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Effects of willow (Salix spp.) browse upon ewe reproduction and rumen microbiology under drought feeding conditions

A thesis presented in partial fulfilment of the requirements for the degree of

Doctor of Philosophy

in

Animal Science

At Massey University, Palmerston North,

New Zealand

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ABSTRACT

A series of grazing experiments were conducted in the summer/autumn of 2003 and 2004 at Massey University's Riverside dryland farm near Masterton in Wairarapa on the East Coast of NZ, to study the effects of grazing willow fodder blocks (6,000 stems/ha) upon the production and reproductive performance of ewes relative to ewes grazing drought pastures. Drought pastures were simulated in this study and included short drought pasture and long drought pasture. Pasture with a low pre-grazing mass of approximately 1500 kg DM/ha, a dead matter content of >50 % and a sward height of 5-7 cm was defined as short drought pasture typical of drought conditions. Long drought pasture was similar to pasture growing in the willow fodder blocks, with a pre-grazing pasture mass of >4000 kg DM/ha, a sward height of > 30cm and a dead matter content of 30-60 %. Willow fodder blocks were established on low-lying wet, marshy areas of the farm that had very low or zero productivity in the undeveloped state. Pasture development in the fodder blocks was noticed with the growth of unsown grasses and legumes, as the areas dried up following the planting of willow stakes, due to evapotranspiration from the trees. Forage in the willow fodder blocks included both trees and pasture that was grown under the trees. The nutritive value of short drought pasture was low with an ME of 8 MJ/kg DM; long drought pasture ranged between 8-10 MJ ME/kg DM; willow pasture contained 8 MJ ME/kg DM in 2003 and 10 MJ ME/kg DM in 2004. The nutritive value of edible willow tree (<5 mm diameter) was superior to drought pasture with an ME of >10 MJ/kg DM. The concentrations of the secondary compounds such as condensed tannins (CT; 30-40 g/kg DM) and phenolic glycosides (PG; 15-35 g/kg DM) were higher in willow

trees compared to their concentrations (CT; 2-3 g/kg DM) and (PG; 2-9 g/kg DM) in control drought pastures.

Experiments involving short drought pasture, long drought pasture and willow fodder blocks as treatment groups were grazed by ewes for 10 weeks in regular breaks from mid February to early May. Ewes were mated during this period and were joined together after mating and grazed on normal pasture until weaning. Live weight (LW) change and body condition score (BCS) were recorded throughout the experiments, whilst reproductive performance of ewes was measured as the number of lambs recorded at ultrasound pregnancy scanning, lambing, docking and weaning. Measurements on wool production were also recorded at weaning.

In 2003, experimental ewes grazed control drought pastures (short and long) and willow fodder blocks (restricted and full access) as treatment groups (n=100 ewes/group; Chapter 2). Ewes grazing short drought pasture had an allowance of 0.8 kg DM/ewe/d whilst ewes with restricted access had an allowance of 0.8 kg DM/ewe/d from drought pasture and 0.4 kg DM/ewe/d from willow fodder blocks. Ewes in full access treatment group had no access to pasture but were confined to willow fodder blocks at an allowance of 2.0 kg DM/ewe/d, which was the same allowance given to long drought pasture ewes. Ewes grazing short drought pasture lost weight at approximately 100g/d and recorded a low reproductive rate (90 lambs weaned/100 ewes mated) with a high proportion of single lamb births. Live weight loss was significantly reduced to 40 g/d in ewes grazing willow fodder blocks (full access) with a 20% units increase in reproductive rate due to more multiple births (P<0.05). Ewes grazing long drought pasture performed intermediate to ewes with full access to fodder blocks and ewes grazing short drought pasture, whilst ewes with

restricted access performed similar to ewes grazing short drought pasture. In 2004 (Chapter 3), the restricted access to willow fodder blocks treatment was eliminated from the study and the number of ewes was increased to 165 ewes per treatment group. Performance of ewes grazing short drought pasture was similar to that of ewes grazing short drought pasture in 2003, with ewes loosing live weight (40g/d) and a low reproductive rate (90 lambs weaned/100 ewes mated) whilst ewes grazing long drought pasture gained LW (54 g/d) and had a higher reproductive rate (P<0.05). Ewes grazing willow fodder blocks performed better than ewes grazing short drought pasture by maintaining LW and their reproductive rate was intermediate to ewes grazing short and long drought pasture.

