

Copyright is owned by the Author of the thesis. Permission is given for a copy to be downloaded by an individual for the purpose of research and private study only. The thesis may not be reproduced elsewhere without the permission of the Author.

**Gonadal growth and regression in Japanese quail
(*Coturnix coturnix japonica*) and the effect of
gonadotropin-releasing hormone (GnRH) on
luteinising hormone (LH) and ovarian growth**

A thesis presented
in partial fulfilment of the requirements
for the degree of
DOCTOR OF PHILOSOPHY
in Physiology

at Massey University, Palmerston North
New Zealand

SHARON JANE HENARE

2004



Candidate's statement

This is to certify that the research carried out for my doctoral thesis entitled: "Gonadal growth and regression in Japanese quail (*Coturnix coturnix japonica*) and the effect of gonadotropin-releasing hormone (GnRH) on luteinising hormone (LH) and ovarian growth" in the Institute of Veterinary, Animal and Biomedical Sciences, Massey University, Turitea Campus, Palmerston North, New Zealand is my own work and that the thesis material has not been used in part or in whole for any other qualification.

Sharon Jane Henare

September 2004



Supervisor's statement

This is to certify that the research carried out for the doctoral thesis entitled "Gonadal growth and regression in Japanese quail (*Coturnix coturnix japonica*) and the effect of gonadotropin-releasing hormone (GnRH) on luteinising hormone (LH) and ovarian growth" was done by Sharon Jane Henare in the Institute of Veterinary, Animal and Biomedical Sciences, Massey University, Turitea Campus, Palmerston North, New Zealand. The thesis material has not been used in part or in whole for any other qualification, and I confirm that the candidate has pursued the course of study in accordance with the requirements of the Massey University regulations.

Dr John F. Cockrem

24 September, 2004



Certificate of regulatory compliance

This is to certify that the research carried out in the doctoral thesis entitled "Gonadal growth and regression in Japanese quail (*Coturnix coturnix japonica*) and the effect of gonadotropin-releasing hormone (GnRH) on luteinising hormone (LH) and ovarian growth" in the Institute of Veterinary, Animal and Biomedical Sciences, at Massey University, New Zealand:

- (a) is the original work of the candidate, and reference to work other than that of the candidate, has been appropriately acknowledged by appropriate attribution in the text and/or in the acknowledgements;
- (b) that the text does not exceed 100, 000 words;
- (c) research practice, ethical and genetic technology policies have been complied with as appropriate.

Candidate: Miss Sharon Jane Henare

Supervisor: Dr John F. Cockrem

24 September 2004

Abstract

Improvements in breeding success are needed for conservation of endangered birds such as the New Zealand kakapo. A potential method to stimulate breeding is treatment with exogenous hormones. Hormone treatment is used in captive breeding programmes for endangered mammals but reliable techniques are not available for birds. Gonadotropin releasing hormone (GnRH), the principal hormone controlling reproduction, has been used to induce ovarian growth and ovulation in seasonally anoestrous mammals. The goal of the research in this thesis was to determine the potential of GnRH for hormone treatment in birds.

The Japanese quail (*Coturnix coturnix japonica*) was used in the current research. Female quail held outdoors with male quail and with access to nesting materials and nest sites showed clear seasonal patterns in the width of the cloacal opening (indicative of oviduct development) and FSH concentrations, whilst LH concentrations were low during winter and increased during spring and summer. Prolactin concentrations were elevated in birds incubating eggs in nests and birds caring for young. Photoperiodically induced gonadal growth and regression were described in detail for male and female quail under controlled conditions. Testicular and ovarian growth was preceded by increased LH and FSH concentrations and accompanied by increased gonadal steroid concentrations.

Administration of various types of GnRH stimulated luteinising hormone (LH) secretion in sexually regressed female Japanese quail. LH responses to cGnRH-II were greater than those to cGnRH-I. Low doses of buserelin stimulated similar LH responses to cGnRH-II, whilst high doses of buserelin and D-Lys⁶Trp⁷Tyr⁸-GnRH induced sustained LH secretion. Single daily injections of various doses of cGnRH-II, buserelin or D-Lys⁶Trp⁷Tyr⁸-GnRH in saline or polyvinylpyrrolidone (PVP) did not induce elevated baseline LH or stimulate ovarian growth. Repeated injections of D-Lys⁶Trp⁷Tyr⁸-GnRH did not increase LH concentrations over a short-term period. Continuous infusion of D-Lys⁶Trp⁷Tyr⁸-GnRH by osmotic mini-pump severely blunted the LH response and did not stimulate ovarian growth.

Future studies using quail exposed to marginally stimulatory photoperiods will offer the opportunities to determine the effects of GnRH in birds under conditions which mimic photoperiod changes during the breeding season. Further studies on the potential development of a hormone treatment programme will continue to offer a promising future for endangered avian species including the New Zealand kakapo.

Acknowledgements

I would like to acknowledge my supervisor, Dr John Cockrem, for providing support, guidance and the opportunity to attend the 7th meeting of the International Society of Avian Endocrinology during my studies. I would also like to thank Comalco and the Kakapo Recovery Programme, Department of Conservation and the Institute of Veterinary, Animal and Biomedical Sciences (IVABS) Postgraduate Research Fund for funding this research. Personal financial support was provided by the Tuapapa Putaiao Maori Fellowship Scheme, Foundation for Research, Science and Technology (FRST), Maori Education Trust and the Massey University Scholarship Committee.

