Copyright is owned by the Author of the thesis. Permission is given for a copy to be downloaded by an individual for the purpose of research and private study only. The thesis may not be reproduced elsewhere without the permission of the Author.

Coordinated transcriptional regulation between a reactive oxygen species-responsive gene network and the circadian clock in *Arabidopsis thaliana*

A thesis presented in partial fulfillment of the requirements for the degree of

Doctor of Philosophy

in

Plant Biology

at Massey University, Palmerston North,

New Zealand.

Alvina Grace Lai

2012

Abstract

Most organisms have evolved endogenous biological clocks as internal timekeepers to fine-tune physiological processes to the external environment. Energetic cycles such as photosynthesis and glycolytic cycles are physiological processes that have been shown to be under clock control. This work sought to understand the mechanism of the synchrony between the circadian oscillator and products of energetic cycles. The fact that plants rely on photosynthesis for survival, and that photosynthesis relies on the sun, this would have meant that oxygen levels would have fluctuated across the day. A common by-product of oxygen metabolism and photosynthesis is the Reactive Oxygen Species (ROS). Evidence has proposed ROS as regulators of cellular signaling and plant development. However, if ROS levels are left unmanaged, it may cause oxidative stress in organisms, which could damage cellular components and disrupt normal mechanisms of cellular signaling. Therefore, it is advantageous for plants to be able to anticipate such periodic burst in ROS. My research investigates the role of the circadian clock in regulating ROS homeostasis in the model plant Arabidopsis thaliana. I found that ROS production and scavenging wax and wane in a periodic manner under diurnal and circadian conditions. Not only that, at the transcriptional level, ROSresponsive genes exhibited time-of-day specific phases under diurnal and circadian conditions, suggesting the role of the circadian clock in ROS signaling. Mutations in the core-clock regulator, CIRCADIAN CLOCK ASSOCIATED 1 (CCA1), affect both the transcriptional regulation of ROS genes and ROS homeostasis. Furthermore, misexpressions of other clock genes such as EARLY FLOWERING 3 (ELF3), LUX ARRHYTHMO (LUX) and TIMING OF CAB EXPRESSION 1 (TOC1) also have profound effects on ROS signaling and homeostasis, thus suggesting a global clock effect on ROS networks. Taken together, CCA1 is proposed as a master regulator of ROS signaling where the response to oxidative stress is dependent on the time of CCA1 expression. Plants exhibit the strongest response at dawn, the time when CCA1 peaks. Moreover, CCA1 can associate to the Evening Element or CCA1-Binding Site on promoters of ROS genes in vivo to coordinate transcription. A common feature of circadian clocks is the presence of multiple interlocked transcriptional feedback loops. It is shown here that the oscillator incorporates ROS as a component of the loop where ROS signals could feed back to affect circadian behavior by changing *CCA1* and *TOC1* transcription. The clock regulates a plethora of output pathways; particularly the transcription of an output gene *FLAVIN BINDING KELCH REPEAT F-BOX 1* (*FKF1*) is affected by ROS signals. Temporal coordination of ROS signaling by *CCA1* and the reciprocal control of circadian behavior by ROS revealed a mechanistic link of which plants match their physiology to the environment to confer fitness.

Preface

Additional published work undertaken during the course of study, as stated below, is included in section 7. Publication.

Lai, A. G., Denton-Giles, M., Mueller-Roeber, B., Schippers, J. H. M., & Dijkwel, P. P. 2011. Positional information resolves structural variations and uncovers an evolutionarily divergent genetic locus in accessions of *Arabidopsis thaliana*. *Genome Biology and Evolution* **3**: 627–640.

Authors' contributions:

AGL carried out sample preparations, LR-PCR, sequence analysis, data interpretation and validation experiments. MD-G participated in sequence analysis. JHMS and BM-R participated in research design and data interpretation. AGL and PPD conceived and designed the project. AGL drafted the manuscript. PPD critically reviewed the manuscript. All authors contributed to the draft, read and approved the final manuscript.

