

Copyright is owned by the Author of the thesis. Permission is given for a copy to be downloaded by an individual for the purpose of research and private study only. The thesis may not be reproduced elsewhere without the permission of the Author.

STUDIES OF Pasteurella haemolytica:

- (i) COMPARISON OF SEROTYPING TECHNIQUES
- (ii) PREVALENCE OF SEROTYPES IN NEW ZEALAND SHEEP

A THESIS PRESENTED IN PARTIAL FULFILMENT OF  
THE REQUIREMENTS FOR THE DEGREE OF  
MASTER OF SCIENCE IN MICROBIOLOGY AT  
MASSEY UNIVERSITY, NEW ZEALAND

DIANE VALERIE PRINCE

1985

1. (a) I give my permission for my thesis, entitled

STUDIES OF PASTEURELLA HAEMOLYTICA:

(i) COMPARISON OF SEROTYPING TECHNIQUES

(ii) PREVALENCE OF SEROTYPES IN

NEW ZEALAND SHEEP  
to be made available to readers in the Library under the conditions determined by the Librarian.

(b) I agree to my thesis, if asked for by another institution, being sent away on temporary loan under conditions determined by the Librarian.

(c) I also agree that my thesis may be copied for Library use.

2. I do not wish my thesis, entitled

to be made available to readers or to be sent to other institutions without my written consent within the next two years.

Signed *Diane Prince*

Date *15/2/85*

Strike out the sentence or phrase which does not apply.

The Library  
Massey University  
Palmerston North, N.Z.

The copyright of this thesis belongs to the author. Readers must sign their name in the space below to show that they recognise this. They are asked to add their permanent address.

Name and Address

Date

.....  
.....  
.....  
.....  
.....

## ABSTRACT

P.haemolytica, which is the aetiological agent of pasteurellosis, is the major cause of mortalities in sheep in Great Britain and is a problem in several other countries including New Zealand. Furthermore, P.haemolytica, acting as a secondary invader, exacerbates lesions of chronic non-progressive pneumonia (CNP) initiated by Mycoplasma ovipneumoniae and this disease causes considerable economic loss to the New Zealand sheep farming industry.

P.haemolytica exists as 15 serotypes and immunity is serotype specific. P.haemolytica vaccines are marketed overseas and their use in New Zealand is under active consideration. It is logical however, to establish which serotypes are present in New Zealand before a vaccine is produced, but there is at present, no information on this point. This is largely because of technical difficulties in typing isolates. This situation stimulated the present investigation which has 2 major aims: To develop an improved method of typing P.haemolytica and to gather some data on the prevalence of the various serotypes in New Zealand.

With respect to the first aim, several approaches were taken to the problem of differentiating between serotypes.

(1) Indirect heamagglutination (IHA), the standard method by which P.haemolytica is serotyped, was found to be laborious and gave many cross-reactions.

(2) SDS-PAGE of total protein showed similar patterns for all

serotypes within a biotype, whereas the 2 biotypes had different patterns. Thus, SDS-PAGE cannot be used to identify the serotype, but could be useful for identifying the species and the biotype.

(3) Latex beads, coated with antibody prepared against whole cells, agglutinated homologous cells, but also gave many cross-reactions. This test should, in principle, become type-specific if purified capsule were used as the immunising antigen, but further work is required to prepare capsular polysaccharide free from endotoxin which stimulates the production of cross-reacting antibody.

(4) Gel precipitation was simple to perform and was serotype specific. However, serotype A2 required a concentrated antigen preparation for the detection of a precipitation line. The concentrated antigen could not be routinely used with the other types because of cross-reactions between antigens.

(5) Counter immunoelectrophoresis (CIE) was not serotype specific due to endotoxin causing cross-reactions.

(6) Bacterial agglutination was a rapid and simple test to perform and showed some limited cross-reactions.

Since IHA, gel precipitation and agglutination showed some potential, they were then compared using 40 isolates from the lungs of 60 sheep with CNP. These findings reinforced the conclusions drawn when prototype strains were used.

We initially proposed that the serotype of isolates should

be primarily determined by agglutination tests, but since this is not 100% specific, the results must be confirmed by gel precipitation. This approach was investigated using 110 isolates from the nasopharynx of sheep and it was satisfactory for all serotypes except A2. Strains within the A2 serotype showed some heterogeneity at least when examined by gel precipitation and a hypothesis is presented to explain this.

We conclude that at present, the best method of serotyping isolates is by gel precipitation, but that all isolates which are not positively serotyped by this approach should be retested by IHA.

Information concerning the prevalence of serotypes was obtained during the above studies. 110 isolates were obtained from the nasopharynx of 50 sheep from each of 4 farms. All "A" serotypes, except A12 and A14, were isolated. Serotype A2 made up 35% of these isolates. 60 lungs with CNP lesions were obtained from 20 farms in the Manawatu region and 40 isolates of P.haemolytica were obtained. All were serotyped and they were: A1 (25%), A2 (55%), A6 (7.5%), A7 (7.5%), A8 (2.5%), and A9 (2.5%). No "T" types were found in either the nasopharynx or the lungs. The implications of these findings for vaccine production are discussed.

## ACKNOWLEDGMENTS

I would like to thank the Department of Microbiology and Genetics for providing the facilities and opportunity for this study.

In particular, I would like to thank my supervisor, Dr. John K. Clarke, for his intense interest and his advice throughout this work.

Special thanks also go to:

Mr. George Ionas for showing me how to run SDS.PAGE gels,

Dr. M.R. Alley for the collection of nasal swabs from sheep,

ICI-Tasman Ltd., Upper Hutt for 2 reasons:

- i Employing me during summer
- ii The gift of the 15 prototype strains of P.haemolytica which were used in this work.