I would like to extend my gratitude to Professor Robert Millar for the generous donation of the cGnRH-II analogue and acknowledge Dr Mitoshi Kikuchi, Dr Richard Talbot and Professor Masaru Wada for assaying samples. Special thank you to Miss Jane Candy for guidance and assistance with radioimmunoassay work and Dr Greg Anderson and Dr Jamroen Thiengtham for practical advice when establishing the LH assay. I would also like to thank Mr Kevin McGill from NIWA for climate data.

Many thanks to past and present members of the Conservation Endocrinology Research Group; Raewyn Wheeler, Dr Ellen Bennett, Emma Hawke, Guy Hessel, Dr Jane Girling, Dr Kate Littin, Dominic Adams, Heather Hesterman, Marcus Sheehan, Jane Candy, Janis Bridges, and Cathy Davidson for the endless help, good laughs and the great dinner parties! Special thanks to Dr Wei-Hang Chua for your help in the lab, help with the computer, collecting blood samples at 2 in the morning, need I say more! I would also like to thank the past and present staff and students of the Comparative Physiology and Anatomy Group and the staff at the Massey University Library for continual encouragement and interest in my research. Thanks also to Mrs Allain Scott from the IVABS postgraduate research office.

Special thank you to Nick Roskrige and Zirsha Wharemate for being positive and always opening their office doors for a cup of tea and a bickie! I would also like to acknowledge Professor David Mellor for encouraging my initial interest in physiology.

Finally I would like to thank my family for their support and encouragement during the numerous years of my studies, for politely ignoring my piles of data and research all around the house and not complaining when I ate all the biscuits!

Table of Contents

1. General introduction and literature review.....	1
1.1 Photoperiodic control of reproduction in birds.....	1
1.1.1 Seasonality.....	1
1.1.2 Photoperiodism.....	2
1.2 Neuroendocrine regulation of reproduction in birds.....	4
1.2.1 Encephalic photoreceptors and the circadian clock.....	6
1.2.2 GnRH neuronal system.....	6
1.2.2.1 Molecular biology of GnRH.....	8
1.2.2.2 Pattern of cGnRH secretion.....	14
1.2.3 Peripheral endocrine system.....	16
1.2.3.1 Gonadotropins.....	16
1.2.3.2 Gonadal steroid feedback.....	21
1.2.3.3 Gonadotropin inhibitory hormone (GnIH).....	22
1.2.4 Prolactin.....	22
1.2.5 The role of thyroid hormones in seasonal reproduction.....	23
1.3 Photoperiodic responses of Japanese quail.....	25
1.3.1 Gonadal growth on artificial long daylengths.....	26
1.3.2 Gonadal growth on artificial short daylengths.....	27
1.3.3 Gonadal regression in quail transferred from long to short days	28
1.4 Hormonal stimulation using GnRH in birds.....	28
1.4.1 Stimulation of ovarian development in birds using GnRH.....	30
1.4.2 Stimulation of ovulation in birds using GnRH.....	30
1.4.3 Conclusion.....	31
1.5 Outline of thesis.....	31
2. Annual reproduction cycle of female Japanese quail held in a semi-natural environment.....	33
2.1 Introduction.....	33
2.2 Materials and methods.....	35
2.2.1 Animals and housing.....	35
2.2.2 Experimental design.....	36
2.2.3 Radioimmunoassay.....	37
2.2.3.1 Luteinising hormone (LH).....	38
2.2.3.2 Follicle stimulating hormone (FSH).....	38
2.2.3.3 Prolactin.....	39
2.2.3.4 Thyroxine.....	40
2.2.3.5 Triiodothyronine.....	41
2.2.4 Meteorological data.....	41
2.2.5 Statistics.....	44
2.3 Results.....	44
2.3.1 Reproductive activity.....	44
2.3.1.1 Male behaviour.....	44
2.3.1.2 Female behaviour.....	45
2.3.2 Body weight.....	49
2.3.3 Width of cloacal opening.....	53
2.3.4 Luteinising hormone (LH).....	59
2.3.5 Follicle stimulating hormone (FSH).....	61

