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# Functional protein display on the surface of biobeads produced by recombinant *Escherichia coli*

A thesis presented in partial fulfilment of the requirements for the degree of

Master of Science

In

Biochemistry
at Massey University, Palmerston North,
New Zealand

Shuxiong Chen

#### Appendix D

#### MASSEY UNIVERSITY

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#### **Abstract**

Polyhydroxyalkanoic acids (PHAs) are biopolyesters produced by various bacteria. They are deposited as spherical water-insoluble cytoplasmic inclusions (beads) containing an amorphous hydrophobic polyester core and surrounded by a phospholipid monolayer and embedded proteins, including PHA synthase (PhaC), the key enzyme required for PHA bead formation. Although inactive PhaC cannot produce PHA beads, fusing inactive PhaC to green fluorescent protein (GFP) leads to GFP protein bead formation. Both PHA and protein beads could serve as a versatile platform for display of desired proteins suitable for various biotechnological and medical applications.

The tuberculin skin test (TST) for diagnosing bovine tuberculosis (TB) in cattle uses the purified protein derivative (PPD) that is prepared from *Mycobacterium bovis*. However, some antigens in the PPD are also present in environmental mycobacteria. Therefore, the TST lacks specificity if animals are exposed to non-pathogenic environmental mycobacteria. In this study, three specific TB antigens, CFP10, ESAT6, and Rv3615c—which are present in pathogenic but absent in most non-pathogenic mycobacteria—were displayed on the surface of PHA beads. The results demonstrated that these triple antigen-displaying PHA beads can differentiate TB-infected from non-infected cattle, making this an attractive alternative to current skin test diagnostic reagents.

IgG binding domains displayed on GFP protein beads have a higher IgG binding ability when compared to their counterpart displayed on PHA beads. However, it is unclear whether an enhancement of IgG binding ability due to GFP protein beads could be achieved by immobilization on other fluorescent protein (FP) beads. The results showed that other FP (including yellow, red and cyan) beads displaying IgG binding domains have an approximately 1.5–2 fold greater IgG binding ability when compared to PHA beads displaying the same binding domains. To investigate whether protein beads displaying iron-binding peptides could be magnetized while maintaining IgG binding function, an iron binding peptide was displayed. The results demonstrated that protein beads displaying both IgG and iron binding peptides can be magnetised by iron oxide and retain a strong IgG binding ability. Finally, this study revealed that different cell disruption techniques could affect the morphology and functionality of FP protein bead.

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#### List of abbreviations

A full list of abbreviations used

°C Degree Celsius

3xFe Triple iron-binding peptides
AGE Agarose gel electrophoresis

Ap Ampicillin

APS Ammonium persulfate

BL21/ E. coli production strain

BL21 69/ E. coli production strain harbours plasmid

pMCS69, encoding PhaA and PhaB enzymes

BSA Bovine serum albumin

BCG vaccine strain Live-attenuated Mycobacterium bovis Bacile

Calmette-Guerin vaccine strain

CFP10 The 10-kDa culture filtrate protein

CFP10-PhaC The 10-kDa culture filtrate protein–PHA synthase CFP10-Rv3615c-PhaC-ESAT6 The 10-kDa culture filtrate protein-Rv3615c-PHA

