Copyright is owned by the Author of the thesis. Permission is given for a copy to be downloaded by an individual for the purpose of research and private study only. The thesis may not be reproduced elsewhere without the permission of the Author.

MALIGNANT HYPERTHERMIA

ALLELE SPECIFIC EXPRESSION AND MUTATION SCREENING OF THE RYANODINE RECEPTOR 1

A dissertation presented to Massey University in partial fulfilment of the requirements for the degree of Doctor of Philosophy in Biochemistry

Hilbert Grievink 2009 To be conscious that you are ignorant is a great step to knowledge

Benjamin Disraeli (1804-1881)

ACKNOWLEDGEMENTS

I would like to take this opportunity to first and foremost thank both my supervisors Assoc. Prof Max Scott and Assoc. Prof. Kathryn Stowell for their expertise, guidance and support during the last three years.

Kathryn, I will be forever grateful for the opportunities you have offered me and helped me realize. I sincerely thank you for presenting me with the possibility to do a PhD after my internship in your research group. You have always been enthusiastic, inspiring and supportive in your advice and guidance. Writing my thesis made me not only realize how much I have learned during these three years but also made me thankful for how much I enjoyed them.

Furthermore I would like to thank everybody in the Twilite Zone for the great times, elaborate pot-luck dinners and awesome lab outings. Will, thanks for all those great games of squash(ing). And Robyn, thank you for an awesome trip around Europe, the good times in the lab, and the much needed distraction (by Robert's emails) from the seemingly endless thesis writing.

I would also like to acknowledge Elaine Langton (Wellington Hospital) and Neil Pollock (Palmerston North Hospital) for supplying blood samples for genomic DNA extractions used for HRM. Thanks also goes to, Jo-Anne Stanton and Chris Mason from the Department of Anatomy and Structural Biology at University of Otago, for their contribution to the 454 data analysis. In addition, I am most grateful to Anthony Thrush and Paula Shields from Roche Diagnostics New Zealand, for their constant technical support over the last three years. Finally, this work would not have been possible without the financial support from the Royal Society of New Zealand Marsden Fund given to Assoc. Prof. Kathryn Stowell.

I am also greatly indebted to my family who have always shown me, love, support and understanding even when I decided to move to the other side of the world. I am looking

forward to being a bit closer to home again. Last but not least I would especially like to thank my wife Liat. Thank you for all your understanding, patience and encouragement over the last three years, all while undertaking your own PhD. I am really sorry for all those times I bored you with my far too detailed monologs of how cool or frustrating my results were. I wish I could promise you it won't happen again.

ABSTRACT

Malignant hyperthermia (MH) is a dominant skeletal muscle disorder caused by mutations in the ryanodine receptor skeletal muscle calcium release channel (RyR1). Allele-specific differences in RyR1 expression levels might provide insight into the observed incomplete penetrance and variations in MH phenotypes between individuals.

Firstly, an H4833Y allele-specific PCR (AS-PCR) assay was designed that allowed for the relative quantification of the two *RYR*1 mRNA alleles in heterozygous samples. In four MHS skeletal muscle samples and two lymphoblastoid cell lines (LCLs), the wild type allele was found to be expressed at higher levels than the mutant RyR1 allele. These differences were not caused by variations in *RYR*1 mRNA stabilities. Secondly, high-throughput amplicon sequencing was employed for the quantification of both the T4826I and H4833Y causative MH mutations in heterozygous MHS samples. With the exception of one, all detected H4833Y and T4826I mutation frequencies were about 50%. This included a control, which was constructed and proven to have a 3:1 ratio of the wild type (H4833) versus the mutant (Y4833) *RYR*1 allele. This suggested that that the high-throughput amplicon sequencing approach as used here, was not suitable for accurate quantification of the two RyR1 alleles in heterozygous H4833Y MHS samples.

To detect possible variations in RyR1 alleles at the protein level, the RyR1 was to be isolated from microsomes prepared from a H4833Y MHS frozen skeletal muscle tissue. Microsomes isolated from MHS skeletal muscle tissues lacked the immunoreactive band that was believed to be the full length RyR1. Poor muscle quality, due to long term storage was believed to be the main cause of RyR1 depletion.

