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## Photobioreactor production of microalgae for potential fuel oils

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Tiyaporn Luangpipat

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#### **Abstract**

This work focussed on a detailed characterization of the freshwater microalga *Chlorella vulgaris* as a producer of potential fuel oils. Uniquely, growth and oil production of *C. vulgaris* were characterized in full strength seawater-based media, something that has not been previously reported. *C. vulgaris* was selected for a detailed study after a screening of six potential oil producing microalgae. For photoautotrophic growth, always under carbon sufficiency and at normal growth temperature, the characterization study covered: the biomass growth rate; lipid content in the biomass; productivities of the lipids and the biomass; the biomass loss in the dark; the lipid/biomass yields on macronutrients; and the energy content of the biomass. The above key production parameters were characterized in a purpose-built tubular photobioreactor (~80 L) and in stirred tank photobioreactors (~7.5 L) under conditions of nitrogen sufficiency and at various levels of nitrogen limitation. Production was evaluated in both batch and continuous cultures at various dilution rates using indoor light to mimic sunlight. The production temperature mimicked the relatively warm conditions that would be encountered in a potential production system located outdoors in a tropical climate.

In seawater media at 25–27 °C, *C. vulgaris* was shown to have a crude oil productivity of >37 mg L<sup>-1</sup> d<sup>-1</sup> and the energy content of the biomass could exceed 25 kJ g<sup>-1</sup>, depending on the culture conditions. Both these values were high compared with the reported data for this alga in freshwater media. Compared with continuously illuminated culture, day–night cycling of irradiance reduced oil productivity by ~31%, but the energy content of the biomass were reduced by only about 8%. In seawater, the alga could be grown as rapidly and stably as in freshwater. The lipid content of the biomass commonly exceeded 30% by dry weight and in exceptional cases a lipid content of more than 50% (by weight)

was achieved. Biomass calorific values of  $\geq 27$  kJ g<sup>-1</sup> could be attained in some cases. Nitrogen starvation enhanced the lipid contents of the biomass by >3-fold relative to the lipid contents for the nonstarved case. Steady-state continuous cultures were shown to be possible. Both batch and continuous operations were feasible, especially in stirred tanks, but the culture was more failure prone, or relatively less productive, in the tubular photobioreactor.

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## **Table of Contents**

Abstract	iii
Acknowledgements	V
Table of Contents	vii
List of Figures	xi
List of Tables	xvii
Abbreviations	xix
Chapter 1	1
Introduction	1
Chapter 2	5
Literature Review	5
2.1 Microalgae and their culture	5
2.2 Attributes of a commercial alga for production of fuel oils	5
2.3 Microalgae versus plants	9
2.4 Algae culture requirements	12
2.4.1 Culture media.	13
2.4.2 Temperature	15
2.4.3 pH	17
2.4.4 Light	17
2.5 Algal oils	21
2.6 Culture systems for microalgae	33
2.6.1 Open culture systems	33
2.6.2 Closed photobioreactors	36
2.7 Objectives	43

Chapter 3	45
Materials and Methods	45
3.1 General methodology	45
3.2 Microalgae	46
3.3 Growth media	50
3.3.1 BG 11 culture medium	50
3.3.2 Seawater	52
3.4 Inoculum preparation and Duran bottle cultures	53
3.5 Tubular photobioreactor	54
3.6 Stirred tank photobioreactor	59
3.6.1 Batch culture	59
3.6.2 Continuous culture	60
3.7 A 10 L stirred photobioreactor (Corning photobioreactor)	62
3.8 Biomass separation by centrifugation	63
3.8.1 Batch centrifugation	63
3.8.2 Continuous centrifugation	64
3.9 Biomass freeze drying	65
3.10 Measurements	66
3.10.1 Biomass concentration	66
3.10.2 Irradiance	72
3.10.3 Nitrate	74
3.10.4 Phosphate	76
3.10.5 Total lipids	78
3.10.6 Neutral lipids	79
3.10.7 Triglyceride in neutral lipids	79

