7.2, 37 °C.	63
Table 4.4 $T_{1/2}$ values of GXG, GXX and G3X obtained during melting and annealing, using 0.5 or 1.0 °C/min temperature ramp in the presence of 110 KCl in 10 mM Li cacodylate buffer at pH 7.2.....	71
Table 4.5 Sequences of various ONs and their k_{on} values obtained in NaCl (110 mM).....	76
Table 5.1 Probes and their sequences.	87

Table 5.2 Standard protocol for manual solid-phase synthesis of pyrrole-imidazole polyamides.	88
Table 5.3 Sequences and dissociation constants (K_d) of triplexes formed with target I in 10 mM HEPES buffer in 50 mM NaCl, 5 mM MgCl ₂ at pH 7.2, 37 °C.	92
Table 6.1 Abbreviations and sequences of target duplexes, complementary DNAs and RNAs containing polypurine binding sites.	99
Table 6.2 Modified and unmodified triplex-forming oligonucleotides (TFO) and their sequences. See Figure 6.5 for definition of K, L ^U , M ^U and M ^A	100
Table 6.3 Fluorescence data of parallel TFOs in 10 mM Li cacodylate, 100 mM NaCl and 10 mM MgCl ₂ at pH 5.0, 20 °C at 1.0 μM strand concentration.	106
Table 6.4 Fluorescence data of antiparallel TFOs in 10 mM Li cacodylate, 100 mM NaCl and 10 mM MgCl ₂ at pH 7.2, 20 °C at 1.0 μM strand concentration.	110
Table 8.1 Manual synthesis cycle of MGB.	128
Table 8.2 Results of mass spectroscopy analysis of ONs synthesised.	139

Abbreviations

(EG) ₆	hexaethylene-glycol
μL	microlitre
μM	micromole/litre
μmol	micromole
A	adenosine
Å	Ångström
<i>ABL</i>	Abelson murine leukemia viral oncogene
Abs	absorbance
ACN	acetonitrile
AcOH	acetic acid
aq	aqueous
ATR	attenuated total reflectance
<i>BCR</i>	breakpoint cluster region
bp	basepair
C	cytosine
CD	circular dichroism
COMBO-FISH	combinatorial oligonucleotide probes in fluorescence <i>in situ</i> hybridization
CPG	controlled pore glass
CTAB	cetyl trimethylammonium bromide
CuAAC	copper-assisted azide-alkyne cycloaddition
DCA	dichloroacetic acid
DCI	4,5-dicyanoimidazole
DCM	dichloromethane
ddH ₂ O	double-distilled water
dH ₂ O	distilled water
DIEA	<i>diisopropylethylamine</i>
DMA	<i>N,N</i> -dimethylacetamide
DMF	<i>N,N</i> -dimethylformamide
DMPA	<i>N,N</i> -dimethylaminopropylamine
DMSO	dimethylsulfoxide
DMT	4,4'-dimethoxytrityl
DNA	deoxyribonucleic acid

Dp	(<i>N,N</i> -dimethylamino)propylamide
dsDNA	double-stranded DNA
ϵ	extinction coefficient
EDTA	<i>N,N</i> -ethylenediaminetetraacetic acid
eq	equivalent
ESI	electrospray ionisation
EtBr	ethidium bromide
EtOAc	ethyl acetate
EtOH	ethyl alcohol
F_c	fluorescence intensity of DNA complex
FISH	fluorescence <i>in situ</i> hybridization
FRET	Förster resonance energy transfer
F_{ss}	fluorescence intensity of single-stranded DNA
FT-IR	Fourier transform infrared spectroscopy
G	guanosine
HATU	<i>O</i> -(7-azabenzotriazol-1-yl)-1,1,3,3-tetramethyluronium hexafluorophosphate
HEPES	4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid
HIV	human immunodeficiency virus
HPLC	high pressure liquid chromatography
<i>HRAS</i>	Harvey rat sarcoma viral oncogene
Im	<i>N</i> -methylimidazole
INA	intercalating nucleic acid
IR	infrared
K_d	dissociation constant
<i>KRAS</i>	Kirsten rat sarcoma viral oncogene
L	litre
LNA	locked nucleic acid
M	mole/litre
MALDI	matrix-assisted laser desorption/ionization
MeOH	methyl alcohol
MGB	minor groove binder
min	minute
mL	millilitre
mM	millimole/litre
mmol	millimole
mRNA	mitochondrial ribonucleic acid

<i>MYC</i>	myelocytomatosis viral oncogene
NaOAc	sodium acetate
N-FISH	non-denaturing fluorescence <i>in situ</i> hybridization
nm	nanometre
NMP	<i>N</i> -methyl-2-pyrrolidone
NMR	nuclear magnetic resonance
ON	oligonucleotide
P	pyrene
p	terminal phosphate
PAGE	polyacrylamide gel electrophoresis
Ph	phenyl
PhSH	thiophenol
PNA	peptide nucleic acid
ppm	parts per million
Ps	psoralen
Py	<i>N</i> -methylpyrrole
qRT-PCR	quantitative real time polymerase chain reaction
RNA	ribonucleic acid
s	second
SNP	single nucleotide polymorphism
<i>SRC</i>	sarcoma viral oncogene
ssDNA	single-stranded DNA
ssRNA	single-stranded RNA
T	thymidine
$T_{1/2}$	mid-transition point
TBTA	tris[(1-benzyl-1 <i>H</i> -1,2,3-triazol-4-yl)methyl]amine
TCA	trichloroacetic acid
TDS	thermal difference spectra
TEAA	triethylammonium acetate
TFA	trifluoroacetic acid
TFO	triplex-forming oligonucleotide
THF	tetrahydrofuran
TINA	twisted intercalating nucleic acid
TLC	thin-layer chromatography
T_m	melting temperature
TO	thiazole orange

TOF	time of flight
TSP	trimethylsilyl propionate
U	uridine
UV	ultraviolet
Vis	visible
β	β -alanine
γ	γ -aminobutyric acid
λ_{em}	emission wavelength
λ_{ex}	excitation wavelength
Φ_F	fluorescence quantum yield