2.3.6 Prolactin.....	63
2.3.7 Thyroxine (T4).....	67
2.3.8 Triiodothyronine (T3).....	70
2.3.9 Molt.....	70
2.3.10 Summary of results.....	72
2.4 Discussion.....	74
2.4.1 Behaviour.....	74
2.4.2 Endocrinology.....	76
2.5 Conclusion.....	79
3. Photoperiodic control of gonadal growth and gonadal regression in male and female Japanese quail	81
3.1 Introduction.....	81
3.2 Materials and methods.....	84
3.2.1 Animals and housing.....	84
3.2.2 Experimental design.....	85
3.2.2.1 Determination of the pattern and rate of gonadal growth.....	85
3.2.2.2 Determination of the pattern and rate of gonadal regression.....	85
3.2.3 Data collection.....	86
3.2.3.1 Cloacal foam production and egg production.....	86
3.2.3.2 Blood and tissue samples.....	86
3.2.3.3 Rate of gonadal growth and regression.....	87
3.2.4 Radioimmunoassays.....	87
3.2.4.1 Luteinising hormone (LH).....	87
3.2.4.2 Follicle stimulating hormone (FSH).....	88
3.2.4.3 Prolactin.....	89
3.2.4.4 Testosterone.....	90
3.2.4.5 Estradiol.....	91
3.2.4.6 Progesterone.....	92
3.2.4.7 Thyroxine (T4).....	93
3.2.4.8 Triiodothyronine (T3).....	94
3.2.5 Statistics.....	97
3.3 Results.....	98
3.3.1 Gonadal growth and regression in male Japanese quail.....	98
3.3.1.1 Body weight.....	98
3.3.1.2 Cloacal gland area.....	101
3.3.1.3 Cloacal foam production.....	109
3.3.1.4 Paired testis weight.....	111
3.3.1.5 Luteinising hormone (LH).....	118
3.3.1.6 Follicle stimulating hormone.....	122
3.3.1.7 Testosterone.....	127
3.3.1.8 Prolactin.....	133
3.3.1.9 Thyroxine (T4).....	137
3.3.1.10 Triiodothyronine.....	142
3.3.1.11 Summary of results for gonadal growth and regression.....	148
3.3.2 Gonadal growth and regression in female Japanese quail.....	152
3.3.2.1 Body weight.....	152
3.3.2.2 Width of cloacal opening.....	155
3.3.2.3 Ovary weight.....	163

3.3.2.4 Oviduct weight.....	173
3.3.2.5 Luteinising hormone (LH).....	179
3.3.2.6 Follicle stimulating hormone (FSH).....	183
3.3.2.7 Estradiol.....	188
3.3.2.8 Progesterone.....	194
3.3.2.9 Prolactin.....	200
3.3.2.10 Thyroxine (T4).....	204
3.3.2.11 Triiodothyronine (T3).....	209
3.3.2.12 Summary of results for gonadal growth and regression in female Japanese quail.....	215
3.4 Discussion.....	219
3.4.1 Testicular growth in male Japanese quail.....	219
3.4.2 Testicular regression in male Japanese quail.....	224
3.4.3 Ovarian growth in female Japanese quail.....	227
3.4.4 Ovarian regression in female Japanese quail.....	231
3.5 Conclusion.....	231
4. Effect of a single GnRH injection on LH concentrations and effect of daily injections on LH concentrations and ovarian development.....	233
4.1 Introduction.....	233
4.2 Materials and methods.....	234
4.2.1 Animals and housing.....	234
4.2.2 Experimental design.....	234
4.2.2.1 Experiment one.....	235
4.2.2.2 Experiment two.....	235
4.2.2.3 Experiment three.....	236
4.2.3 Hormone preparation and administration.....	237
4.2.3.1 cGnRH-I preparation and administration.....	237
4.2.3.2 cGnRH-II preparation and administration.....	237
4.2.3.3 Buserelin preparation and administration.....	237
4.2.3.4 D-Lys ⁶ Trp ⁷ Tyr ⁸ -GnRH preparation and administration	238
4.2.4 Data collection.....	239
4.2.4.1 Experiments one and two.....	238
4.2.4.2 Experiment three.....	238
4.2.5 Radioimmunoassays.....	238
4.2.5.1 Luteinising hormone.....	239
4.2.6 Statistics.....	241
4.3 Results.....	241
4.3.1 Experiment one: Acute LH response to exogenous GnRH.....	241
4.3.1.1 Acute LH response to a single subcutaneous cGnRH-I injection.....	241
4.3.1.2 Acute LH response to a single subcutaneous cGnRH-II injection.....	247
4.3.1.3 Acute LH response to a single subcutaneous buserelin injection.....	252
4.3.2 Experiment two Acute LH response to exogenous GnRH.....	257
4.3.2.1 Acute LH response to a single subcutaneous D-Lys ⁶ Trp ⁷ Tyr ⁸ -GnRH.....	257
4.3.3 Comparisons between experiments one and two.....	262
4.3.3.1 Comparisons of LH responses to 0.5 µg GnRH/kg.....	262
4.3.3.2 Comparisons of LH responses to 1 µg GnRH/kg.....	264