Table of contents

Abstract	ii
Preface	iv
Acknowledgements	v
Abbreviations	ix
List of figures	xv
List of tables	xvii
1. Introduction	1
1.1 The clock paradigm	2
1.1.1 Biological clocks and circadian rhythms	2
1.1.2 Circadian gating of environmental responses	7
1.1.3 Growing with time	10
1.2 The clock's architecture	11
1.2.1 The <i>Arabidopsis</i> oscillator	11
1.2.2 Myb transcription factors and a pseudo-response regulator form	the core
loop	12
1.2.3 PSEUDO-RESPONSE REGULATORS (PRR) 7, PRR9 and PRR5 to	ake the
morning shift	13
1.2.4 GIGANTEA and three F-box proteins take the evening shift	14
1.2.5 Additional indispensable clock regulators	15
1.3 The oxygen paradigm	17
1.3.1 Reactive oxygen species (ROS) and the oxygen paradox	17
1.3.2 Enzymatic and non-enzymatic detoxification of ROS	19
1.3.3 Non-toxic levels of ROS is essential for plant development	20
1.3.4 The dynamics of ROS signaling network	21
1.3.5 ROS as signals for transcriptional coordination	23

	1.4 Circadian regulation of other stress-responsive pathways and the ROS netwo	rk?
		25
	1.5 Aims of this research	27
2	. Materials and Methods	29
	2.1 Plant materials and growth conditions	30
	2.2 H ₂ O ₂ and catalase assays	30
	2.3 ROS hypersensitivity assay	34
	2.4 ROS treatments for qPCR analysis	34
	2.5 Transcript analysis by qPCR	34
	2.6 ChIP-qPCR assay	38
	2.7 Luminescence assay	42
	2.7.1 Seed surface sterilization	42
	2.7.2 Video-intensified microscopy (VIM) imaging of luminescence	42
	2.7.3 Luminescence data processing	45
	2.7.4 Data analysis with BioDare	45
	2.8 Bioinformatics analyses	50
	2.9 Statistical analysis	50
3	. Results	51
	3.1 ROS homeostasis and signaling are regulated by diurnal cycles	52
	3.2 The circadian clock regulates ROS homeostasis and signaling	56
	3.3 A functional clock is required for ROS homeostasis	59
	3.4 The Evening Element is enriched in ROS-responsive genes	68
	3.5 ROS-responsive genes display time-of-day specific expression phase	in
	anticipation of oxidative stress under regular growth condition	76
	3.6 CCA1 regulates plants' response to oxidative stress	84
	3.7 WRKY11, MYB59, PAL1 and ZAT12 are direct targets of CCA1 in vivo	89
	3.8 ROS signals feed back to affect circadian behavior	92

4.	Discussion97
	4.1 Coupling of the circadian clock and metabolism
	4.2 The circadian clock communicates temporal information to regulate ROS
	network transcriptomes
	4.3 The circadian clock mediates the time-of-day sensitivity to oxidative stress
	signals
	4.4 Biological importance of non-photic influences on the circadian oscillator 103
	4.5 Future research
	4.6 Concluding remarks
5.	Appendices
	5.1 Genes of the <i>Arabidopsis</i> circadian clock their proposed molecular function 109
	5.2 Primer sequences of the 167 ROS-responsive genes used in time-course
	expression studies
	5.3 Primer sequences of ROS-responsive genes selected for expression study in clock
	mutants
	5.4 Primer sequences of ROS transcription factors and regulatory genes used for
	expression analysis in response to MV treatments
	5.5 Primer sequences of ROS genes used for ChIP-qPCR assay
	5.6 Primer sequences of reference genes used for qPCR normalization control 117
	5.7 Standard curves
	5.8 The 167 ROS-responsive genes used in expression profiling
	5.9 Diurnal and circadian experimental conditions
6.	Bibliography135
7.	Publication159
	Overview
	laietal 2011