Furthermore, I would like to thank my family and friends (especially Lyn, Nick and my hockey coach) for their support throughout everything.

## LIST OF CONTENTS

	<u>Page</u>
TITLE PAGE . . . . .	i
ABSTRACT . . . . .	ii
ACKNOWLEDGEMENTS . . . . .	v
LIST OF CONTENTS . . . . .	vi
LIST OF FIGURES . . . . .	ix
LIST OF TABLES . . . . .	xi
INTRODUCTION . . . . .	1
CHAPTER 1: <u>Historical Review</u>	
1.10 Classification of <u>P.haemolytica</u> . . . . .	4
1.11 Biotypes . . . . .	4
1.12 Serological Typing . . . . .	6
1.13 Somatic Antigens . . . . .	7
1.14 Relationship of biotype to serotype . . . . .	7
1.20 Commensal Association in Sheep . . . . .	8
1.30 Diseases Associated with <u>P.haemolytica</u> in Sheep . . . . .	8
1.31 Pneumonic Pasteurellosis . . . . .	9
1.32 Septicaemic Pasteurellosis . . . . .	10
1.33 Importance of Pasteurellosis . . . . .	12
1.34 Chronic Non-Progressive Pneumonia . . . . .	12
1.35 Importance of CNP . . . . .	15
1.40 Immunity to <u>P.haemolytica</u> . . . . .	16
1.50 Vaccine to <u>P.haemolytica</u> . . . . .	20
1.61 Indirect Haemagglutination . . . . .	22
1.62 SDS-PAGE . . . . .	22
1.63 Antibody-Coated Latex Bead Agglutination . . . . .	23
1.64 Gel Precipitation . . . . .	24
1.65 Counter Immunoelectrophoresis . . . . .	25

1.66 Agglutination . . . . .	25
CHAPTER 2 <u>Isolation and Identification of P.haemolytica</u>	
2.1 Introduction . . . . .	27
2.2 Materials and Methods . . . . .	27
2.3 Results . . . . .	32
2.4 Discussion . . . . .	32
CHAPTER 3 <u>Comparison of Isolates by SDS-PAGE</u>	
3.1 Introduction . . . . .	34
3.2 Materials and Methods . . . . .	35
3.3 Results . . . . .	37
3.4 Discussion . . . . .	48
CHAPTER 4 <u>Assessment of Serotyping Techniques and the Production of Antisera Using the Prototype Strains of P.haemolytica</u>	
4.1 Introduction . . . . .	50
4.2 Materials and Methods	
4.21 Antiserum Production to the Capsular Polysaccharide in Different Species of Animal . . . . .	52
4.22 Indirect Haemagglutination . . . . .	53
4.23 Latex Bead Agglutination . . . . .	54
4.24 Gel Precipitation . . . . .	56
4.25 Counter Immunoelectrophoresis . . . . .	57
4.26 Bacterial Agglutination . . . . .	60
CHAPTER 5 <u>Comparison of Techniques using P.haemolytica Isolated from Ovine Lung</u>	
5.1 Introduction . . . . .	85
5.2 Materials and Methods . . . . .	85
5.3 Results . . . . .	86

5.4 Discussion . . . . .	89
CHAPTER 6 <u>Serotyping Nasal Isolates of P.haemolytica</u>	
6.1 Introduction . . . . .	91
6.2 Materials and Methods . . . . .	92
6.3 Results . . . . .	93
6.4 Discussion . . . . .	99
CHAPTER 7 <u>General Discussion</u> . . . . .	102
APPENDIX . . . . .	118
BIBLIOGRAPHY . . . . .	129

## LIST OF FIGURES

<u>Figure</u>	<u>Page</u>
1 Colonies of <u>P.haemolytica</u> serotype A1 .. .. .	31
2 Growth curves for <u>P.haemolytica</u> serotypes A1 or T4 .. ..	39
3 Total proteins of <u>P.haemolytica</u> serotype A1 at various loadings .. .. .	40
4 Comparison of the total proteins of various "A" serotypes	41
5 Comparison of the total proteins of various "A" serotypes and all "T" serotypes .. .. .	42
6 Comparison of the total proteins of various isolates of serotype A1 .. .. .	43
7 Comparison of the total proteins of various isolates of serotype A6 .. .. .	44
8 Comparison of the total proteins of various isolates of serotype A2 .. .. .	a 45 b 46
9 Comparison of the total proteins of 4 isolates of gram negative rods (not <u>P.haemolytica</u> ) with "A" and "T" strains of <u>P.haemolytica</u> .. .. .	47
10 IHA of antisera to <u>P.haemolytica</u> A2 against RBC's sensitised with antigen from each of the 15 serotypes .	63
11 Gel precipitation of T3 antibody against standard and concentrated T3 antigen and also phenol extracted concentrated antigen .. .. .	67
12 Gel precipitation of T3 antibody against phenol extracted T3 antigen which was otherwise untreated or periodate treated .. .. .	68
13 Gel precipitation of T3 antibody against concentrated T3 antigen, untreated and pronase treated .. .. .	69
14 Gel precipitation of T3 antibody against standard T3 antigen and ultracentrifuged T3 antigen .. .. .	70

15	Optimum time required for CIE of <u>P.haemolytica</u> .. .. .	72
16	CIE of various antisera against antigens to the 15 serotypes .. .. .	73
17	Removal of CIE cross-reacting antigens of <u>P.haemolytica</u> A1 by pronase treatment and ultracentrifugation .. .. .	74



nasopharynx from 4 different flocks .. . . . . . 98

XVIII Comparison of prevalence of serotypes from the lungs of  
sheep from the nasopharynx .. . . . . . 99

XIX The 3 groups of A2 isolates obtained from the nasopharynx . . 100