Faster and less expensive screening methodologies are required for the identification of genetic variants in MH research. Thus, in an additional project inexpensive and high-throughput high-resolution melting (HRM) assays were developed to allow screening of the *RYR*1 gene, for mutations associated with MH and/or central core disease (CCD).

ABBREVIATIONS

ACTA1 Skeletal muscle α-actin

apoCaM apo-calmodulin

AS1 Allele-specific primer 1
AS2 Allele-specific primer 2

AS-PCR Allele-specific PCR

ATP Adenosine triphosphate

AVA-CLI Amplicon Variant Analyser Command Line Interface

CaM Calmodulin

CCD Central core disease

cDNA Complementary DNA

CICR Calcium-induced Ca²⁺ release channel

CLI Command Line Interface

CSQ Calsequestrin

Ct values PCR crossing points

CV Coefficient of variance

DEPC Diethylpyrocarbonate

DHPR Dihydropyridine receptor

DMSO Dimethylsulfoxide

dNTP Deoxyribonucleoside triphosphate

dsDNA Double-stranded DNA

DTT Dithiothreitol

E Amplification efficiency

ECCE Excitation-coupled Ca²⁺ entry

EC-coupling Excitation-contraction coupling

EDTA Ethylenediaminetetraacetic acid

eIF4E Eukaryotic initiation factor 4E

EMHG European Malignant Hyperthermia Group

emPCR Emulsion PCR

ER Endoplasmic reticulum

FKBP-12 12 kDa FK506 binding protein

FKBP12.6 12.6 kDa FK506 binding protein

gDNA Genomic DNA

GUI Graphical user interface

HPRT Hypoxanthine-guanine-phosphoribosyltransferase

HRC Histidine-rich calcium binding protein

HRM High-resolution melting

IP₃R Inositol 1,4,5-triphosphate receptors

IPTG Isopropyl-beta-D-thiogalactopyranoside

IRE Iron-responsive element

IRES Internal ribosome entry sites

IVCT In vitro contracture test

JFM Junctional face membrane

KcsA Bacterial K⁺ channel

LCL Lymphoblastoid cell line

m7G 7 methylguanosine

MALDI-TOF Matrix-assisted laser desorption/ionization time-of-flight

MH Malignant Hyperthermia

MHE Malignant hyperthermia equivocal

MHN Malignant hyperthermia negative

MHS Malignant hyperthermia susceptible

MHS1 to 5 Malignant hyperthermia loci 1 to 5

miRNA MicroRNA

MmD Multi minicore disease

mRNA Messenger RNA

MthK *Methanobacterium autotrophicum* potassium channel

MYH7 Beta-myosin heavy chain

NAMHG North American Malignant Hyperthermia Group

NTC Non template control
ORF Open reading frame

PABP Poly(A)-binding protein

PMCA Plasma membrane calcium ATPase

PPi Pyrophosphate

RT-PCR Reverse transcription-PCR

RyR1 Skeletal muscle ryanodine receptor 1 isoform

RyR2 Cardiac muscle ryanodine receptor 2 isoform

RyR3 Brain ryanodine receptor 3 isoform

SEPN1 Selenoprotein gene

SERCA Sarco-Endoplasmic Reticulum Ca²⁺-ATPase

SNP Single nucleotide polymorphism

SOCE Store-operated Ca²⁺ entry

SR Sarcoplasmic reticulum

sstDNA Single strand template DNA

T_m Melting temperature

TRPC Transient receptor potential channel

T-tubule Transverse-tubule

UTR Untranslated region

XALD X-linked adrenoleukodystrophy

X-Gal 5-Bromo-4-Chloro-3-Indolyl-BD-Galactopyranoside

LIST OF FIGURES

Figure 1.1: The mutation hotspots on the <i>RYR</i> 1 gene 20
Figure 1.2: Proposed mechanism for induction of MH caused by abnormalities in the
Ca ²⁺ release channel of skeletal muscle sarcoplasmic reticulum.
