Copyright is owned by the Author of the thesis. Permission is given for a copy to be downloaded by an individual for the purpose of research and private study only. The thesis may not be reproduced elsewhere without the permission of the Author.

## Investigation of the immunostimulatory effects of some New Zealand honeys and characterization of an active component

A Thesis submitted in partial fulfilment of the requirements for the degree of

Master of Technology in FOOD TECHNOLOGY at Massey University, Albany New Zealand

Swapna Gannabathula

June 2010

#### Abstract

Medicinal use of honey has re-emerged recently indicating that honey accelerates wound healing activity. Honey has been shown to stimulate TNF- $\alpha$  production from monocytes and macrophages which is apparently correlated with a high molecular weight fraction, and not lipopolysaccharide (LPS, an immunostimulatory endotoxin) levels. Cytokine production by honey has been attributed to the endotoxin content. The aim of this study was to investigate the ability of Comvita sourced honeys to elicit a TNF- $\alpha$  cytokine response from acute monocytic leukemia (THP-1) cells as well as identify the responsible component.

Five honey samples were used together with sugar and methylglyoxal controls. The samples were incubated with THP-1 cells, with and without LPS. After incubation, the cell culture supernatants were collected and TNF- $\alpha$  was measured by the enzyme-linked immune sorbent assay (ELISA). The most active honey samples were further heat-treated to remove enzyme/protein/peptide-like stimulation; the samples were treated with polymixin B (PmB) to remove LPS-like stimulation and not protein fraction. The samples were then filtered by molecular weight centrifugal filters to separate constituents according to their size and the fractions were re-analysed.

All five honey samples in the absence of LPS stimulated TNF- $\alpha$  release from THP-1 cells, whereas untreated, sugar- and methylglyoxal-treated cells did not. The cytokine production was partially inhibited by heating, but mostly by PmB. In the filtered honey samples, the activity was observed in the >30 kDa fraction. These results suggest that the activity may be associated with one or more components which are partially heat-labile, LPS-like stimulated with a high molecular weight.

Further, honey samples were analyzed for the concentration of LPS present. The tests revealed that the cytokine stimulation was higher than would be expected from the concentration of LPS present in the honey. The possibility that this component was a plant-derived  $\beta$ -glucan, which is known to have LPS-like activity and can interfere with detection of LPS in the LAL assay, was investigated. Subsequent analyses confirmed the presence of arabinogalactan, a large complex carbohydrate.

The data presented in this study suggests that arabinogalactans in honey may stimulate inflammatory responses and the release of cytokines that are crucial in regulating wound-healing. This heralds a significant advancement in the usage and understanding of medicinal honey.

#### **Acknowledgements**

I would particularly like to take this opportunity to convey my sincere gratitude and appreciation to the following people and institutions for their advice, support, help and encouragement throughout the tenure of my study.

First of all, I thank God for guarding safety, good health and showing me the right way throughout my life.

Sincere and appreciate acknowledgements to my supervisor, Dr. Doug Rosendale, for offering me this opportunity, his advice, and support throughout the course of this study. His constant inspiration, enthusiasm, strong motivation and guidance in the subject area were appreciated. Not only did he give me prompt advice and answers to my questions, but he also enabled me to be able to think about aspect of the work from different angles.

I would like to express equally important appreciative acknowledgments to my supervisors, Professor Margot Skinner and Dr. Tony Mutukumira, for their continuous support, guidance, encouragement, and kind co-operation throughout my study. I am very much thankful to Dr.Tony Mutukumira for giving me the opportunity to study at the Institute of Food, Nutrition and Human Health, Massey University. A special thanks to Professor. Margot Skinner for her kind helps as I wrote my thesis.

Thanks to Dr Ralf Schlothauer and Dr Jonathan Stephens, from Comvita New Zealand Limited, for creating and directing the programme, for providing me honey samples and their support, guidance, enthusiasm, strong motivation, and proof reading of my thesis.

I am thankful to Dr. Jeff Greenwood, Edward Walker and Judie Farr, for their kind help, assistance in learning cell culture techniques and ELISAs.