4.3.3.3	Comparisons of LH responses to 2.5 µg GnRH/kg.....	265
4.3.3.4	Comparisons of LH responses to 5 µg GnRH/kg.....	266
4.3.4	Experiment three.....	267
4.3.4.1	Daily injections of cGnRH-II in saline or PVP.....	271
4.3.4.2	Daily injections of D-Lys ⁶ Trp ⁷ Tyr ⁸ -GnRH in saline or PVP.....	282
4.4	Discussion.....	288
4.4.1	Acute LH response to a single subcutaneous GnRH injection...	288
4.4.2	Effect of a single daily injection of GnRH for seven days.....	290
4.5	Conclusion.....	292
5.	Stimulation of LH secretion by repeated D-Lys⁶Trp⁷Tyr⁸-GnRH injections.....	293
5.1	Introduction.....	293
5.2	Materials and methods.....	294
5.2.1	Animals and housing.....	294
5.2.2	Experimental design.....	295
5.2.3	Hormone preparation and administration.....	296
5.2.4	Data collection.....	296
5.2.5	LH radioimmunoassay.....	296
5.2.6	Statistics.....	296
5.3	Results.....	297
5.3.1	LH response to the first injection of saline, 2.5 or 5 µg D-Lys ⁶ Trp ⁷ Tyr ⁸ -GnRH.....	297
5.3.2	LH response to the second injection of saline, 2.5 or 5 µg D-Lys ⁶ Trp ⁷ Tyr ⁸ -GnRH.....	300
5.3.3	LH response to the third injection of saline, 2.5 or 5 µg D-Lys ⁶ Trp ⁷ Tyr ⁸ -GnRH.....	304
5.3.4	LH response to the fourth injection of saline, 2.5 or 5 µg D-Lys ⁶ Trp ⁷ Tyr ⁸ -GnRH.....	307
5.3.5	LH responses to repeated injections of 2.5 µg D-Lys ⁶ Trp ⁷ Tyr ⁸ -GnRH.....	310
5.3.6	LH responses to repeated injections of 5 µg D-Lys ⁶ Trp ⁷ Tyr ⁸ -GnRH.....	312
5.3.7	LH concentrations at the end of the experiment.....	313
5.4	Discussion.....	315
5.5	Conclusion.....	317
6.	Effect of D-Lys⁶Trp⁷Tyr⁸-GnRH delivered by osmotic pump on LH secretion and ovarian growth.....	318
6.1	Introduction.....	318
6.2	Materials and methods.....	319
6.2.1	Animals and housing.....	319
6.2.2	Experimental design.....	320
6.2.3	Hormone preparation and administration.....	321
6.2.4	Data collection.....	321
6.2.5	LH radioimmunoassay.....	322
6.2.6	Statistics.....	322
6.3	Results.....	323
6.3.1	Width of cloacal opening.....	323
6.3.2	Luteinising hormone.....	326

6.3.3 Ovary weight.....	332
6.3.4 Oviduct weight.....	334
6.4 Discussion.....	336
6.5 Conclusion.....	338
7. General discussion.....	339
8. References.....	347
9 Appendices.....	373
Appendix 1 Annual reproductive cycle of female Japanese quail held in a semi-natural environment.....	373
Appendix 2. Papers in preparation for publication	380

List of Figures

- Figure 1.1** Diagram of the hypothalamic-pituitary-gonadal axis. In response to stimulatory photoperiodic conditions, gonadotropin-releasing hormone-I (GnRH-I) neurons secrete GnRH which stimulates the pituitary gland to synthesise and secrete luteinising hormone (LH) and follicle stimulating hormone (FSH). LH and FSH bind to receptors within the testes or the ovary and stimulate the production of the gonadal steroids – androgens, estrogens, progestins and inhibin. The steroids participate in the regulation of the HPG axis through feedback systems and support the development and maintenance of secondary sexual characteristics and behaviour. Further control of the GnRH-I system is provided by neuropeptides and neurotransmitters – epinephrine (E), norepinephrine (NE), dopamine (DA), vasotocin (VT), β -endorphin (END), enkephalin, (ENK) and the γ -aminobutyric acid-ergic (GABA) systems.5
- Figure 1.2** Comparison of eleven GnRH peptides. Bold type amino acids indicate changes with respect to the mammalian form.....7
- Figure 2.1** Annual cycle of daylength (including civil twilight) (A), minimum and maximum daily air temperatures (B) and daily rainfall (C) for Palmerston North, New Zealand (40° 21' S, 175° 37' E) from 1 January 1999 to 31 January 2000.....44
- Figure 2.2** Periods of egg laying and daily percentage of female quail incubating eggs in nests. Quail were maintained in a semi-natural environment under natural conditions from 11 January 1999 to 6 January 2000.....47
- Figure 2.3** Changes in mean body weight of female quail maintained in a semi-natural environment under natural conditions from 11 January 1999 to 6 January 2000. Results are shown as mean \pm standard error.....51
- Figure 2.4** Individual body weights of male quail maintained in a semi-natural environment under natural conditions from 11 January 1999 to 6 January 2000. Results are shown as individual data points.....52
- Figure 2.5** Mean cloacal opening diameters of female quail maintained in a semi-natural environment under natural conditions from 11 January 1999 to 6 January 2000. Results are shown as mean \pm standard error..... 54
- Figure 2.6** Individual profiles of the width of the cloacal opening, LH and FSH in female quail maintained in a semi-natural environment under natural conditions from 11 January 1999 to 6 January 2000. “I” indicates birds that were incubating eggs at the time of the collection of the blood sample.....55
- Figure 2.7** Individual profiles of the width of the cloacal opening, LH and FSH in female quail maintained in a semi-natural environment under natural conditions from 11 January 1999 to 6 January 2000. “I” indicates birds