3.10.8 Calorific values	80
3.10.9 Nile red staining of cells (modified from Elsey et al., 2007)	80
3.10.10 Gram staining (Mudili, 2007; Cappuccino & Natalie, 2011)	81
3.10.11 Algae morphology	81
3.10.12 Calculations of kinetic parameters (Doran, 1995; Shuler & Kargi, 2002)	81
Chapter 4	85
Results and Discussion	85
4.1 Studies in Duran bottles	85
4.1.1 Salinity effects on growth	85
4.1.2 Light/dark cycle effects on <i>Chlorella vulgaris</i>	96
4.1.3 <i>Chlorella vulgaris</i> biomass loss in the dark	100
4.2 Culture profiles in tubular photobioreactor	103
4.2.1 Batch cultures	112
4.2.1.1 Effect of nitrate concentration	112
4.2.1.2 Effect of light	117
4.2.1.3 Effect of copper	118
4.2.2 Continuous cultures	121
4.3 Culture profiles in stirred-tank bioreactor	124
4.3.1 Batch cultures	137
4.3.2 Continuous cultures	146
4.4 Investigation of MGA-1-NZ cultures	156
4.4.1 MGA-1-NZ culture in Duran bottles	156
4.4.2 MGA-1-NZ culture in 10 L Corning stirred photobioreactor	157
4.5 Characteristics of the algal lipids	165
4.5.1 Chlorella vulgaris	166

4.5.1.1 Concentrations of elements in Chlorella vulgaris total lipid extract	166
4.5.1.2 Fractionation of Chlorella vulgaris total lipid extract	167
4.5.1.3 Fatty acid profiles of Chlorella vulgaris total lipid extract	169
4.5.2 The alga MGA-1-NZ	175
4.5.2.1 Concentrations of elements in MGA-1-NZ total lipid extract	175
4.5.2.2 Fractionation of MGA-1-NZ total lipid extract	175
4.5.2.3 Fatty acid profiles of MGA-1-NZ total lipid extract	175
4.6 Comparison of the lipid productivity of C. vulgaris	179
Chapter 5	185
Summary and Conclusions	185
5.1 Summary	185
5.2 Conclusions	190
References	193
Appendix 1	225
Standard deviation calculations	225
Appendix 2	227
Experimental data	227

# **List of Figures**

Figure 2.1 A microalga ( <i>Chlorella vulgaris</i> ) with a dispersed cell morphology	6
Figure 2.2 Microalgae with filamentous morphologies	7
Figure 2.3 Oil productivity of various sources.	10
Figure 2.4 A typical photosynthesis response curve.	19
Figure 2.5 Some components of algal crude oil.	23
Figure 2.6 Open systems for growing microalgae.	34
Figure 2.7 Closed systems for growing microalgae.	39
Figure 2.8 A tubular photobioreactor with a single continuous looped tube	41
Figure 3.1 <i>Chlorella vulgaris</i> in BG-11 made in freshwater	47
Figure 3.2 <i>Choricystis minor</i> in BG-11 made in freshwater.	47
Figure 3.3 <i>Neochloris</i> sp. in BG-11 made in freshwater.	48
Figure 3.4 <i>Pseudococcomyxa simplex</i> in BG-11 made in freshwater.	48
Figure 3.5 Scenedesmus sp. in BG-11 made in freshwater	49
Figure 3.6 MGA-1-NZ in BG-11 made in seawater.	49
Figure 3.7 A typical set up for algae growth in gas sparged culture bottles	54
Figure 3.8 Tubular photobioreactor.	55
Figure 3.9 Broth recirculation in the tubular loop photobioreactor	56
Figure 3.10 Main control panel of the tubular photobioreactor	56
Figure 3.11 A stirred tank photobioreactor illuminated using LED lamps.	61
Figure 3.12 Continuous culture in a stirred tank photobioreactor.	61
Figure 3.13 The 10 L Corning stirred photobioreactor	63
Figure 3.14 The cell paste after centrifugation.	64
Figure 3.15 Continuous centrifugation.	65
Figure 3.16 Freeze-dried biomass	66