Abbreviations

3-AT 3-aminotriazole

ABA Abscisic acid

ACT2 ACTIN 2

AGI Arabidopsis Gene Identifier

AOX Alternative oxidase

APX Ascorbate peroxidase

APX4 ASCORBATE PEROXIDASE 4

AsA Ascorbate

ASN1 ASPARAGINE SYNTHASE 1

AtCP1 Ca2+ BINDING PROTEIN 1

ATPase Adenosine triphosphatase

bHLH128 Basic helix-loop-helix DNA-binding superfamily

BMAL1 Brain and muscle ARNT-like 1

BME β-mercaptoethanol

BOA BROTHER OF LUX ARRHYTHMO

°C Degrees celcius

CAB2 CHLOROPHYLL A/B BINDING PROTEIN 2

cADPR Cyclic adenosine diphosphate ribose

CAT CATALASE

CBF CRT/DRE BINDING FACTORS

CBS CCA1-binding site

CCA1 CIRCADIAN CLOCK ASSOCIATED 1

CCD Charged couple device

CCR2 COLD CIRCADIAN RHYTHM RNA-BINDING 2

CCT CONSTANS (CO), CO-like, TOC1

cDNA Complementary DNA

CHE CCA1 HIKING EXPEDITION

ChIP Chromatin immunoprecipitation

CLOCK Circadian locomotor output cycles kaput

cm Centimeter

CRY CRYPTOCHROME

CSD2 Cd/Zn superoxide dismutase

Cytb6f Cytochrome b6f

DAB 3,3-diaminobenzidine

DD Constant darkness

DHA Dehydroascorbate

DHAR DHA reductase

DMSO Dimethyl sulfoxide

DNA Deoxyribose nucleic acid

DNase Deoxyribonuclease

DPI Diphenylene iodonium

EC Evening complex

EE Evening element

ELF EARLY FLOWERING

ERF2 ETHYLENE RESPONSE FACTOR 2

Fd Ferredoxin

FeSOD Fe superoxide dismutases

FFT-NLLS Fast fourier transform-nonlinear least squares

FIO1 FIONA 1

FKF1 FLAVIN BINDING KELCH REPEAT F-BOX 1

FNR Ferredoxin NADPH reductase

FW Fresh weight

g Gram

GA Gibberellin

GCL Glutamate cysteine ligase

GAST1 GIBBERELLIC ACID STIMULATED TRANSCRIPT1

GDH1 GLUTAMINE DEHYDROGENASE

GFP Green fluorescent protein

GI GIGANTEA

GLN 1.3 GLUTAMINE SUNTHASE

GLR Glutaredoxin

Glu Glutamate

GO Gene ontology

GPX Glutathione peroxidase

GR Glutathione reductase

GSH Glutathione

GSSG Oxidized glutathione

h Hour(s) H₂O Water

H₂O₂ Hydrogen peroxide

HRP Horseradish peroxidase

HSFA4A HEAT SHOCK TRANSCRIPTION FACTOR A4A

IPP2 ISOPENTENYL PYROPHOSPHATE 2

JMJD5 JUMONJI DOMAIN PROTEIN 5

KI Potassium iodide

krpm Kilo-revolutions per minute

LD 12 h light 12 h dark

LHCB LIGHT-HARVESTING CHLOROPHYLL A/B-BINDING PROTEIN

LHY LATE ELONGATED HYPOCOTYL

LKP2 LOV KELCH PROTEIN 2

LL Constant light

LUC Luciferase

LUX LUX ARRHYTHMO

LWD LIGHT-REGULATED WD 1

M Molar

MAPK Mitogen-activated protein kinase

MDA Monodehydroascorbate

MDAR Monodehydroascorbate reductase

MES18 METHYL ESTERASE 18

μΕ Micro-Einstein

μg Micro-gram

μl Micro-litre

μM Micro-molar

mg Milli-gram

mL Milli-liter

min Minute(s)

mM Milli-molar

mRNA Messenger ribonucleic acid

MS Murashige and Skoog

MV Methyl viologen

MYB59 MYB DOMAIN PROTEIN 59

NAD Nicotinamide adenine dinucleotide

NADP Nicotinamide adenine dinucleotide phosphate

NADPH Nicotinamide adenine dinucleotide phosphate (reduced form)

ng Nano-gram

nm Nano-meter

NOX NADPH oxidase

NTR NADPH thioredoxin reductase

NTRX NADP-linked thioredoxin

¹O₂ Singlet oxygen

O₂ Oxygen

O²⁻ Superoxide anion

OH Hydroxyl radical

PAL1 PHENYLALANINE AMMONIA-LYASE 1

PC Plastocyanin

PCR Polymerase chain reaction

PDX1 PYRIDOXINE BIOSYNTHESIS 1

PER PERIOD

PHY PHYTOCHROME

PIF7 PHYTOCHROME INTERACTING FACTOR 7

PIN PIN-FORMED

PQ Plastoquinone

PRMT5 PROTEIN ARGININE METHYL TRANSFERASE 5

PRR PSEUDO-RESPONSE REGULATOR

PRX Peroxiredoxin

PrxR Peroxireductase

PSI Photosystem I

PSII Photosystem II

PUP1 PURINE PERMEASE 1

qPCR Quantitative polymerase chain reaction

RAV1 RAV family transcription factor

RBOH Respiratory burst oxidase homologue

RBOHC RESPIRATORY BURST OXIDASE HOMOLOG C

RNA Ribonucleic acid

RNase Ribonuclease

ROS Reactive oxygen species

RuBisCO Ribulose-1,5-bisphosphate carboxylase oxygenase

RVE REVEILLE

s.d. Standard deviation

s.e.m Standard error of mean

SA Salicylic acid

SAND GLYCERALDEHYDE-3-PHOSPHATE DEHYDROGENASE C2

SCF Skp1-Cullin-F-box

sec Second(s)