I would like to express my thanks to Dr. Paul Loong from ESR and Dr. Ian Sims from IRL for their work outlined in Chapter 5.

I would like to acknowledge Graham Fletcher, Graeme Summers and Cristina Cruz for their valuable guidance, friendly attitude, answering my silly questions, suggesting to me the right path and support not only throughout this study but also outside of it.

My personal thanks to, Jill Chantarachoti, Sravani Gupta, Jessicah Win, Sabrina Tian, Sean Chen and Andrew Boey, for their kind help, co-operation and patience and, it was great to share daily moments with them and Monday morning teas.

Words cannot describe my heartiest gratitude towards my husband Subrahmanyeswarlu Gannabathula. This thesis would not have been possible without the advice, love, companionship, support, encouragement and guidance from him. His lively, optimism, faithfulness and his passion for study, understanding and patiently kept telling me it could be done, and whose hard work can be seen on every page. Subbu you are one in a million and came into my life as light, happiness and without you I could not have been what I am and where I am now.

Comvita New Zealand Limited and the New Zealand Foundation for Research Science and Technology for providing me with a scholarship, research funding, arranging research facilities, and conference travel grant; the New Zealand Institute for Plant & Food Research Limited for providing research facilities; the Institute of Food, Nutrition and Human Health, Massey University for providing the Master candidature registration and travel grant.

Last but not least I am really indebted to my parents Jhansi Rani Maraoni (Mom) Ramdas Maraoni (Dad) and my daughters Yashaswini Gannabathula and Harika Gannabathula for their love, companionship, help, and continuous support through the whole process of this study. The words of encouragement, moral support and valuable guidance from my parents made it possible to complete my study.

To my Husband Subrahmanyeswarlu, and daughters, Yashaswini and Harika May you live a long and happy life, With all my love

### List of Contents

| Abstract   | ii    |
|--|-------|
| Acknowledgements                                 | iv    |
| List of Contents                                 | vii   |
| List of Figures                                  | xiv   |
| List of Tables                                   | xviii |
| List of Abbreviations                            | xix   |
| Chapter 1 Introduction and literature review     | 1     |
| <b>1.0</b> Introduction                          | 1     |
| <b>1.1</b> Honey                                 | 1     |
| <b>1.2</b> Past review on honey                  | 2     |
| <b>1.2.1</b> Composition of honey                | 2     |
| <b>1.2.2</b> Production of honey                 | 4     |
| <b>1.2.3</b> Microorganisms in honey             | 4     |
| 1.2.3.1 Sources of microbial contamination       | 5     |
| <b>1.2.4</b> Varieties of honey                  | 6     |
| <b>1.2.4.1</b> Floral origin                     | 6     |
| 1.2.4.2 Geographical origin                      | 7     |
| <b>1.3</b> Properties of honey                   | 7     |
| <b>1.3.1</b> The antibacterial activity of honey | 8     |
| 1.3.1.1 Osmotic effects                          | 8     |
| 1.3.1.2 Hydrogen peroxide                        | 9     |
| <b>1.3.1.3</b> Methylglyoxal                     |       |

| <b>1.3.1.4</b> Acidity   | 10 |
|--|----|
| 1.3.1.5 Phytochemicals   | 11 |
| <b>1.3.1.6</b> Antimicrobial peptides                          | 11 |
| <b>1.3.2</b> Antifungal activity of honey                      | 12 |
| <b>1.3.3</b> Anti-oxidant activity of honey                    | 12 |
| <b>1.3.4</b> Wound healing activity of honey                   | 13 |
| 1.3.4.1 Historical usage of honey                              | 14 |
| <b>1.3.4.2</b> Wound healing compounds in honey                | 15 |
| 1.3.4.3 Clearing infections                                    | 16 |
| <b>1.3.4.4</b> Deodorizing and debriding                       | 16 |
| 1.3.4.5 Ant-inflammatory activity of honey                     | 17 |
| <b>1.3.4.6</b> Wound healing with stimulation/immunomodulation |    |
| <b>1.3.4.7</b> Honey wound healing clinical trials             | 19 |
| <b>1.4</b> Infection and immunity                              | 19 |
| 1.4.1Wound healing   | 21 |
| <b>1.4.2</b> Anti-inflammatory cytokines                       | 21 |
| <b>1.4.3</b> Pro-inflammatory cytokines                        | 21 |
| 1.4.4 Tissue re-growth   | 22 |
| 1.5 Aims of this project                                       | 23 |
| Chapter 2 Materials and methods                                | 24 |
| 2.1 Materials  | 24 |
| 2.1.1 Chemicals and reagents                                   | 24 |
| 2.1.2 Other materials  | 25 |
| <b>2.1.3</b> Cell line   |    |