Figure 1.3: Possible mechanisms of how RyR1 mutations can affect intracellula
calcium concentrations27
Figure 1.4: Side view of the three dimensional structure of RyR1 30
Figure 1.5: Solid body representations of the three isoforms of ryanodine receptor 31
Figure 1.6: Schematic representation of the major components of the skeletal muscle
triad 35
Figure 2.1: Schematic representation of the QuickChange® Site-Directed Mutagenesis
procedure50
Figure 3.1: Dye redistribution during melting might interfere with heteroduples
detection66
Figure 3.2: Visualizing of the amplified genomic DNA for cloning68
Figure 3.3: EcoR1 digests of 4861 transformants compared to untreated 4861 plasmic
construct69
Figure 3.4: Representative results of the mutagenesis PCR products after Dpn
digestion70
Figure 3.5: HRM data analysis of the 4861 61 bp amplicons, using the LightCycler®
480 Gene Scanning Software
Figure 3.6: Difference plots of gDNA HRM analyses of the 4861 81 bp amplicons 74
Figure 3.7: Difference plots of gDNA HRM analyses of the 4826 77 bp amplicons. $_$ 75
Figure 3.8: Difference plots of gDNA HRM analyses of the 4833 78 bp amplicons. $_$ 76
Figure 3.9: HRM analysis of possible SNP genotypes at the 4861 position, using the
LightCycler® 480 HRM Master (81 bp amplicons) 77
Figure 3.10: HRM analysis of possible SNP genotypes at the 4861 position by adding
wild type DNA amplicons, using the LightCycler® 480 HRM Master (81 bp
amplicons).

Figure 3.11: HRM analysis of possible SNP genotypes at the 4861 position, using the
LightCycler® 480 HRM Master (61 bp amplicons)79
Figure 4.1: Allele specific differences in gene expression84
Figure 4.2: Principle of the AS-PCR of RYR1 cDNA85
Figure 4.3: Visualizing the amplified cDNA fragments used for cloning87
Figure 4.4 Transformants before and after digest with <i>Eco</i> R188
Figure 4.5: Testing allele-specific PCR primer specificities using plasmid constructs. 89
Figure 4.6: Visualizing the AS-PCR products89
Figure 4.7: Real-time PCR results for the determination of reverse transcription
linearity93
Figure 4.8: Determination of the PCR amplification efficiency of mutant RYR1 cDNA.
94 Figure 4.9: Linear relationship of cDNA synthesis as a function of RNA content95
Figure 4.10: Constructed standard curves for the determination of the PCR
amplification efficiencies97
Figure 4.11: Difference plot generated by HRM analysis of the 4833 78 bp amplicons.
104
Figure 4.12: Constructed standard curves for the determination of the PCR amplification efficiencies106
Figure 4.13: mRNA expression levels in LCL #1295 after actinomycin D incubations.
108
Figure 5.1: Clonal amplification of annealed DNA fragments115
Figure 5.2: Bead deposition into PicoTiterPlate116
Figure 5.3: Sequencing-by-synthesis
Figure 5.4: Schematic representation of a PCR product generated by the amplification
using bar coded fusion primers119
Figure 5.5: Visualizing the amplified products used for library preparation124
Figure 5.6: Visualizing the amplified products from serially diluted template used for
library preparation129
Figure 5.7: Visualizing cDNA PCR products generated for allele frequency
determination control129
Figure 6.1: Trypsin cleavage sites around the 4833 amino acid139

