

Copyright is owned by the Author of the thesis. Permission is given for a copy to be downloaded by an individual for the purpose of research and private study only. The thesis may not be reproduced elsewhere without the permission of the Author.

Genetic diversity and gene expression analysis of
Phytophthora pluvialis, a foliar pathogen of conifers

**A thesis presented in partial fulfilment of the
requirements for the degree of**

**Doctor of Philosophy (PhD)
in
Genetics**

**Massey University, Manawatū,
New Zealand.**

Simren Brar

2018

Abstract

Phytophthora pluvialis is the causal agent of red needle cast on *Pinus radiata* in New Zealand. It was first isolated in 2008 but had previously been recovered from tanoak (*Notholithocarpus densiflorus*) and Douglas fir (*Pseudotsuga menziesii*) trees in Oregon, USA in 2002. *Phytophthora pluvialis* was subsequently described as a new species in 2013 and classified as a clade 3 *Phytophthora* species. The aims of this study were to (1) gain a better understanding of the genetic diversity and population structure of *P. pluvialis* and (2) examine gene expression profiles of *P. pluvialis* from naturally infected *P. radiata* seedlings. Studying the genetic diversity and population structure of *P. pluvialis* provided insight into the mode of reproduction of this pathogen and helped determine if *P. pluvialis* was introduced into New Zealand. This information is also important for the development of management strategies for *P. pluvialis*. Twenty-seven single nucleotide polymorphism (SNP) markers were designed to genotype a total of 360 isolates of *P. pluvialis* collected from New Zealand and the USA. The genotypic data showed that the population in New Zealand has lower diversity than the USA population. A minimum spanning network (MSN) showed two unique clusters in the New Zealand population, suggesting there may have been two separate introductions of *P. pluvialis*. For the second study, samples were collected from 45 *P. radiata* grafted plants that were part of a field trial, with the aim of identifying genes that are highly expressed and may be important for virulence. Interestingly, *Phytophthora kernoviae* was found in more of the samples than *P. pluvialis*. Needle samples were collected, RNA was extracted and sequenced, and the normalised reads that mapped to the genome of *P. pluvialis* were compared to those from *P. pluvialis* grown in culture. Differentially expressed genes (DEGs) of *P. pluvialis* that showed higher expression in the field trial included potential orthologs of sugar transporter, GH12 and effector genes with known pathogenicity functions in other species. This is the first study to examine the genetic diversity of *P. pluvialis* in New Zealand and the USA., and to examine the gene expression of a *Phytophthora* forest pathogen in the field. The results from these studies provide useful tools for forest disease management. The SNP markers can be used to monitor the population of *P. pluvialis* in New Zealand. The highly expressed genes can be used to help identify resistance genes in *P. radiata* that can be incorporated into future breeding programs.

Acknowledgements

It has been a privilege to work under the supervision of Dr. Rosie Bradshaw. You truly are the epitome of a research supervisor and scientist. I am extremely appreciative of your support and encouragement throughout this project. I would like to thank my co-supervisors; Dr. Rebecca McDougal, Dr. Nari Williams at Scion. Their passion for *Phytophthora* research was contagious and their research expertise was invaluable for this project. I would like to thank Scion and then Healthy Trees Healthy Future program for funding this project (MBIE C04X1305).

I was lucky to have worked with a great lab group at Massey. Pierre this project would not have been possible without you. Thank you for being so patient with me, I really learned a lot from you. I would like to thank Pranav for all his help with literally anything and everything and my bestie Melissa for keeping me entertained and Lukas my PhD partner.

I would like to thank all of the collaborators who brought a wealth of knowledge to this project. At Oregon State University; Dr. Nik Grunwald, Dr. Everett Hansen, Wendy Sutton, Paul Reeser, Dr. Jared LeBoldus, and a very special thank you to Dr. Javier Tabima. My collaborators at The University of British Columbia; Dr. Richard Hamelin and Dr. Nicolas Feau.

I am blessed to have supportive family and friends who have encouraged me to pursue my passion. Papa ji and Abbotsford Papa ji, you both are my inspiration. You taught me the value of an education and encouraged me to find and pursue my passion to the fullest of my abilities. Mum, Dad and Rajen, you have always supported my dreams and celebrated in my accomplishments. This whole experience has been so special because I was able to share it with all of you. Aman thank you for being so supportive and full of positive energy and encouragement. Dadi Masi ji I miss your smile everyday. You always encouraged me to see the world and enjoy life. Beji, Big mom and Big dad, I know you miss me but thank you for not being upset when I decided to move to NZ. I have to thank the Bhoajis for doing such a great job on raising me. Rummy mamiji, our 3-hour conversation in your kitchen really influenced my decision to start a PhD. I am lucky to have such incredible friends. Jess you know you are my number one! Stephanie, Julia and Nadia I love that it feels like no time has passed when we see each other, and Sonal my day just is not complete without an update from you. To all my Palmy friends thank you for making my time in NZ so memorable.