| 2.2 Solutions and media   |
|---|
| <b>2.3</b> Honey samples  |
| 2.4 General Methods   |
| <b>2.4.1</b> Preparation of honey samples   |
| <b>2.4.2</b> Preparation of artificial honey (AH)   |
| 2.4.3 Treatment of honey samples  |
| <b>2.4.3.1</b> Size fractionation of honey samples  |
| 2.4.3.2 Heat treatment of honey samples and LPS   |
| <b>2.4.3.3</b> Treatment of honey samples with PmB  |
| <b>2.5</b> <i>In vitro</i> analysis   |
| <b>2.5.1</b> Culturing of THP-1 cells   |
| <b>2.5.2</b> Preparation of cells for frozen storage  |
| <b>2.5.3</b> Thawing cells  |
| <b>2.5.4</b> Cytotoxicity of honey samples  |
| <b>2.6</b> <i>In vitro</i> assay for immunostimulation  |
| <b>2.6.1</b> THP-1 cells differentiation  |
| 2.6.2 Optimization LPS dose   |
| <b>2.6.3</b> Optimization of capture and detection antibodies for ELISA                         |
| <b>2.7</b> Treatment of honey samples for <i>in vitro</i> cell-based assay                      |
| <b>2.7.1</b> Effect of honey with and without LPS on TNF- $\alpha$ production in THP-1 cells37  |
| 2.7.2 Effect of PmB treated honey samples with and without added LPS                            |
| <b>2.7.3</b> Effect of heat treated honey samples with and without added LPS                    |
| <b>2.7.4</b> Effect of combined treatments (heat and PmB) of honey samples with and without LPS |
| <b>2.7.5</b> Effect of irradiated honey samples with and without LPS                            |

| 2.7.6 Effect of size fractioned honey samples alone or with added LPS  | 43          |
|--|-------------|
| 2.8 Analytical methods   | 44          |
| <b>2.8.1</b> Enzyme linked immune sorbent assay (ELISA) for the measurement of t necrosis factor (TNF- $\alpha$ ) production | umour<br>44 |
| 2.8.2 Limulus Amebocyte Lysate (LAL) assay   | 45          |
| <b>2.8.3</b> Effect of TNF-α production from arabinogalactan (AG)  | 46          |
| 2.8.4 Carbohydrate analysis  | 47          |
| 2.8.4.1 Centrifugal concentration  | 47          |
| 2.8.4.2 Monosaccharide analysis  | 47          |
| 2.8.4.3 Constituent sugar analysis   | 48          |
| 2.8.4.4 Glycosyl linkage analysis  | 48          |
| 2.8.4.5 Proton nuclear magnetic resonance (NMR) spectroscopy   | 49          |
| 2.8.4.6 Size-exclusion chromatography  | 49          |
| 2.9 Data Analysis  | 49          |
|  |             |

### Chapter 3 The stimulation of tumor necrosis factor alpha (TNF-a) production by active honey 50

| 3.0 Introduction                        | 50 |
|---|----|
| 3.1 Cytotoxic activity of honey samples | 50 |
| 3.1.1 Introduction                      | 50 |
| <b>3.1.2</b> Method                     | 51 |
| 3.1.3 Results and Discussion            | 51 |
| <b>3.2</b> Optimization of TNF-α ELISA  | 55 |
| 3.2.1 Introduction                      | 55 |
| <b>3.2.2</b> Method                     | 55 |