Figure 3.17 The calibration curve for <i>Chlorella vulgaris</i> in BG-11 made in freshwater68
Figure 3.18 The calibration curve for <i>Chlorella vulgaris</i> in BG-11 made in seawater69
Figure 3.19 The calibration curve for <i>Choricystis minor</i> in BG-11 made in freshwater69
Figure 3.20 The calibration curve for <i>Neochloris</i> sp. in BG-11 made in freshwater70
Figure 3.21 The calibration curve for <i>Pseudococcomyxa simplex</i> in BG-11 made in
freshwater
Figure 3.22 The calibration curve for <i>Scenedesmus</i> sp. in BG-11 made in freshwater71
Figure 3.23 The calibration curve for MGA-1-NZ in BG-11 made in seawater71
Figure 3.24 Relationship between the percentage light output and irradiance (QSL-2101
sensor) for LEDs of tubular photobioreactor
Figure 3.25 Relationship between the percentage light output and irradiance (QSL-2101
sensor) for LEDs units A and B of the stirred tank photobioreactor unit 273
Figure 3.26 Relationship between the percentage light output and irradiance (QSL-2101
sensor) for LEDs units C and D of the stirred tank photobioreactor unit 1
Figure 3.27 A nitrate calibration curve for BG-11 made with freshwater
Figure 3.28 A nitrate calibration curve for BG-11 made with seawater
Figure 3.29 A phosphate calibration curve.
Figure 4.1 Growth curves of <i>Chlorella vulgaris</i> in different BG11 formulations in 1 L
culture bottles
Figure 4.2 Growth curves of <i>Choricystis minor</i> in different BG11 formulations in 1 L culture
bottles86
Figure 4.3 Growth curves of <i>Neochloris</i> sp. in different BG11 formulations in 1 L culture
bottles87
Figure 4.4 Growth curves of <i>Pseudococcomyxa simplex</i> in different formulations of BG11 in
1 L culture bottles.

Figure 4.5 Growth curves of <i>Chlorella vulgaris</i> in different BG11 formulations in 2 L	
culture bottles	89
Figure 4.6 Growth curves of <i>Choricystis minor</i> in different BG11 formulations in 2 L c	ulture
bottles	90
Figure 4.7 Growth curves of <i>Neochloris</i> sp. in different BG11 formulations in 2 L cultures	re
bottles	90
Figure 4.8 Growth curves of <i>Psuedococcomyxa simplex</i> in different formulations of BC	611 in
2 L culture bottles.	91
Figure 4.9 Growth curves of <i>Scenedesmus</i> sp. in different formulations of BG11 in 2 L	
culture bottles	91
Figure 4.10 Growth curves of <i>Chlorella vulgaris</i> in BG11 made with seawater, illumina	ated
continuously (24/0) and every 12 h (12/12) by fluorescent lamps.	97
Figure 4.11 <i>Chlorella vulgaris</i> biomass loss in the dark during incubation at various	
temperatures. At 25 °C, the data sets marked (1) and (2) are from two parallel flasks	100
Figure 4.12 Plots of $ln(C/C_0)$ versus time for <i>C. vulgaris</i> at various temperatures in the	dark.
At 25 °C, the data sets marked (1) and (2) are from two parallel flasks.	102
Figure 4.13 Culture profile of $C$ . $vulgaris$ in the run PBR 1 with 1133 mg $L^{-1}$ of initial	
nitrate concentration.	112
Figure 4.14 Culture profile of $C$ . $vulgaris$ in the run PBR 9 with 537 mg $L^{-1}$ of initial n	itrate
concentration and 0.02 mg L <sup>-1</sup> of copper concentration.	113
Figure 4.15 Culture profile of $C$ . $vulgaris$ in the run PBR 13 with 291 mg L <sup>-1</sup> of initial	
nitrate concentration. See Table 4.6 for further details.	113
Figure 4.16 Broth foaming in the degassing column.	114
Figure 4.17 Culture profile of $C$ . $vulgaris$ in the run PBR 10 with 1372 mg L <sup>-1</sup> of initial	1
nitrate concentration under a light level of 314-458 µmol m <sup>-2</sup> s <sup>-1</sup>	116