	that were incubating eggs at the time of the collection of the blood sample.....	56
Figure 2.8	Individual profiles of the width of the cloacal opening, LH and FSH in female quail maintained in a semi-natural environment under natural conditions from 11 January 1999 to 6 January 2000. “I” indicates birds that were incubating eggs at the time of the collection of the blood sample.....	57
Figure 2.9	Individual profiles of the cloacal gland area, LH and FSH in male quail maintained in a semi-natural environment under natural conditions from 11 January 1999 to 6 January 2000.....	58
Figure 2.10	Mean LH concentration of female quail maintained in a semi-natural environment under natural conditions from 11 January 1999 to 6 January 2000. Results are shown as mean \pm standard error.....	60
Figure 2.11	Changes in mean FSH concentration of female quail maintained in a semi-natural environment under natural conditions from 11 January 1999 to 6 January 2000. Results are shown as mean \pm standard error.....	62
Figure 2.12	Changes in mean prolactin concentration of female quail maintained in a semi-natural environment under natural conditions from 11 January 1999 to 6 January 2000. Results are shown as mean \pm standard error.....	64
Figure 2.13	Individual profiles of prolactin concentrations in female and male quail maintained in a semi-natural environment under natural conditions from 11 January 1999 to 6 January 2000. Results are shown as individual data points. “I” indicates birds that were incubating eggs at the time of the collection of the blood sample.....	65
Figure 2.14	Prolactin concentrations in female quail that were not incubating eggs, incubating eggs or caring for young maintained in a semi-natural environment under natural conditions from 11 January 1999 to 6 January 2000. Results are shown as mean \pm standard error.....	66
Figure 2.15	Changes in mean T4 and T3 concentrations of female quail maintained in a semi-natural environment under natural conditions from 11 January 1999 to 6 January 2000. Results are shown as mean \pm standard error...	68
Figure 2.16	Individual profiles of T4 and T3 concentrations in female quail maintained in a semi-natural environment under natural conditions from 11 January 1999 to 6 January 2000.....	69
Figure 2.17	Individual profiles of T4 and T3 concentrations in male quail maintained in a semi-natural environment under natural conditions from 11 January 1999 to 6 January 2000.....	71
Figure 2.18	Summary graphs of variables measured in female quail maintained in a semi-natural environment under natural conditions from 11 January 1999 to 6 January 2000.....	73

- Figure 3.1** Parallelism demonstrated for Japanese quail plasma samples A: testosterone, B: estradiol, C: progesterone, D: thyroxine and E: triiodothyronine. Curves with filled squares are standard curves and all other curves are samples.....96
- Figure 3.2** Changes in body weight of male quail transferred from short days to long days, quail transferred from long days to short days and quail held on short days at 10 °C (short day control) and long days at 20 °C (long day control). Results are shown as mean \pm standard error.....99
- Figure 3.3** Cloacal gland areas of individual quail transferred from short days to long days (A), quail transferred from long days to short days (B) and quail held on long days at 20 °C (LD) or short days at 10 °C (SD).....103
- Figure 3.4** Changes in cloacal gland area of male quail transferred from short days to long days, quail transferred from long days to short days and quail held on short days at 10 °C (short day control) and long days at 20 °C (long day control). Results are shown as mean \pm standard error.....104
- Figure 3.5** Relationships between cloacal gland area and paired testis weight (A) and cloacal gland area and testosterone (B) in quail transferred from short days to long days for 35 days.....105
- Figure 3.6** Relationships between cloacal gland area and paired testis weight (A) and cloacal gland area and testosterone (B) in quail transferred from long days to short days for 35 days.....106
- Figure 3.7** Changes in the number of male quail transferred from short days to long days, quail transferred from long days to short days and quail held on short days at 10 °C (short day control) and long days at 20 °C (long day control).....110
- Figure 3.8** Paired testis weights of individual quail transferred from short days to long days (A), quail transferred from long days to short days (B) and quail held on long days at 20 °C (LD) and short days at 10 °C (SD)...113
- Figure 3.9** Changes in paired testis weights of male quail transferred from short days to long days, quail transferred from long days to short days and quail held on short days at 10 °C (short day control) and long days at 20 °C (long day control). Results for control groups are shown on day 36 for simplicity. Results are shown as mean \pm standard error.....114
- Figure 3.10** Relationships between paired testis weight and T, LH and FSH (A-C) in quail transferred from short days to long days for 35 days. Note different scales on x axes.....115
- Figure 3.11** Relationships between paired testis weight and T, LH and FSH (A-C) in quail transferred from long days to short days for 35 days. Note different scales on x and y axes.....116