| <b>3.2.3</b> Results and Discussion   | 56  |
|---|-----|
| <b>3.3</b> Optimization of TNF- $\alpha$ cytokine production from THP-1 cells | s57 |
| <b>3.3.1</b> Differentiation of THP-1 cells with PMA                          |     |
| 3.3.1.1 Introduction  |     |
| <b>3.3.1.2</b> Method   | 59  |
| 3.3.1.3 Results and Discussion  | 59  |
| <b>3.3.2</b> Response of differentiated THP-1 cells to LPS                    | 60  |
| <b>3.3.2.1</b> Introduction   | 60  |
| <b>3.3.2.2</b> Method   | 61  |
| <b>3.3.2.3</b> Results and Discussion   | 61  |
| <b>3.4</b> Effect of honeys on TNF- $\alpha$ production by THP-1 cells        | 62  |
| 3.4.1 Introduction  | 62  |
| <b>3.4.2</b> Method   | 63  |
| 3.4.3 Results and Discussion  | 63  |
| 3.5 Conclusion  | 65  |

# Chapter 4 Characterization of active component(s) in honey that stimulatetumor necrosis factor (TNF)-a production66

| 4.0 Introduction  | 66 |
|---|----|
| 4.1 Effect of pre-treatment with PmB on activity of honey samples | 67 |
| 4.1.1 Introduction  | 67 |
| <b>4.1.2</b> Method   | 68 |
| 4.1.3 Results and Discussion                                      | 68 |
| <b>4.2</b> Effect of heat treatment on activity of honey samples  | 70 |
| 4.2.1 Introduction  | 70 |

| <b>4.2.2</b> Method  | 70      |
|--|---------|
| <b>4.2.3</b> Results and Discussion  | 71      |
| 4.3 Effect of combined treatments (heat and PmB) on activity of honey sa                       | mples73 |
| <b>4.3.1</b> Introduction  | 73      |
| <b>4.3.2</b> Method  | 73      |
| 4.3.3 Results and Discussion   | 73      |
| <b>4.4</b> Effect on TNF- $\alpha$ production of irradiated honey sample                       | 75      |
| <b>4.4.1</b> Introduction  | 75      |
| <b>4.4.2</b> Method  | 75      |
| 4.4.3 Results and Discussion   | 76      |
| 4.5 Effect of fractioned honey samples   | 77      |
| <b>4.5.1</b> Introduction  | 77      |
| <b>4.5.2</b> Method  | 78      |
| 4.5.3 Results and Discussion   | 79      |
| 4.6 Conclusion   | 82      |
| Chapter 5 Chemical characterization and identification of immunostimulatory component of honey | 84      |
| 5.0 Introduction   | 84      |
| 5.1 Measurement of LPS in honey  | 85      |
| 5.1.1 Introduction   | 85      |
| <b>5.1.2</b> Method  | 85      |
| 5.1.3 Results and Discussion   | 85      |
| 5.3. Carbohydrate (polysaccharide) analysis  | 89      |
| 5.3.1 Introduction   |         |

| <b>5.3.2</b> Method  | 89           |
|--|--------------|
| 5.3.3 Results and Discussion   | 90           |
| <b>5.3.3.1</b> Centrifugal concentration   | 90           |
| <b>5.3.3.2</b> Constituent sugar analysis  | 91           |
| <b>5.3.3.3</b> Glycosyl linkage analysis   | 92           |
| <b>5.3.3.4</b> Proton nuclear magnetic resonance (NMR) spectroscopy                    | 94           |
| <b>5.3.3.5</b> Size-exclusion chromatography   | 97           |
| <b>5.5</b> Effect on TNF- $\alpha$ production of the arabinogalactan component of Kanu | ıka honey.98 |
| 5.5.1 Introduction   | 98           |
| 5.5.2 Method   | 99           |
| 5.5.3 Results and Discussion   | 99           |
| 5.6 Conclusions  | 101          |
| Chapter 6 Summary  | 102          |
| 6.1 Summary  | 103          |
| Chapter 7 Conclusions and Future study   | 104          |
| 7.1 Conclusions  | 104          |
| <b>7.2</b> Future study  | 104          |
| Appendices   | 106          |
| Appendix I: Publication  | 106          |
| Appendix II: Data analysis   | 108          |
| References   | 116          |