Figure 4.18 Culture profile of <i>C. vulgaris</i> in the run PBR 14 with 236 mg L <sup>-1</sup> of initial
nitrate concentration under a light level of 314-458 μmol m <sup>-2</sup> s <sup>-1</sup> 116
Figure 4.19 Culture profile of <i>C. vulgaris</i> in the run PBR 3 with 1088 mg $L^{-1}$ of initial
nitrate concentration under a light level of 251-361 μmol m <sup>-2</sup> s <sup>-1</sup>
Figure 4.20 Culture profile of <i>C. vulgaris</i> in the run PBR 6 without copper under a light leve
of 115-171 μmol m <sup>-2</sup> s <sup>-1</sup> .
Figure 4.21 Continuous growth profile of $C$ . $vulgaris$ in PBR 8 at a dilution rate of 0.3 d <sup>-1</sup> .
Figure 4.22 Continuous growth profile of $C$ . $vulgaris$ in PBR 9 at a dilution rate of 0.12 d <sup>-1</sup> .
Figure 4.23 Growth profile of <i>C. vulgaris</i> in STR 1 with normal nitrate (BG-11 seawater)
under illumination of 292 μmol m <sup>-2</sup> s <sup>-1</sup> by fluorescent lights.
Figure 4.24 Growth profile of <i>C. vulgaris</i> in STR 2 with half nitrate under illumination of
292 μmol m <sup>-2</sup> s <sup>-1</sup> by fluorescent lights.
Figure 4.25 Growth profile of <i>C. vulgaris</i> in STR 4 with half nitrate under illumination of
292 μmol m <sup>-2</sup> s <sup>-1</sup> by fluorescent lights.
Figure 4.26 Growth profile of <i>C. vulgaris</i> in STR 5 with half nitrate under illumination of
292 μmol m <sup>-2</sup> s <sup>-1</sup> by fluorescent lights.
Figure 4.27 Growth profile of <i>C. vulgaris</i> in STR 6 with half nitrate under illumination of
292 μmol m <sup>-2</sup> s <sup>-1</sup> by fluorescent lights.
Figure 4.28 Growth profile of <i>C. vulgaris</i> in STR 10 with normal nitrate under illumination
of 200 μmol m <sup>-2</sup> s <sup>-1</sup> by light-emitting diodes.
Figure 4.29 Culture profile of <i>C. vulgaris</i> in STR 7. On day 31, the dilution rate was set at
0.157 d <sup>-1</sup> . Irradiance level was 292 μmol m <sup>-2</sup> s <sup>-1</sup> (fluorescent light)146