For Mum, Dad, and Rajen

Table of contents

Abstract	iii
Acknowledgements	v
Table of contents	ix
List of figures	xii
List of tables	xiii
List of appendices	xiv
Abbreviations	xv
Chapter 1. Introduction	1
1.1 Lifecycle of <i>Phytophthora</i> species.....	2
1.2 Dissemination of <i>Phytophthora</i> species.....	4
1.3 Aerial <i>Phytophthora</i> species	6
1.3.1 <i>Phytophthora ramorum</i>	6
1.3.2 <i>Phytophthora pinifolia</i>	9
1.3.3 <i>Phytophthora pluvialis</i>	10
1.4 Other Clade 3 <i>Phytophthora</i> species.....	11
1.5 <i>Phytophthora</i> species present in New Zealand	14
1.5.1. <i>Phytophthora agathidicida</i>	15
1.5.2. <i>Phytophthora kernoviae</i>	15
1.6 Other pathogens on <i>Pinus radiata</i>	16
1.6.1 Dothistroma needle blight.....	16
1.6.2 Cyclaneusma needle cast	17
1.7 Host: <i>Pinus radiata</i>	18
1.8 Management.....	19
1.8.1 Chemical Control.....	20
1.8.2 Biological Control	21
1.8.3 Breeding for Resistance	22
1.9 Objectives	23
Chapter 2. Population genetics of <i>Phytophthora pluvialis</i>	25
2.1. Introduction.....	25
2.2. Materials and Methods.....	29
2.2.1. <i>Phytophthora pluvialis</i> isolates from the USA.....	29
2.2.2. <i>Phytophthora pluvialis</i> isolates from New Zealand	29

2.2.3. DNA extraction.....	30
2.2.4. Confirmation of species.....	30
2.2.5. Genome annotation of <i>Phytophthora pluvialis</i> NZFS3000.....	33
2.2.6. SNP discovery	34
2.2.7. SNP Data analysis.....	39
2.3. Results.....	40
2.3.1. Identification of isolates	43
2.3.2. Preliminary analysis of genetic diversity.....	43
2.3.3. SNP Validation	44
2.3.4. Analysis of genetic diversity in New Zealand and the USA	45
2.4. Discussion	53
2.5. DRC 16 – Statement of contribution to doctoral thesis containing publications.....	62
Chapter 3. Gene expression of <i>Phytophthora pluvialis</i> in field samples.....	65
3.1 Introduction.....	65
3.2 Materials and Methods.....	70
3.2.1. Field trial and sampling	70
3.2.2. Confirmation of <i>Phytophthora pluvialis</i> in samples.....	73
3.2.3. RNA extraction.....	74
3.2.4. RNA sequencing.....	75
3.2.5. Filtering <i>Phytophthora</i> reads (performed by P. Panda, Scion).....	75
3.2.6. Mapping and gene calling of the <i>Phytophthora pluvialis</i> reads (Massey)	76
3.2.7. Analysis of differentially expressed genes	79
3.2.8. GO and KEGG Pathway analysis and Carbohydrate Active Enzyme (CAZy) enrichment analysis	80
3.2.9 Effector candidates	81
3.2.10. Searching for orthologs of putative pathogenicity genes from other <i>Phytophthora</i> species.....	82
3.3 Results.....	82
3.3.1. Field trial design	82
3.3.2. Confirmation of <i>Phytophthora pluvialis</i> in the field samples	84
3.3.3. RNA sequencing and mapping	86
3.3.4. Sample selection and grouping.....	89
3.3.5. Differentially expressed genes in <i>Phytophthora pluvialis</i>	91

3.3.6. GO and KEGG analysis.....	92
3.3.7. Top 20 most highly expressed DEGs	97
3.3.8. CAzy Analysis.....	108
3.3.9. Effector candidates	112
3.4 Discussion	121
3.4.1 Highly expressed <i>Phytophthora pluvialis</i> genes in naturally infected <i>Pinus radiata</i>	121
3.4.2 Limitations of using field infected samples.....	125
3.4.3 Interaction in the forest between <i>Phytophthora pluvialis</i> and <i>Phytophthora kernoviae</i>	127
3.4.4 Future work.....	130
4.0 Thesis discussion	133
5.0 Conclusions.....	139
6.0 Appendix (attached).....	141
7.0 Literature Cited	143