### List of Figures

| Figure 1.1     | Ire 1.1Flow diagrams summarizing the activities performed in a wound when |        |
|----------------|---|--------|
|                | honey is applied  | 14     |
| Figure 2.1     | Summary of honey sample treatments to determine the active                |        |
|                | component   | 31     |
| Figure 2.2     | Summary of in vitro immunostimulation protocol                            | 34     |
| Figure 2.3     | Summary of treatments with and without lipopolysaccharide (LF             | PS) in |
|                | in vitro based assays   | 37     |
| Figure 3.1 (a) | Effect of camptothecin on cell viability                                  | 52     |
| Figure 3.1 (b) | Effect of artificial honey on cell viability                              | 52     |
| Figure 3.2     | Cytotoxicity effects of 16 honey samples on THP-1 cells                   | 54     |
| Figure 3.3     | Cytotoxicity effects of 16 honey samples (0.5 % (w/v)) expresse           | d      |
|                | relative to negative control (100 %) and positive control (10 $\mu$ M     |        |
|                | camptothecin)   | 55     |
| Figure 3.4     | Optimizing the tumor necrosis factor- $\alpha$ ELISA by varying capture   | re and |
|                | detection antibodies using high and low concentrations of tumor           |        |
|                | necrosis factor-α standard  | 56     |
| Figure 3.5     | The standard curve of tumor necrosis factor- $\alpha$                     | 57     |
| Figure 3.6     | Effects of differentiating THP-1 cells with phorbol 12-myristate          | 13-    |
|                | acetate (PMA) on magnitude of lipopolysaccharide (LPS) -stimu             | lated  |
|                | TNF- $\alpha$ production  | 58     |
| Figure 3.7     | Structure of phorbol 12-myristate 13-acetate                              | 58     |

| Figure 3.8             | <b>3.8</b> THP-1 cells before and after differentiation with 10 nM phorbol 12-  |             |
|------------------------|---|-------------|
|                        | myristate 13-acetate  | 59          |
| Figure 3.9             | Structure of lipopolysaccharide   | 60          |
| Figure 3.10            | Tumor necrosis factor- $\alpha$ production by THP-1 cells in response to  |             |
|                        | lipopolysaccharide dose after differentiation by 3 days exposure to   | 10          |
|                        | nM (or) 50 nM phorbol 12-myristate 13-acetate   | 61          |
| Figure 3.11            | Tumor necrosis factor- $\alpha$ production by THP-1 cells in response to  |             |
|                        | varying concentrations of lipopolysaccharide after differentiation b  | oy 3        |
|                        | days exposure to 10 nM phorbol 12-myristate 13-acetate  | 62          |
| <b>Figure 3.12 (a)</b> | Tumor necrosis factor- $\alpha$ production from differentiated THP-1 cell stimulated by different honey samples, lipopolysaccharide and con | ls<br>trols |
|                        |   | 64          |
| <b>Figure 3.12 (b)</b> | Tumor necrosis factor- $\alpha$ production from differentiated THP-1 cell   | ls          |
|                        | stimulated by different honey samples in the absence and presence   | of          |
|                        | lipopolysaccharide  | 65          |
| Figure 4.1             | Tumor necrosis factor- $\alpha$ production from differentiated THP-1 cell   | ls          |
|                        | after treatment with lipopolysaccharide and honey samples with an   | ıd          |
|                        | without pre-treatment with polymixin B  | 69          |
| Figure 4.2 (a)         | Tumor necrosis factor- $\alpha$ production from differentiated THP-1 cell   | ls          |
|                        | after treatment with lipopolysaccharide and final concentration of  |             |
|                        | various honeys before and after heat treatment (80°C)   | 71          |
| Figure 4.2 (b)         | Tumor necrosis factor- $\alpha$ production from differentiated THP-1 cell   | ls          |
|                        | after heat treatment of honey samples with and without  |             |
|                        | lipopolysaccharide  | 72          |