Figure 4.30 Culture profile of <i>C. vulgaris</i> in STR 8. The dilution rate was $0.22 \mathrm{d}^{-1}$ (day 30)
and 0.079 d <sup>-1</sup> (day 55). Irradiance level was 1123 $\mu$ mol m <sup>-2</sup> s <sup>-1</sup> (light emitting diodes) 147
Figure 4.31 Culture profile of <i>C. vulgaris</i> in STR 9. On day 41, the dilution rate was set at
0.079 d <sup>-1</sup> . Irradiance level was 292 μmol m <sup>-2</sup> s <sup>-1</sup> (fluorescent light)
Figure 4.32 Culture profile of <i>C. vulgaris</i> in STR 11 at various dilution rates (Table 4.8).
The incident irradiance levels were 600 and 862 μmol m <sup>-2</sup> s <sup>-1</sup> (light-emitting diodes) as noted
in Table 4.8
Figure 4.33 Culture profile of <i>C. vulgaris</i> in STR 12 at various dilution rates (Table 4.8).
The incident irradiance level were 177 and 76 μmol m <sup>-2</sup> s <sup>-1</sup> (light-emitting diodes) as noted
in Table 4.8
Figure 4.34 Relationship between dilution rate and the steady state biomass productivity. 155
Figure 4.35 Relationship between dilution rate and the steady state lipid productivity 156
Figure 4.36 Culture profiles of MGA-1-NZ in BG11 seawater medium in 2 L Duran bottle.
Figure 4.37 Culture profile of MGA-1-NZ in BR 1
Figure 4.38 Culture profile of MGA-1-NZ in BR 2
Figure 4.39 Culture profile of MGA-1-NZ in BR 3

## **List of Tables**

Table 1.1 Some companies engaged in commercializing algal fuels	3
Table 2.1 Various factors that affect lipid content and composition	29
Table 3.1 BG11 stock 1	51
Table 3.2 BG11 stock 2	51
Table 3.3 BG11 stock 3	51
Table 3.4 BG11 stock 4	52
Table 3.5 Vitamins solution	52
Table 4.1 Kinetic parameters for the algae cultured in 1 L bottles (Figures 4.1-4.4)	88
Table 4.2 Total lipid contents and calorific values for the algae cultured in 2 L bottles	92
Table 4.3 Kinetic parameters for the algae cultured in 2 L bottles	93
Table 4.4 Comparison of continuous illumination and light-dark cycling (12 h/12 h) on	
culture kinetics and biomass properties of Chlorella vulgaris	99
Table 4.5 <i>C. vulgaris</i> biomass consumption rate constant <i>k</i> in the dark at various	
temperatures	103
Table 4.6 Summary of <i>C. vulgaris</i> production in the photobioreactor runs	104
Table 4.7 Total lipid content and calorific value of <i>C. vulgaris</i> biomass produced in the	
photobioreactor	109
Table 4.8 Kinetic parameters of <i>C. vulgaris</i> production in tubular photobioreactor	110
Table 4.9 Summary of <i>C. vulgaris</i> cultures in stirred-tank bioreactors	124
Table 4.10 Total lipid contents and calorific values of <i>C. vulgaris</i> biomass from stirred-t	ank
bioreactors	131
Table 4.11 Culture kinetic parameters for <i>C. vulgaris</i> from stirred-tank bioreactors	133
Table 4.12 Lipid production by <i>C. vulgaris</i> in stirred-tank bioreactors	135
Table 4.13 Summary of biomass concentrations at various steady states	150

Table 4.14 Summary of MGA-1-NZ cultures in 10 L Corning stirred photobioreactor	159
Table 4.15 Total lipid contents and calorific values of MGA-1-NZ biomass	162
Table 4.16 Growth kinetic parameters of MGA-1-NZ	163
Table 4.17 Lipid production parameters of MGA-1-NZ	164
Table 4.18 Concentrations of various elements in C. vulgaris total lipid extract from stir.	red-
tank photobioreactor runs STR 1 and STR 2	166
Table 4.19 Neutral lipid content and triglyceride content of <i>C. vulgaris</i> oils	168
Table 4.20 Fatty acid profiles of <i>C. vulgaris</i> total lipid extract from photobioreactor runs	s .170
Table 4.21 Fatty acid profiles of <i>C. vulgaris</i> total lipid extract from stirred-tank	
photobioreactor runs	172
Table 4.22 Proportions of the various classes of fatty acids in lipids from some tubular	
photobioreactor and stirred-tank photobioreactor runs	174
Table 4.23 Concentrations of elements in crude oil of MGA-1-NZ	175
Table 4.24 Fatty acid profiles of MGA-1-NZ total lipids	177
Table 4.25 Proportions of the various classes of fatty acids in lipids of MGA-1-NZ	179
Table 4.26 The lipid productivity of <i>C. vulgaris</i> in Duran bottles, tubular photobioreactor	rs
and stirred-tank bioreactors	180

