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Institute of Fundamental Sciences

A comparison of next-generation sequencing protocols for microbial profiling

A thesis

submitted in partial fulfillment
of the requirements for the degree of
Master of Science in Genetics

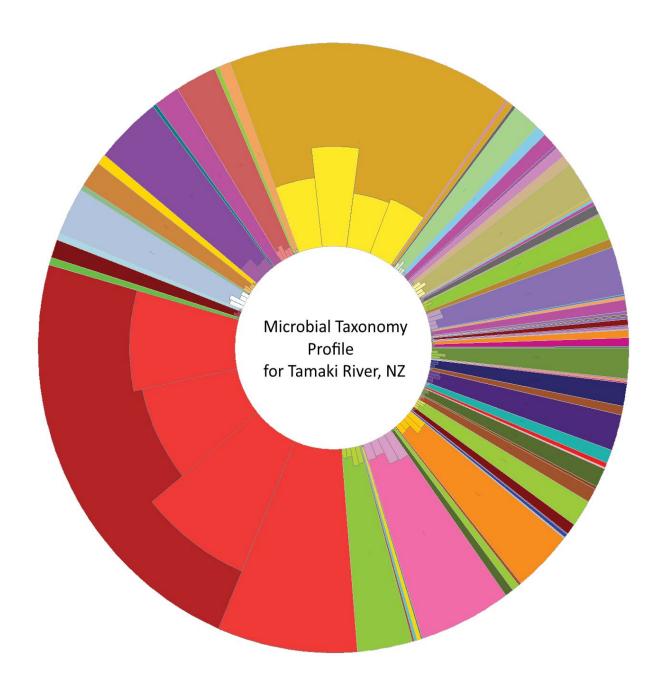
By

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2016

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One of the responsibilities faced by the Environmental Genome Project is to provide the science base upon which society can make better informed risk management decisions.

-Samuel Wilson-

Abstract

The introduction of massive parallel sequencing has revolutionized analyses of microbial communities. Illumina and other Whole Genome Shotgun Sequencing (WGS) sequencing protocols have promised improved opportunities for investigation of microbial communities. In the present work, we compared and contrasted the findings from different NGS library preparation protocols (Illumina Nextera, Nextera-XT, NEXTFlex PCR-free and Ion-Xpress-400bp) and two sequencing platforms (MiSeq and Ion-Torrent). Short reads were analysed using the rapid database matching software PAUDA and visualization software MEGAN5, which provides a conservative approach for taxonomic identification and functional analyses. In analyses of a Tamaki River water sample, biological inferences were made and compared across platforms and protocols. For even a relatively small number of reads generated on the MiSeq sequencing platform important pathogens were identified in the water sample. Far greater phylogenetic resolution was obtained with WGS sequencing protocols than has been reported in similar studies that have used 16S rDNA Illumina sequencing protocols. TruSeq and Nextera-XT sequencing protocols produced similar results. The latter protocol offered cheaper, and faster results from less DNA starting material. Proteobacteria (alpha, beta and gamma), Actinobacteria and Bacteroidetes were identified as major microbial elements in the Tamaki River sample. Our findings support the emerging view that short read sequence data and enzymatic library prep protocols provide a cost effective tool for evaluating, cataloguing and monitoring microbial species and communities. This is an approach that complements, and provides additional insight to microbial culture "water testing" protocols routinely used for analysing aquatic environments.

Acknowledgement

There are many people I would like to express my gratitude and cordial thanks in helping me out in preparing my Master's Thesis. This dissertation would not have been possible without your support and strong collaboration between different academia backgrounds.

To my lovely wife, I know I could not have done this without your constant encouragement and great patience at all times. I know that during this challenging period, you have been understanding and have given me much help, showered me with your love and support and there are no words to express my appreciation for having you by my side. To my family members; my parents, in-laws and brother, I would like to send my appreciation and would like to say thank you for being patient and for your unequivocal moral support during my master degree course. To my mentor Trish McLenachan, without you my thesis would be incomplete! Thank you for your valuable guidance, advice and patience with my writing. I send you immeasurable and deepest gratitude for your contribution in making this study worthwhile and possible.

To my principal supervisor Professor Peter-James Lockhart, co-supervisor Professor Nigel French and my bioinformatics co-supervisor Dr Patrick Biggs, I would like to thank you for being very supportive, understanding, patient and for providing precious academic and technical advice. To my principal supervisor Peter, I would like to thank you for your guidance throughout my course especially in having faith in me to finish my dissertation on time. Despite project challenges you have continued to encourage me with your knowledge and invaluable thoughts both on an academic and personal level. Special thanks to Dr Patrick Biggs for your continual support and enlightenment regarding bioinformatics analyses.