| Figure 4.3      | e 4.3Effect of tumor necrosis factor-α production from differentiated THP-1<br>cells stimulated with lipopolysaccharide alone, heat treated |             |
|-----------------|---|-------------|
|                 |   |             |
|                 | lipopolysaccharide alone and the final concentration of two   | heat        |
|                 | treated honey samples before and after polymixin B  | 74          |
| Figure 4.4 (a)  | Effect on tumor necrosis factor- $\alpha$ production from differenti  | ated THP-   |
|                 | 1 cells after Kanuka blend honey sample was treated with in   | creasing    |
|                 | doses of gamma irradiation  | 76          |
| Figure 4.4 (b)  | Effect on tumor necrosis factor- $\alpha$ production from differenti  | ated THP-   |
|                 | 1 cells of Kanuka blend honey sample was treated with diffe   | erent doses |
|                 | of gamma irradiation in the presence and absence of   |             |
|                 | lipopolysaccharide  | 77          |
| Figure 4.5      | Structure of lipopolysaccharide and proteins  | 78          |
| Figure 4.6 (a)  | Effect of tumor necrosis factor- $\alpha$ production from the difference  | entiated    |
|                 | THP-1 cells after treatment with lipopolysaccharide and Kan   | nuka young  |
|                 | honey sample without fractionation and fractionated honey   | sample 79   |
| Figure 4.6 (b)  | Effect on tumor necrosis factor- $\alpha$ production from the differ  | entiated    |
|                 | THP-1 cells after treatment with lipopolysaccharide and wit   | h Kanuka    |
|                 | old honey sample without fractionation and fractionated hor   | ey sample   |
|                 |   | 80          |
| Figure 4.6 (a1) | Effect on tumor necrosis- $\alpha$ production of fractions from Kar   | uka young   |
|                 | honey with and without lipopolysaccharide of different mole   | ecular      |
|                 | weight fractions  | 81          |
| Figure 4.6 (b1) | Effect of tumor necrosis- $\alpha$ production of fractionated Kanuk   | a old       |
|                 | honey with and without lipopolysaccharide of different mole   | ecular      |
|                 | weight fractions  | 82          |
| Figure 5.1(a)   | The correlation between stimulated tumor necrosis factor- $\alpha$  | by original |
|                 | honey samples and lipopolysaccharide levels in honey  | 87          |

| Figure 5.1 (b) | the correlation between stimulated tumor necrosis factor- $\alpha$ by heat  |      |
|----------------|---|------|
|                | treated honey samples and lipopolysaccharide levels in honey                | 87   |
| Figure 5.2     | Summary of various treatments carried out at Industrial Research            |      |
|                | Limited to identify the arabinogalactan active component in honey           | 90   |
| Figure 5.3     | Proton nuclear magnetic resonance spectra of five honey samples             | 94   |
| Figure 5.4     | The structural model of $\beta$ -arabinogalactan-protein from honey         | 99   |
| Figure 5.5     | Effect of tumor necrosis factor- $\alpha$ production from differentiated TI | HP-1 |
|                | cells with purified arabinogalactan from honey                              | 100  |

### List of Tables

| Table 2.1 | New Zealand honey samples analyzed in the study   | 28                 |
|-----------|---|--------------------|
| Table 2.2 | Phenolic compounds and methylglyoxal present in New Zea<br>Manuka, Kanuka, Rewarewa and Clover honeys   | land<br>29         |
| Table 2.3 | Preparation of Tumor necrosis factor- $\alpha$ standard concentrations  | 36                 |
| Table 5.1 | Quantification of endotoxin levels in honey samples   | 86                 |
| Table 5.2 | Yield of high molecular weight material from honey samples through Vivaspin 15R cartridges  | put<br>91          |
| Table 5.3 | Constituent sugars composition high molecular weight fraction<br>honey samples analyzed using the reductive hydrolysis method   | s of<br>91         |
| Table 5.4 | Glycosyl linkage composition high molecular weight fractions<br>honey samples analyzed by gas chromatography-mass spectroso<br>(GC-MS) of partially methylated alditol acetates | s of<br>copy<br>93 |