- Figure 3.12** LH concentrations of individual quail transferred from short days to long days (A), quail transferred from long days to short days (B) and quail held on long days at 20 °C (LD) and short days at 10 °C (SD).....119
- Figure 3.13** Changes in LH concentrations of male quail transferred from short days to long days, quail transferred from long days to short days and quail held on short days at 10 °C (short day control) and long days at 20 °C (long day control). Results for control groups are shown on day 36 for simplicity. Results are shown as mean \pm standard error.....120
- Figure 3.14** FSH concentrations of individual quail transferred from short days to long days (A), quail transferred from long days to short days (B) and quail held on long days at 20 °C (LD) and short days at 10 °C (SD). Note different scales on y axes.....123
- Figure 3.15** Changes in FSH concentrations of male quail transferred from short days to long days, quail transferred from long days to short days and quail held on short days at 10 °C (short day control) and long days at 20 °C (long day control). Results for control groups are shown on day 36 for simplicity. Results are shown as mean \pm standard error.....124
- Figure 3.16** Relationships between LH and FSH in quail transferred from short days to long days (A) and in quail transferred from long days to short days (B) for 35 days. Note different scales on x and y axes.....125
- Figure 3.17** T concentrations of individual quail transferred from short days to long days (A), quail transferred from long days to short days (B) and quail held on long days at 20 °C (LD) and short days at 10 °C (SD).....128
- Figure 3.18** Changes in T concentrations of male quail transferred from short days to long days, quail transferred from long days to short days and quail held on short days at 10 °C (short day control) and long days at 20 °C (long day control). Results for control groups are shown on day 36 for simplicity. Results are shown as mean \pm standard error.....129
- Figure 3.19** Relationships between T and LH (A) and FSH (B) in male quail transferred from short days to long days for 35 days.....130
- Figure 3.20** Relationships between T and LH (A) and FSH (B) in male quail transferred from long days to short days for 35 days.....131
- Figure 3.21** Prolactin concentrations of individual quail transferred from short days to long days (A), quail transferred from long days to short days (B) and quail held on long days at 20 °C (LD) and short days at 10 °C (SD). Note different scales on y axes.....134
- Figure 3.22** Changes in prolactin concentrations of male quail transferred from short days to long days, quail transferred from long days to short days and quail held on short days at 10 °C (short day control) and long days at 20 °C (long day control). Results for control groups are shown on day 36 for simplicity. Results are shown as mean \pm standard error.....135

- Figure 3.23** T4 concentrations of individual quail transferred from short days to long days (A), quail transferred from long days to short days (B) and quail held on long days at 20 °C (LD) and short days at 10 °C (SD).....138
- Figure 3.24** Changes in T4 concentrations of male quail transferred from short days to long days, quail transferred from long days to short days and quail held on short days at 10 °C (short day control) and long days at 20 °C (long day control). Results for control groups are shown on day 36 for simplicity. Results are shown as mean \pm standard error.....139
- Figure 3.25** Relationships between T4 and LH in male quail transferred from short days to long days (A) and in male quail transferred from long days to short days for 35 days. Note different scales on x and y axes.....140
- Figure 3.26** T3 concentrations of individual quail transferred from short days to long days (A), quail transferred from long days to short days (B) and quail held on long days at 20 °C (LD) and short days at 10 °C (SD).....143
- Figure 3.27** Changes in T3 concentrations of male quail transferred from short days to long days, quail transferred from long days to short days and quail held on short days at 10 °C (short day control) and long days at 20 °C (long day control). Results for control groups are shown on day 36 for simplicity. Results are shown as mean \pm standard error.....144
- Figure 3.28** Relationships between T3 and LH (A) and T4 (B) concentrations in quail transferred from short days to long days. Note different scales on y axes.....145
- Figure 3.29** Relationships between T3 and LH (A) and T4 (B) concentrations in quail transferred from long days to short days. Note different scales on y axes.....146
- Figure 3.30** Summary of variables measured in male quail transferred from short days to long days for 35 days. Results are presented as mean \pm standard error. Note different scales on y axes.....149
- Figure 3.31** Summary of variables measured in male quail transferred from long days to short days for 35 days. Results are presented as mean \pm standard error. Note different scales on y axes.....150
- Figure 3.32** Changes in body weight of female birds transferred from short days to long days, birds transferred from long days to short days and birds held on short days at 10 °C (short day control) and long days at 20 °C (long day control). Results are shown as mean \pm standard error.....153
- Figure 3.33** Width of cloacal opening of individual quail transferred from short days to long days (A), quail transferred from long days to short days (B) and quail held on long days at 20 °C (LD) and short days at 10 °C (SD)...156

- Figure 3.34** Changes in the width of the cloacal opening of female quail transferred from short days to long days, quail transferred from long days to short days and quail held on short days at 10 °C (short day control) and long days at 20 °C (long day control). Results for control groups are shown on day 36 for simplicity. Results are shown as mean \pm standard error.....157
- Figure 3.35** Relationships between the width of the cloacal opening and ovary weight (A), and the width of the cloacal opening and oviduct weight (B) of female quail transferred from short days to long days for 35 days. Note different scale on x axes.....158
- Figure 3.36** Relationships between the width of the cloacal opening and ovary weight (A), and the width of the cloacal opening and oviduct weight (B) of female quail transferred from long days to short days for 35 days. Note different scales on the y axes.....159
- Figure 3.37** Ovary weights of individual quail transferred from short days to long days (A), quail transferred from long days to short days (B) and quail held on long days at 20 °C (LD) and short days at 10 °C (SD).....165
- Figure 3.38** Changes in ovary weight of female quail transferred from short days to long days, quail transferred from long days to short days and quail held on short days at 10 °C (short day control) and long days at 20 °C (long day control). Results for control groups are shown on day 36 for simplicity. Results are shown as mean \pm standard error.....166
- Figure 3.39** Relationships between ovary weight and oviduct weight (A), LH (B), FSH (C), estradiol (D) and P₄ (E) of quail transferred from short days to long days for 35 days. Note different scales on y axes.....167
- Figure 3.40** Relationships between ovary weight and oviduct weight (A), LH (B), FSH (C), estradiol (D) and P₄ (E) of quail transferred from long days to short days. Note different scales on y axes.....168
- Figure 3.41** Daily egg production of female quail transferred from short days to long days, quail transferred from long days to short days and quail held on short days at 10 °C (short day control) and long days at 20 °C (long day control).....171
- Figure 3.42** Oviduct weights of individual quail transferred from short days to long days (A), quail transferred from long days to short days (B) and quail held on long days at 20 °C (LD) and short days at 10 °C (SD).....174
- Figure 3.43** Changes in oviduct weight of female quail transferred from short days to long days, quail transferred from long days to short days and quail held on short days at 10 °C (short day control) and long days at 20 °C (long day control). Results for control groups are shown on day 36 for simplicity. Results are shown as mean \pm standard error.....175