For financial support, I would like to acknowledge and thank the Institute of Veterinary, Animal and Biomedical Sciences (IVABS), Institute of Fundamental Sciences (IFS), Ministry of Health (MOH), Protozoal Research Unit (PRU, Hopkirk Research Institute). Lastly to all my fellow friends and colleagues, you guys kept me going and most importantly were understanding and patient with my workload and study. You guys are awesome!

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List of Acronyms

% Percent

°C Degrees Celsius

μl Microlitre(s)

μ**M** Micromolar

100 PE 2 x 100 base pair paired-end read

150 PE 2 x 150 base pair paired-end read

250 PE 2 x 250 base pair paired-end read

300 PE 2 x 300 base pair paired-end read

A Adenine

A260 Nanodrop absorbance at 260 nanometres

A280 Nanodrop absorbance at 280 nanometres

AFLP Amplified Fragment Length Polymorphism

ATL A-Tailing Mix

ATM Amplicon Tagment Mix

ATP Adenosine Triphosphate

BAM Binary Alignment Matrix

BGI Beijing Genomics Limited

BIPES Illumina Multiplexed Paired-end Sequencing Adapter

BLAST Basic Local Alignment Search Tool

bp Base pair(s)

C Cytosine

CCD Charge-coupled Device

cDNA Complementary Deoxyribonucleic Acid

contig Continuous Sequence

CTA A-Tailing Control

CTE End-Repair Control

CTL Ligation Control

ddNTP Dideoxy Nucleotide Triphosphate

dH₂O Distilled Water

DNA Deoxyribonucleic Acid

dNTP Deoxy Nucleotide Triphosphate

ds Double Stranded

EB Elution Buffer

eDNA Environmental Deoxyribonucleic Acid

EDTA Ethylenediamine Tetra-Acetic Acid

emPCR Emulsion Polymerase Chain Reaction

ERP End-Repair mix

EtBr Ethidium Bromide

E-value A parameter that describes the number of expected matches when searching a

sequence database of a particular size and composition

FC Flowcell

fq Fastq File Format

g Gram(s)

G Guanine

Gb Gigabytes

gDNA Genomic DNA

HiFi High fidelity enyzme

HMW High Molecular Weight

HT1 Hybridization Buffer

Inc. Incorporated

Indel Small Insertion or deletion

ISFET Ion Sensitive Field Effect Transistor

KEGG Kyoto Encyclopedia of Genes and Genomes

LCA Lowest Common Ancestor

LIG Ligation Mix

Log¹⁰ Logarithm to the base 10

M Molar

Mb Megabytes

MDA Multiple Displacement Amplification

MEGAN Metagenome Analyzer

MGS Massey Genome Service

min Minute(s)

ml Millilitre(s)

mm Millimetre(s)

mM Millimolar

MPSS Massive Parallel Signature Sequencing

mRNA Messenger Ribonucleic Acid

mtDNA Mitochondrial Deoxyribonucleic Acid

ng Nanogram(s)

NGS Next-generation Sequencing

No Number

NPM Nextera PCR Master Mix

NPS Non-point Source

nt Nucleotide

NT Neutralize Tagment Buffer

NZGL New Zealand Genomics Limited

OTU Operational Taxonomic Unit

PAUDA Protein Alignment Using a DNA Aligner

PCoA Principal Coordinate Analysis

PCR Polymerase chain reaction

pDNA Pseudo DNA

PE Paired-end

PGM Personal Genome Machine

PhiX Bacteriophage PhiX174

PMM PCR Master Mix

pmol Picomole(s)

PPC PCR Primer Cocktail

PP_i Pyrophosphate

Q₁₀ Phred Quality Score 1 error in 10

Q₂₀ Phred Quality Score 1 error in 100

Q₃₀ Phred Quality Score 1 error in 1000

QC Quality Control

qPCR Quantitative Polymerase Chain Reaction

Q-score Phred Quality Score

RNA Ribonucleic Acid

rpm Revolutions per Minute

rRNA Ribosomal Ribonucleic Acid

RSB Resuspension Buffer

RTA Real-Time Analysis

s Second(s)

SAM Sequence Alignment Map

SBS Sequencing by Synthesis

SCIMM Sequence Clustering with Interpolated Markov Models

SEED Database infrastructure for comparative genomics in MEGAN5 software

SMRT Single Molecule Real Time

spp. Species

SPRI Solid Phase Reversible Immobilization

ss Single Stranded

STL Stop Ligation Buffer

T Thymine

TAE Tris-Acetate EDTA buffer

TAP Taxonomic Assignment Pipeline

Taq Thermus aquaticus

TB Tuberculosis

TD Tagmentation Buffer

TE Tris EDTA Buffer

V Volts

WGS Whole Genome Shotgun Sequencing

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