## List of Abbreviations

| NH <sub>4</sub> OH                 | Ammonium hydroxide                         |
|------------------------------------|--|
| ATCC                               | American Type Culture Collection           |
| ANOVA                              | Analysis of variance                       |
| AG                                 | Arabinogalactan                            |
| AGP                                | Arabinogalactan-protein                    |
| AH                                 | Artificial honey                           |
| BSA                                | Bovine serum albumin                       |
| (CH <sub>3</sub> ) <sub>2</sub> SO | Camptothecin                               |
| CO <sub>2</sub>                    | Carbon dioxide                             |
| °C                                 | Degrees Celsius                            |
| DMSO                               | Dimethylsulfoxide                          |
| KH <sub>2</sub> HPO <sub>4</sub>   | Dipotassium hydrogen orthophosphate        |
| Na <sub>2</sub> HPO <sub>4</sub>   | Disodium hydrogen orthophosphate anhydrous |
| ECM                                | Extra cellular matrix                      |
| ESR                                | Environmental Science Research             |
| ELISA                              | Enzyme-linked immune sorbent assay         |
| FBS                                | Fetal bovine serum                         |
| GC-MS                              | Gas chromatography-mass spectrometry       |
| g                                  | Gram or Acceleration due to gravity        |
| >                                  | Greater than                               |
| HPAEC                              | High-performance anion-exchange            |
|                                    | chromatography                             |
| h                                  | Hour                                       |
| THP-1                              | Human monocytic leukemia cells             |
| HMF                                | Hydroxymethylfurfural                      |
| HRP                                | Horse radish peroxidase                    |
| $H_2O_2$                           | Hydrogen peroxide                          |
| IRL                                | Industrial Research Limited                |
| IU                                 | International Unit                         |
| IL-1                               | Interleukin-1                              |

| IL-6              | Interleukin-6                       |
|-------------------|-------------------------------------|
| Kg                | Kilogram                            |
| KGy               | kilo Grays                          |
| KDa               | Kilo Daltons                        |
| <                 | Less than                           |
| L                 | Litre                               |
| LPS               | Lipopolysaccharide                  |
| Ltd               | Limited                             |
| LiNO <sub>3</sub> | Lithium nitrate                     |
| рН                | -Log $[H^+]$                        |
| MS                | Mass spectroscopy                   |
| MHz               | Megahertz                           |
| MGO               | Methylglyoxide                      |
| CH <sub>3</sub> I | Methyl iodide                       |
| MeOH              | Methanol                            |
| ML                | Millilitre                          |
| μL                | Microliter                          |
| mg                | Milligram                           |
| μm                | Micrometer                          |
| μΜ                | Micromolar                          |
| mM                | Millimolar                          |
| mm                | Millimetre                          |
| min               | Minutes                             |
| Μ                 | Molar                               |
| ng                | Nanogram                            |
| nm                | Nanomolar                           |
| N <sub>2</sub>    | Nitrogen                            |
| NMR               | Nuclear magnetic resonance          |
| ppm               | Part per million                    |
| %                 | Percent                             |
| % (w/v)           | Percent by weight per volume        |
| PMA               | Phorbol 12-myristate 13-acetate     |
| PBS               | Phosphate Buffered Saline           |
| PBS-T             | Phosphate Buffered Saline -Tween 20 |
|                   |                                     |

| PmB                              | Polymixin B  |
|----------------------------------|--|
| rpm                              | Revolution per minute  |
| RPMI                             | Roswell park memorial institute medium   |
| Complete RPMI                    | RPMI medium supplemented with 10 % FBS,  |
|                                  | penicillin at 50 IU units/mL and streptomycin                                      |
|                                  | at 5 µg/mL   |
| SEC                              | Size-exclusion chromatography  |
| NaN <sub>3</sub>                 | Sodium azide   |
| NaBD <sub>4</sub>                | Sodium borodeuteride   |
| NaH <sub>2</sub> PO <sub>4</sub> | Sodium dihydrogen phosphate monohydrate  |
| TMB                              | Tetra methyl benzidine   |
| TFA                              | Tri fluoro acetic acid   |
| TNF-α                            | Tumor necrosis factor  |
| USP                              | United States Pharmacopeia   |
| v/v                              | Volume per volume  |
| WST-1 reagent                    | 4-[3-(4-iodophenyl)-2-(4-nitrophenyl)-2H-5-<br>tetrazolio]-1, 3-benzenedisulfonate |