In 2005, a short grazing trial with rumen fistulated sheep was conducted to study the effect of supplementing willow to ewes grazing drought pastures upon plasma amino acid concentrations (Chapter 4) and upon the microbiology of the rumen (Chapter 5 and 6). Grazing occurred during summer/autumn for 10 weeks with two treatment groups; control (short drought pasture; n=7) at an allowance of 0.8 kg DM/ewe/d and ewes grazing short drought pasture at 0.8 kg DM/ewe/d plus a supplement of fresh willow at 1.4 kg fresh willow/ewe/d (n=7). Blood samples for the quantification of plasma amino acids were collected at week 5 and 10, with L W and BCS measured at fortnightly intervals. Short drought pasture in this experiment had a low pasture mass (2000 kg DM/ha) and a low nutritive value (8 MJ/kg DM), whilst willow had a higher ME of 10 MJ/kg DM. Both groups of ewes lost live weight at the rate of 50 g/d. Plasma concentration of 3 methyl histidine (3-MTH; 88 vs 127 μ mole/L) at week 5 and non essential amino acids (NEAA; 1082 vs 1417 μ mole/L) at week 5 and (1155 vs 1324 μ mole/L) at week 10, were substantially lower (P<0.05) in

willow supplemented ewes than control ewes. It was concluded that the increased reproductive rate from willow supplementation in ewes grazing drought pasture might be partly explained by reduced body protein catabolism, besides also increasing plasma branched chain amino acids (BCAA) and essential amino acids (EAA) concentrations.

To investigate the effects of willow supplementation on rumen microbes, rumen samples were collected during the 2005 experiment with fistulated ewes over a 10 week period. The study involved the use of a molecular technique (Chapter 5), denaturing gradient gel electrophoresis (DGGE), to compare the rumen microbial populations between the control and supplemented ewes and a cultivation technique (Chapter 6) to study the effect on rumen bacteria of ewes grazing drought pastures with and with out willow supplementation. DGGE analysis of the V3 region of 16S ribosomal RNA genes in DNA extracted from samples of rumen contents taken fortnightly over a 10 week feeding period showed a distinct difference in banding patterns between treatment groups which progressively developed over time, showing rumen microbial adaptation to willow supplementation. However, phylogenetic analysis of the DNA sequences retrieved from the DGGE bands from willowsupplemented and control ewes did not cluster by treatment group. It was deduced that willow supplementation induced a change in rumen bacterial populations through selecting sub-populations of organisms already present in the rumen. The changes in the rumen bacterial populations is attributed to the ability of these bacteria to metabolise secondary compounds in willow such as phenolic glycosides and flavanoid monomers and their ability to resist the inhibitory effects of condensed tannins.

The cultivation study involved enumeration, isolation and purification of bacterial colonies on Complete Carbohydrate, Salicin, Xylan, Cellulose and Willow media followed by full characterisation of a representative set of pure bacterial cultures. Total bacterial counts on the above media at week 5 and week 10 were generally lower in willow-supplemented ewes compared to control ewes and the 16S rRNA gene sequences of the majority of isolates characterised from both Salicin and Xylan media, were most closely related to species from the *Pseudobutyrivibrio* genus. Isolates from Willow medium clustered as two distinct groups. One group (mostly isolated from control ewes) was made up of mainly of organisms not usually associated with the rumen and probably represent non-resident organisms that are passing through the rumen. The other group of bacteria were mainly retrieved from willow-supplemented ewes and were most closely related to species of the *Olsenella* genus. Compared to bacteria isolated on Salicin and Xylan media, isolates on Willow medium showed little ability to ferment various carbohydrates or trypticase (hydrolysed protein) but were able to utilise secondary compounds from willow.

It was concluded that willow fodder blocks are useful sources of supplementary fodder for mating ewes during drought situations. Both the field and microbiological studies showed adaptation to the willow supplementary diet, including the detection of *Olsenella*-like bacteria for the first time in the rumen. It is suggested that the principal purpose of the rumen investigation is the degradation of secondary compounds present in willow.