synthase-the 6-kDa early secretory antigenic

target

Cm Chloramphenicol
DMSO Dimethylsulfoxide

DNA Deoxyribonucleic acid

DNAase Deoxyribonuclease

dNTPs Deoxyribonucleotide triphosphates

DTH Delayed-type hypersensitivity

ECFP Enhanced cyan fluorescent protein

ECFP-PhaC(C319A)-[GB1]3 Enhanced cyan fluorescent protein-inactive PHA

synthase-triple IgG binding domain of protein G

ELISA Enzyme-linked immunosorbent assay

EtOH Ethanol

ESAT6 The 6-kDa early secretory antigenic target

ESAT6-PhaC The 6-kDa early secretory antigenic target–PHA

synthase

EYFP Enhanced yellow fluorescent protein

EYFP-PhaC(C319A)-[GB1]3 Enhanced yellow fluorescent protein-inactive

PHA synthase-triple IgG binding domain of

protein G

FPs Fluorescent proteins

FP beads Fluorescent protein beads

FP-PhaC(C319A) Fluorescent protein-inactive PHA synthase

g Gravity/gram

[GB1]3 Triple IgG binding domains of protein G

GFP Green fluorescent protein

GFP beads Green fluorescent protein beads

GFP-PhaC(C319A) Green fluorescent protein-inactive PHA synthase

HcR Far red protein HcRed

HcR-PhaC(C319A)-[GB1]3 Far red protein HcRed-inactive PHA

synthase-triple IgG binding domains of protein G

HRP Horseradish peroxidase

IFNγ Interferon-γ

IgG Immunoglobulin G

IPTG Isopropyl â-D-1-thiogalactopyranoside

kDa Kilo Daltons

λ-DNA Phage lambda DNA

LacZ β-galactosidase

LB Luria-Bertani (broth)

MalE Maltose binding domain

Maldi-TOF/MS Matrix-assisted laser desorption ionization

time-of-flight mass spectrometry

OD Optical density

OpdA Organophosphohydrolases
PCR Polymerase chain reaction

PhaA β-ketothiolase

PHAs Polyhydroxyalkanoic acids
PHA<sub>SCL</sub> Short chain length PHAs
PHA<sub>MCL</sub> Medium chain length PHAs

PhaB Acetoacetyl-CoA reductase

PhaC PHA synthase

PhaC-[GB1]3 PHA synthase-triple IgG binding domains of

protein G

PhaC(C319A) Inactive PHA synthase

PhaE Type III PHA synthase subunit

PhaP Phasin

PhaR Phasin regulatory protein

PHB Poly(3-hydroxybutyric acid)

PPDs Purified protein derivatives

PPD-A Avian PPD
PPD-B Bovine PPD

REs Restriction endonucleases
Rv3615c-PhaC Rv3615c-PHA synthase

SD Standard deviation

SEM Scanning Electron Microscopy

SDS-PAGE Sodium dodecyl sulphate polyacrylamide gel

electrophoresis

TB Tuberculosis

TBE Tris-Borate-EDTA buffer

TEM Transmission and Electron Microscopy

TEMED Tetramethylethylenediamine

Tet Tetracycline

TST Tuberculin skin test

Tris Trishydroxymethylaminomethane

v/v Volume per volume w/v Weight per volume

x AVTS tetrapeptide extension

X-Gal 5-bromo-4-chloro-3-indolyl-beta-D-galactopyrano

side

xECFP-PhaC(C319A)-[GB1]3 AVTS tetrapeptide extension-enhanced cyan

fluorescent protein-inactive PHA synthase-triple

IgG binding domain of protein G

AVTS tetrapeptide extension-enhanced yellow xEYFP-PhaC(C319A)-[GB1]3 fluorescent protein-inactive PHA synthase-triple IgG binding domain of protein G xGFP-PhaC(C319A)-MalE AVTS tetrapeptide extension-green fluorescent protein-inactive PHA synthase-maltose binding domain AVTS tetrapeptide extension-green fluorescent xGFP-PhaC-MalE protein- PHA synthase-maltose binding domain xGFP-PhaC(C319A)-ZZ AVTS tetrapeptide extension-green fluorescent protein-inactive PHA synthase-double IgG binding domains of protein A xGFP-3xFe-PhaC(C319A)-ZZ AVTS tetrapeptide extension-green fluorescent protein-triple iron-binding peptides-inactive PHA synthase-double IgG binding domains of protein Α xGFP-PhaC(C319A)-ZZ-3xFe AVTS tetrapeptide extension-green fluorescent protein-inactive PHA synthase-double binding domains of protein A-triple iron-binding peptides xHcR-PhaC(C319A)-[GB1]3 AVTS tetrapeptide extension-far red protein HcRed-inactive PHA synthase-triple repeats of IgG binding domain of protein G XL1-Blue E. coli cloning strain 77 Double IgG binding domain of protein A ZZPhaC Double IgG binding domain of protein A-active

PHA synthase

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