SHAM Salicylhydroxamic acid

SOD Superoxide dismutase

TCA Tricarboxylic acid

TCP TB1, CYC and PCF

TE Tris-ethylene diamine tetra-acetic acid

TF Transcription factor

TIC TIME FOR COFFEE

TOC1 TIMING OF CAB EXPRESSION 1

Trx Thioredoxin

TTFL Transcriptional-translational feedback loop

TUB2 TUBULIN BETA-2

U Unit(s)

UPL7 UBIQUITIN-PROTEIN LIGASE 7

VIM Video-intensified microscopy

VTC2 VITAMIN C 2

WT Wild-type

XCT XAP5 CIRCADIAN TIME-KEEPER

ZAT Zinc-finger transcription factor

ZAT10 SALT TOLERANCE ZINC FINGER

ZT Zeitgeber time

ZTL ZEITLUPE

List of figures

Figure 1.1	An idealized circadian rhythm 4
Figure 1.2	Model of circadian timekeeping 5
Figure 1.3	Circadian gating of environmental responses 9
Figure 1.4	Schematic representation of the <i>Arabidopsis</i> genetic circuit 14
Figure 1.5	Schematic diagram depicting modes of ROS generation and scavenging in plants
Figure 2.1	Real-time imaging of circadian-regulated gene expression with the VIM system
Figure 2.2	Luminescence data analysis with PedroApp and BioDare 49
Figure 3.1	ROS homeostasis and signaling are regulated by diurnal cycles 55
Figure 3.2	The circadian clock regulates ROS homeostasis and signaling 58
Figure 3.3	Mutations in <i>CCA1</i> and <i>LHY</i> resulted in ROS hypersensitivity 61
Figure 3.4	A functional clock is required for ROS homeostasis
Figure 3.5	H ₂ O ₂ and catalase rhythms are regulated by <i>CCA1</i>
Figure 3.6	ROS homeostasis is altered in clock mutants 66
Figure 3.7	Phase enrichments of ROS genes under different ROS-related GO categories
Figure 3.8	CCA1 and LHY are involved in transcriptional coordination of ROS genes expression
Figure 3.9	ELF3, LUX and TOC1 are partly involved in transcriptional coordination of ROS genes expression
Figure 3.10	Clock regulation of non-circadian ROS transcripts
Figure 3.11	Response to ROS is regulated by diurnal cycles and is dependent on the time of <i>CCA1</i> expression
Figure 3.12	Response to ROS is attenuated in <i>CCA1-ox</i>

Figure 3.13	ROS signaling is altered at the transcriptional level in <i>elf3-1, lux-1</i> and
	toc1-1 mutants88
Figure 3.14	CCA1 binds to EE and CBS in promoters of ROS genes in vivo90
Figure 3.15	Not all ROS genes with the CBS are bound by CCA1 in vivo
Figure 3.16	ROS signals feed back to affect circadian behavior94
Figure 3.17	ROS signals feed back into clock-regulated genes
Figure 4.1	Schematic representation of a proposed model depicting the interaction
	between the Arabidopsis circadian clock, ROS networks and clock-
	controlled outputs

List of tables

Table 2.1	Genomic DNA digestion in RNA samples
Table 2.2	Template-primer mixture
Table 2.3	Reverse transcription reaction mixture
Table 2.4	QPCR reaction mixture
Table 2.5	QPCR setup
Table 3.1	GO overrepresentation of the 167 ROS-responsive genes 53
Table 3.2	Enrichments of EE and CBS in ROS-responsive genes obtained from
	publicly available microarray datasets
Table 3.3	Positions of putative EE and CBS in promoters of ROS-responsive genes
	obtained using the ATHENA tool
Table 3.4	Arabidopsis genes under five ROS GO categories that are called
	rhythmic by the DIURNAL tool under at least two diurnal and circadian
	conditions
Table 3.5	GO overrepresentation of genes involved in ROS signaling
Table 3.6	Positions of putative EE and CBS in promoters of selected ROS-
	responsive genes used for expression analysis and ChIP-qPCR 76