Acknowledgements

I extend my sincere thanks to *Professor Tom Barry* who helped me to step into the highest qualified degree programme and had been a pillar of support and encouragement through out my study. I am indebted for his constant inspiration, untiring help, constructive ideas, immense affection and for the excellent facilities provided to conduct this work. No words can fully express my gratitude for his continued patronage and dedication to this work.

My heartfelt thanks to *Senior Scientist. Dr. Graeme Attwood* for providing me the opportunity to work at AgResearch Rumen Microbiology Laboratory, for his timely help, expertise, valuable suggestions and excellent lab facilities provided to undertake rumen microbiology studies.

I express my thanks to Co-Supervisors to *Dr. Nicolas Lopez Villalobos* and *Prof. Peter Kemp* who encouraged me at every stage and provided constructive ideas and valid criticism when ever required despite their busy schedule.

I am indebted to the technical expertise provided by *Geoff Purchas, Collin Morgan* and *Nathan Crombie* at organising field experiments and to *Dong Li, Matt Deighton, Jason Peters, Matt Nicholson, Debjit De, Graham Naylor, Diana Paccheco* for extending their constant help to carry out my Lab work. Without their help my work would have been incomplete.

My research could not have been possible without the financial support provided by *Massey University* through Doctoral scholarship for 3 years, Joan Berry Scholarship for 6 months and Helen E Akers Scholarship for personal finances. I also thank AgResearch who supported financially in the critical, final stages of this study.

I owe all my friends a big thank you for their constant encouragement and support rendered at various stages of this 4 year study. I especially thank *Ithiel* and *Sheelum Satya* for their limitless help in crucial stages, *Jerusha* for giving me moral strength

and Carolina for backing me up in my low times. For all my other friends with whom

I shared all my feelings during my study and names that are not mentioned here, your

company is always remembered and cherished.

I owe my thanks to my brother Pitta Heston Sam Sagar for his timely help in

supporting me financially to come to New Zealand. Without his help I would not have

been to NZ to pursue my career. I also thank my Parents for their courage, patience

and prayers to send me this far to undertake PhD and backing me every moment. I

also thank my sisters and their families for supporting me to complete this study.

I express my love to Sundar kanth who had been my fiancé through out this study

and gratitude for his encouragement, moral support and patience to help me complete

this degree program. His support remained unchanged as my husband to finish the

concluding phase of this thesis.

Finally, I praise and thank the Lord Almighty Jesus Christ, with whose blessing I am

able to submit this research work.

Praise the Lord.....I finished!!!!!!!!

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LIST OF ABBREVIATIONS

BCAA Branched chain amino acid

BCS Body condition score

CA Chlorogenic acid

CC Complete carbohydrate

CP Crude protein

Cr2O3 Chromium sesquioxide

CT Condensed tannins

d day

DGGE Denaturing gradient gel electrophoresis

DM Dry matter

DMI Dry matter intake

DOMD Digestible organic matter

DON Deoxy Nivalenol

DSMD Days of soil moisture deficit

EAA Essential amino acid

ELISA Enzyme linked immunosorbent assay

ENSO El Nino - Southern oscillation phenomenon

EU European union

FM Flavanoid monomer

FV Feeding value

GDP Gross domestic product
GLM Generalised linear model

ha hectare

HCL Hydrochloric acid

hd Head

HPLC High performance liquid chromatography

HT Hydrolysable tannins

IPO Interdecadal pacific oscillation index

IVDMD Invitro dry matter digestibility

LHV Lower heating value

LIG Lignin

LW Live weight

LWC Live weight change

MAF Ministry of agriculture and forestry

ME Metabolisable energy

MTH Methyl histidine

MW Molecular weight

NAN Non ammonia nitrogen

ND Not determined

NDF Neutral detergent fibre

NE Net energy

NEAA Non-essential amino acid

NH3 Ammonia NIV Nivalenol

NIWA National institute for water and atmospheric research

NV Nutritive valueNZ New ZealandOM Organic matter

OMD Organic matter digestibility

OR Ovulation rate

P Probability

PED Potential evapotranspiration deficit

PEG Poly ethylene glycol
PG Phenolic glycoside

SAS Statistical analysis system

SE Standard error

SMD Soil moisture deficit

SOI Southern oscillation index
TLC Thin layer chromatography
UDP Undegradable dietary protein

VFI Voluntary feed intake