- Figure 3.44** Relationships between oviduct weight and LH, FSH, estradiol and progesterone (A-D) of quail transferred from short days to long days for 35 days.....176
- Figure 3.45** Relationships between oviduct weight and LH, FSH, estradiol and progesterone (A-D) of quail transferred from long days to short days for 35 days. Note different scales on y axes.....177
- Figure 3.46** LH concentrations of individual quail transferred from short days to long days (A), quail transferred from long days to short days (B) and quail held on long days at 20 °C (LD) and short days at 10 °C (SD).....180
- Figure 3.47** Changes in LH concentrations of female quail transferred from short days to long days, quail transferred from long days to short days and quail held on short days at 10 °C (short day control) and long days at 20 °C (long day control). Results for control groups are shown on day 36 for simplicity. Results are shown as mean \pm standard error.....181
- Figure 3.48** FSH concentrations of individual quail transferred from short days to long days (A), quail transferred from long days to short days (B) and quail held on long days at 20 °C (LD) and short days at 10 °C (SD). Note different scales on y axes.....184
- Figure 3.49** Changes in FSH concentrations of female quail transferred from short days to long days, quail transferred from long days to short days and quail held on short days at 10 °C (short day control) and long days at 20 °C (long day control). Results for control groups are shown on day 36 for simplicity. Results are shown as mean \pm standard error.....185
- Figure 3.50** Relationships between LH and FSH of quail transferred from short days to long days (A) and quail transferred from long days to short days (B) for 35 days. Note different scales on x and y axes.....186
- Figure 3.51** Estradiol concentrations of individual quail transferred from short days to long days (A), quail transferred from long days to short days (B) and quail held on long days at 20 °C (LD) and short days at 10 °C (SD)...189
- Figure 3.52** Changes in estradiol concentrations of female quail transferred from short days to long days, quail transferred from long days to short days and quail held on short days at 10 °C (short day control) and long days at 20 °C (long day control). Results for control groups are shown on day 36 for simplicity. Results are shown as mean \pm standard error.....190
- Figure 3.53** Relationships between estradiol and LH (A) and estradiol and FSH (B) of quail transferred from short days to long days for 35 days.....191
- Figure 3.54** Relationships between estradiol and LH (A) and estradiol and FSH (B) of quail transferred from long days to short days for 35 days.....192
- Figure 3.55** Progesterone concentrations of individual quail transferred from short days to long days (A), quail transferred from long days to short days (B)

- and quail held on long days at 20 °C (LD) and short days at 10 °C (SD).....195
- Figure 3.56** Changes in progesterone concentrations of female quail transferred from short days to long days, quail transferred from long days to short days and quail held on short days at 10 °C (short day control) and long days at 20 °C (long day control). Results for control groups are shown on day 36 for simplicity. Results are shown as mean \pm standard error.....196
- Figure 3.57** Relationships between progesterone and LH (A), FSH (B) and estradiol (C) of quail transferred from short days to long days for 35 days. Note different scales on y axes.....197
- Figure 3.58** Relationships between progesterone and LH (A), FSH (B) and estradiol (C) of quail transferred from long days to short days for 35 days. Note different scales on y axes.....198
- Figure 3.59** Prolactin concentrations of individual quail transferred from short days to long days (A), quail transferred from long days to short days (B) and quail held on long days at 20 °C (LD) and short days at 10 °C (SD)...201
- Figure 3.60** Changes in prolactin concentrations of female quail transferred from short days to long days, quail transferred from long days to short days and quail held on short days at 10 °C (short day control) and long days at 20 °C (long day control). Results for control groups are shown on day 36 for simplicity. Results are shown as mean \pm standard error.....202
- Figure 3.61** T4 concentrations of individual quail transferred from short days to long days (A), quail transferred from long days to short days (B) and quail held on long days at 20 °C (LD) and short days at 10 °C (SD).....205
- Figure 3.62** Changes in T4 concentrations of female quail transferred from short days to long days, quail transferred from long days to short days and quail held on short days at 10 °C (short day control) and long days at 20 °C (long day control). Results for control groups are shown on day 36 for simplicity. Results are shown as mean \pm standard error.....206
- Figure 3.63** Relationships between thyroxine and LH of quail transferred from short days to long days (A) and quail transferred from long days to short days for 35 days. Note different scales on x and y axes.....207
- Figure 3.64** T3 concentrations of individual female quail transferred from short days to long days (A), quail transferred from long days to short days (B) and quail held on long days at 20 °C (LD) and short days at 10 °C (SD)...210
- Figure 3.65** Changes in T3 concentrations of female quail transferred from short days to long days, quail transferred from long days to short days and quail held on short days at 10 °C (short day control) and long days at 20 °C (long day control). Results for control groups are shown on day 36 for simplicity. Results are shown as mean \pm standard error.....211