Figure 6.2: Separation of microsomal proteins prepared from a frozen MHN skeleta
muscle tissue #1482 143
Figure 6.3: Separation of microsomal proteins prepared from a frozen MHN skeleta
muscle tissue #1482
Figure 6.4: Separation of microsomal proteins prepared from a frozen skeletal muscle
tissues #1482 (MHN) and #145 (MHS)
Figure 6.5: Separation of microsomal proteins prepared from a frozen MHS skeleta
muscle tissues #835 and #116 146
Figure 6.6: Separation of microsomal proteins prepared from a frozen MHN skeleta
muscle tissue #66 147
Figure V.1: Screen shot of the file that contains the Reference sequence in tsv-format
177
Figure V.2: Screen shot of the file that contains the specified Amplicon in tsv-format
178
Figure V.3: Screen shot of the file that contains the specified Variants in tsv-format. 178
Figure V.4: Screen shot of the file that contains the specified Sample in tsv-format. 179
Figure V.5: Screen shot of the file that was used to associate the Amplicons with
Samples in tsv-format 180
Figure V.6: Screen shot of the file that contains the table (in tsv-format) that was used to
upload the ACGA bar coded sequences

LIST OF TABLES

Table 2.1: Reaction components of the standard PCR protocol	46
Table 2.2: Cycle parameters of standard PCR	46
Table 2.3: Reaction components of the pGEM®-T Easy cloning protocol	47
Table 2.4: Reaction components of the <i>Eco</i> RI digestion protocol	49
Table 2.5: Reaction components of the site directed mutagenesis protocol	51
Table 2.6: Site directed mutagenesis cycle parameters	51
Table 2.7: Reaction components of the TURBO DNase treatment protocol	53
Table 2.8: Reaction components for priming RNA with oligo(d)Ts	54
Table 2.9: Reaction components of the oligo(d)T primer extension protocol	54
Table 2.10: Reaction components for the LightCycler® 480 HRM Master	HRM
protocol	55
Table 2.11: Reaction components for the LCGreen PLUS HRM protocol	56
Table 2.12: PCR and HRM parameters	57
Table 2.13: Reaction components for the allele specific PCR protocol	58
Table 2.14: PCR parameters for allele specific PCR	58
Table 3.1: Mutagenic primers for the 4861 amino acid	70
Table 3.2: HRM primer sequences, primer concentrations and amplicon sizes	71
Table 4.1: Primers used in the allele-specific PCR assay	86
Table 4.2: Relative allele frequencies using engineered plasmid constructs	90
Table 4.3: Summary of the linearity of the reverse transcription reactions	96
Table 4.4: Mean PCR amplification efficiencies (n=4)	98
Table 4.5: Intra-assay variability of real-time PCR when screening #470 (n=3)	99
Table 4.6: Inter-assay variability of real-time PCR when screening #470	99
Table 4.7: Relative <i>RYR</i> 1 transcript abundance ratios in muscle tissue (n=3)	101
Table 4.8: Data from the <i>in vitro</i> contracture test	101
Table 4.9: Real-time PCR results for the after prolonged culturing of LCL #1051	102
Table 4.10: Relative RYR1 transcript abundance ratios after prolonged culturing o	f LCL
#1051	103
Table 4.11: Summary of the linearity of the reverse transcription determinations	105

Table 4.12: Mean PCR amplification efficiencies (n=4)	106
Table 4.13: Average real-time PCR results of three independent mRNA stability	y assays
for	107
Table 4.14: Relative RYR1 transcript abundance ratios of LCL #1295 after inc	cubation
with actinomycin D	109
Table 4.15: Relative RYR1 transcript abundance ratios of LCL #1333 after inc	cubation
with actinomycin D	109
Table 5.1: Ten sample DNA library and the bar coded fusion primers used	120
Table 5.2: Seven sample DNA library and the bar coded fusion primers used	121
Table 5.3: Detected SNP frequencies of the ten sample DNA library	126
Table 5.4: Comparison of the detected RyR1 expression levels using high-thr	oughput
sequencing or allele-specific PCR	127
Table 5.5: Relative allele frequencies using purified PCR products	131
Table 5.6: Detected SNP frequencies of the seven sample DNA library	132