List of figures

Figure 2.1. A map and MSN of the isolates from the USA.	41
Figure 2.2. A map and MSN of the isolates from New Zealand	42
Figure 2.4. A MSN showing the relationship between <i>P. pluvialis</i> isolates.....	52
Figure 3.1. The layout of <i>P. radiata</i> grafted plants used in the field trial	72
Figure 3.2. A pipeline showing the programs used to analyse the RNA-seq data.....	78
Figure 3.3. A heat map confirming the presence of <i>P. pluvialis</i> in the field trial.....	85
Figure 3.4. The proportions of reads that mapped to the seven <i>Phytophthora</i> species.	88
Figure 3.5. Numbers of differentially expressed genes (DEGs) in groups 1 and 2.	92
Figure 3.6. Numbers of DEGs in GO categories that showed significant enrichment.	94
Figure 3.7. Numbers of DEGs in KEGG pathway categories that showed significant enrichment.....	96
Figure 3.8. Amino acid alignment of predicted sugar transporter <i>P. pluvialis</i> Pp006603 with homologs from other species.	107
Figure 3.9. Numbers of DEGs in CAzy categories that showed significant enrichment...	110
Figure 3.10. Amino acid alignment of predicted GH12 genes <i>P. pluvialis</i> Pp011155 and Pp003315.	111
Figure 3.11. Amino acid alignment of predicted RxLR effector <i>P. pluvialis</i> gene Pp011236.	115

List of tables

Table 2.1. Primers used to amplify the <i>coxI</i> and <i>coxII</i> spacer DNA sequence and the <i>ypt1</i> gene	32
Table 2.2. The primers used to validate the 27 SNPs	36
Table 2.3. Primer and probe sequences for iPlex assays <i>P. pluvialis</i>	37
Table 2.4. Summary of the 27 SNPs used to genotype the <i>P. pluvialis</i> isolates.....	46
Table 2.5. Population diversity measures for <i>P. pluvialis</i> in New Zealand and the USA. ...	48
Table 2.6. Analysis of molecular variance for <i>P. pluvialis</i> populations sampled across geographic regions in the USA and New Zealand.....	50
Table 3.1. <i>P. pluvialis</i> and <i>P. kernoviae</i> specific probes and primers used to confirm the presence of each pathogen.	74
Table 3.2. RNA sequencing reads for each sample and the percentages mapping to <i>P. pluvialis</i> and <i>P. kernoviae</i>	90
Table 3.3. The top 20 most highly up-regulated <i>P. pluvialis</i> genes in <i>in planta</i> compared to in culture.	102
Table 3.4. The top 20 most highly expressed <i>P. pluvialis</i> genes <i>in planta</i>	104
Table 3.5. Putative <i>P. pluvialis</i> orthologues of pathogenicity-related genes identified in other species.....	106
Table 3.6. Predicted RxLR effectors in the <i>P. pluvialis</i> DEG dataset	116
Table 3.7. Predicted CRN effectors in the <i>P. pluvialis</i> DEG dataset.....	118
Table 3.8. Reciprocal blast matches of predicted <i>P. pluvialis</i> RxLR and CRN effectors with effectors from other oomycetes.....	120

List of appendices

Appendix Table 2.1. Location and host information for the 360 <i>P. pluvialis</i> isolates used in this study.	141
Appendix Table 2.2. The 58 primers that were tested that did not reveal a SNP.....	141
Appendix Table 2.3. SNP genotype information of the 360 <i>P. pluvialis</i> isolates used in this study.	141
Appendix Table 3.1. The R program code used in DEGseq to determine the DEGs for the Group 1 (Pp) and Group 2 (Pp and Pk) samples used in this study.....	141
Appendix Table 3.2. FPMK values for all the DEGs for all 45 samples used in this study.	141
Appendix Table 3.3. The total read alignment rate to seven <i>Phytophthora</i> species.	141
Appendix Table 3.4. Information on Group 1 DEGs.	141
Appendix Table 3.5. Information on Group 2 DEGs.	141

Abbreviations

AFLP	Amplified fragment length polymorphism
AMOVA	Analysis of molecular variance
AA	Auxiliary activity
bp	Base pair
CBM	Carbohydrate binding module
CE	Carbohydrate esterase
CAzy	Carbohydrate-active enzymes
CRN	Crinkler effectors
DEG	Differentially expressed genes
He	Expected heterozygosity
FPKM	Fragments per kilobase per million reads
GO	Gene ontology
GBS	Genotype by sequencing
GH	Glycoside hydrolase
GT	Glycosyl transferase
HTHF	Healthy Trees Healthy Future MBIE research programme
Ha	Hectare
IGV	Integrated Genomics Viewer
ITS	Internal transcribed spacer
IBD	Isolation by distance
KAAS	KEGG automatic annotations
KEGG	Kyoto Encyclopedia of Genes and Genomes
µl	Microlitre
µM	Micromolar
ml	Millilitre
MSN	Minimum spanning network
min	Minutes
MLG	Multilocus genotype
Ho	Observed heterozygosity
PAMP	Pathogen-associated molecular pattern
PNB	Physiological needle blight
PTA	<i>Phytophthora agathidicida</i> / <i>Phytophthora</i> taxon agathis
PGPR	plant growth-promoting rhizobacteria
PL	Polysaccharide lyase
PCA	Principal component analysis
RNC	Red needle cast
RIN	RNA integrity number
RNA-Seq	RNA sequencing
SSR	Simple sequence repeats (microsatellite markers)
SNP	Single nucleotide polymorphism marker