### **Abbreviations**

A Area illuminated by light (m<sup>2</sup>)

 $A_{xxx}$  Spectrophotometric absorbance at a wavelength of xxx nm

BR n Corning stirred photobioreactor run n

C Biomass concentration at time t (g L<sup>-1</sup>)

 $C_0$  Biomass concentration at time zero (g L<sup>-1</sup>)

D Dilution rate  $(d^{-1})$ 

DCW Dry cell weight (g L<sup>-1</sup>)

DHA Docosahexaenoic acid

DNA Deoxyribonucleic acid

DO Dissolved oxygen

d Diameter (m)

EPA Eicosapentaenoic acid

F Flow rate of the feed (mL  $d^{-1}$ )

I Irradiance (μmol m<sup>-2</sup> s<sup>-1</sup>)

k First-order rate constant for biomass loss ( $h^{-1}$ )

LED Light emission diodes

ND Not determined

 $N_f$  Final concentration of nitrate (mg L<sup>-1</sup>)

 $N_i$  Initial concentration of nitrate (mg  $L^{-1}$ )

PAR Photosynthetically active radiation

PBR n Tubular photobioreactor run n

 $P_f$  Final concentration of phosphate (mg L<sup>-1</sup>)

 $P_i$  Initial concentration of phosphate (mg L<sup>-1</sup>)

PTFE Poly (tetrafluoroethylene), or Teflon

 $Q_L$  Final lipid productivity (g  $L^{-1}d^{-1}$ )

 $Q_X$  Final biomass productivity (g  $L^{-1}d^{-1}$ )

q<sub>L</sub> Specific lipid production rate (d<sup>-1</sup>)

 $q_N$  Specific nitrate consumption rate (mg g<sup>-1</sup>d<sup>-1</sup>)

 $q_P$  Specific phosphate consumption rate (mg g<sup>-1</sup>d<sup>-1</sup>)

RNA Ribonucleic acid

RTD Resistance temperature detector, a type of temperature sensor

RUBISCO Ribulose-1,5-bisphosphate carboxylase/oxygenase

RuBPCase Ribulose-1,5-bisphosphate carboxylase/oxygenase

SARDI South Australia Research and Development Institute

STR n Stirred tank bioreactor (BioFlo) run n

TL Total lipids

t Duration of a batch (d)

 $t_f$  Time at the end of a batch (d)

 $t_i$  Time at the beginning of a batch (d)

V Working volume of the reactor, or volume (L)

 $X_f$  Final concentration of biomass (g L<sup>-1</sup>)

 $X_i$  Initial concentration of biomass (g L<sup>-1</sup>)

 $Y_{L/Light}$  Lipid yield on light (g  $\mu$ mol<sup>-1</sup>)

 $Y_{\rm L/N}$  Lipid yield coefficients on nitrate (g mg<sup>-1</sup>)

 $Y_{L/P}$  Lipid yield coefficients on phosphate (g mg<sup>-1</sup>)

 $Y_{X/Light}$  Biomass yield coefficient on light (g  $\mu$ mol<sup>-1</sup>)

 $Y_{X/N}$  Biomass yield coefficient on nitrate (g mg<sup>-1</sup>)

 $Y_{X/P}$  Biomass yield coefficient on phosphate (g mg<sup>-1</sup>)

y Weight fraction of lipids in the biomass

ICP-MS Inductively coupled plasma mass spectrometry

ICP-OES Inductively coupled plasma optical emission spectrometry