TABLE OF CONTENTS

Acknowledgements	iii
Abstract	v
Abbreviations	vi
List of figures	ix
List of tables	xii
1. Introduction	18
1.1 Malignant Hyperthermia	18
1.1.1 History of Malignant Hyperthermia	18
1.1.2 Manifestations of Malignant Hyperthermia	19
1.1.3 Molecular genetics of Malignant Hyperthermia	20
1.1.4 Associated myopathies	21
1.1.5 Physiological basis of malignant hyperthermia	22
1.1.6 Hypersensitivity of MHS RyR1 channels	24
1.1.7 Diagnostic testing	27
1.2 Ryanodine receptor	29
1.3 Maintaining intracellular calcium homeostasis	32
1.3.1 Endogenous RyR1 modulators	32
1.3.2 Proteins involved in SR calcium storage	33
1.3.3 Proteins involved in SR calcium release	34
1.4 The role of gene expression in human disease	38
1.5 PhD project outline	41
2. Materials and methods	43
2.1 Materials	43
2.2 Methods	
2.2.1 Genomic DNA isolation	
2.2.2 Standard PCR protocol	45
2.2.3 Cloning of PCR products	47

2.2.4 Preparation of heat shock competent cells	47
2.2.5 Transformation	48
2.2.6 Rapid boil plasmid preparation	48
2.2.7 Site-directed mutagenesis	49
2.2.8 Isolation of RNA	52
2.2.9 DNase treatment of the isolated RNA	53
2.2.10 First-strand cDNA synthesis	54
2.2.11 High-Resolution Melting reaction conditions	55
2.2.12 Allele-specific PCR reaction conditions	57
2.2.13 Tissue culture	59
2.2.14 Measuring DNA concentrations using the Qubit fluometer	60
2.2.15 Preparation of crude microsomes from skeletal muscle	61
2.2.16 Polyacrylamide gel electrophoresis	62
2.2.17 Western blot analysis	63
2.2.18 DNA sequencing	64
3. High-resolution melting	65
3.1 Introduction	65
3.2 Assay design	67
3.2.1 DNA samples used for assay validation	67
3.2.2 PCR and high resolution melting conditions	71
3.2.3 Data analysis	71
3.3 Results	74
3.3.1 Genomic DNA samples	74
3.3.2 Engineered plasmid constructs	76
3.4 Discussion	79
4. Allele-specific PCR	83
4.1 Introduction	83
4.2 Assay design	85
4.2.1 Primer design	85
4.2.2 Testing allele specificity	86
4.2.3 Validation of allele-specific PCR assay using engineered plasmid constructs _	90
4.2.4 Relative quantification of RyR1 cDNA	91

4	4.3 Results	94
	4.3.1 Screening muscle tissues	94
	4.3.2 mRNA stability assays	_101
4	4.4 Discussion	_109
5.	High-throughput amplicon sequencing	114
į	5.1 Introduction	_114
į	5.2 Assay design	_119
	5.2.1 Bar coded fusion primers	_119
	5.2.2 DNA samples	_121
	5.2.3 PCR conditions and sample preparation	_122
į	5.3 Results	_123
	5.3.1 Ten sample DNA library	_123
	5.3.2 Seven sample DNA library	_128
!	5.4 Discussion	_133
6.	Allelic variation at the protein level	137
(5.1 Introduction	_137
(5.2 Assay design	_138
(5.3 Results	_139
(5.4 Discussion	_147
7.	Summary and future directions	150
•	7.1 SNP detection using HRM	_150
•	7.2 Allele-specific ryanodine receptor 1 expression	_150
	7.2.1 Allele specific RYR1 mRNA expression	_150
	7.2.3 Allele-specific RyR1 expression	_153
Re	ferences	155
Αŗ	ppendices	171
,	Appendix I: PGEM®-T Easy Vector map and sequence reference points	_171
,	Appendix II: Sequence results of the engineered 4861 HRM templates	_172
,	Appendix III: Partial C-terminal amino acid and cDNA sequence of the RYR1	_174
,	Appendix IV: Sequence results of the engineered 4833 plasmid contructs	_175
,	Appendix V: Workflow for using the Amplicon Variant Analyser Command Line Interface	176

Appendix VI: Amplicon Variant Analyser Command Line Interface output files	184
Appendix VII: Results of RyR1 trypsin cleavage	203