- Figure 3.66** Relationships between T3 and LH (A) and between T3 and T4 (B) of quail transferred from short days to long days for 35 days.....212
- Figure 3.67** Correlation between T3 and LH (A) and between T3 and T4 (B) of quail transferred from long days to short days for 35 days.....213
- Figure 3.68** Summary graphs of variables measured in female quail transferred from short days to long days for 35 days. Results are shown as mean \pm standard error. Note different scales on y axes.....216
- Figure 3.69** Summary graphs of variables measured in female quail transferred from long days to short days for 35 days. Results are shown as mean \pm standard error. Note different scales on y axes.....217
- Figure 4.1** Parallelism demonstrated for LH concentrations in Japanese quail plasma samples.....240
- Figure 4.2** LH concentrations in individual female quail treated with 0.25 (A), 0.5 (B), 1 (C), 2.5 (D) or 5 (E) μg cGnRH-I/kg body weight.....243
- Figure 4.3** Mean LH concentrations in female quail treated with saline or 0.25, 0.5, 1, 2.5 or 5 μg cGnRH-I/kg body weight. Results are shown as mean \pm standard error.....244
- Figure 4.4** LH concentrations in individual female quail treated with 0.25 (A), 0.5 (B), 1 (C), 2.5 (D) or 5 μg (E) cGnRH-II /kg body weight.....248
- Figure 4.5** Mean LH concentrations in female quail treated with saline or 0.25, 0.5, 1, 2.5 or 5 μg cGnRH-II/kg of body weight. Results are shown as mean \pm standard error.....249
- Figure 4.6** LH concentrations in individual female quail treated with 0.5 (A), 1 (B), 2.5 (C) or 4 (D) μg buserelin/kg body weight.....253
- Figure 4.7** Mean LH concentrations in female quail treated with saline or 0.5, 1, 0.25 or 4 μg buserelin/kg body weight. Results are shown as mean \pm standard error.....254
- Figure 4.8** LH concentrations in individual female quail treated with 0.2 (A), 1 (B), 2.5 (C) or 5 (D) μg D-Lys⁶ Trp⁷Tyr⁸-GnRH/kg body weight.....258
- Figure 4.9** Mean LH concentrations in female quail treated with saline or 0.2, 1, 2.5 or 5 μg D-Lys⁶Trp⁷Tyr⁸-GnRH/kg body weight. Results are shown as mean \pm standard error.....259
- Figure 4.10** Mean LH concentrations in female quail treated with either buserelin, cGnRH-I, cGnRH-II or D-Lys⁶ Trp⁷Tyr⁸-GnRH at 0.5 μg , 1 μg , 2.5 μg or 5 μg /kg. Results are shown as mean \pm standard error. Note different scales on y axis.....263

- Figure 4.11** Mean width of cloacal opening in female quail injected daily with saline or PVP, or transferred to a long day photoperiod (16L:8D) on day 0. Results are shown as mean \pm standard error.....268
- Figure 4.12** Mean ovary weights, oviduct weights and LH concentrations in female quail euthanased at the start of the experiment (reference), injected daily with saline or PVP or transferred to long daylengths for 7 days. Results are shown as mean \pm standard error.....270
- Figure 4.13** Mean width of cloacal opening in female quail treated daily with 0.5 μ g, 1 μ g, 2.5 μ g or 5 μ g cGnRH-II/kg body weight dissolved in saline (A) or PVP (B) for 7 days. The mean width of cloacal opening of birds transferred to long days is also shown for comparison with treated birds. Results are shown as mean \pm standard error.....272
- Figure 4.14** Mean ovary weights, oviduct weights and LH concentrations in female quail treated daily with 0.5, 1, 2.5 or 5 μ g cGnRH-II/kg dissolved in saline or PVP for 7 days or transferred to long days. Results are shown as mean \pm standard error. Note different scales on y axes.....276
- Figure 4.15** Mean width of cloacal opening in female quail treated daily with 0.2 μ g, 1 μ g, 2.5 μ g or 5 μ g D-Lys⁶Trp⁷Tyr⁸-GnRH/kg body weight dissolved in saline (A) or PVP (B) for 7 days. The mean width of cloacal opening of birds transferred to long days is also shown for comparison with treated birds. Results are shown as mean \pm standard error.....283
- Figure 4.16** Mean ovary weights, oviduct weights and LH concentrations in female quail treated daily with 0.2, 1, 2.5 or 5 μ g D-Lys⁶Trp⁷Tyr⁸-GnRH/kg dissolved in saline or PVP for 7 days or transferred to long days. Results are shown as mean \pm standard error. Note different scales on y axes..287
- Figure 5.1** Individual LH concentrations in birds after a single injection of saline (A), 2.5 (B) or 5 (C) μ g D-Lys⁶Trp⁷Tyr⁸-GnRH/kg.....298
- Figure 5.2** Mean LH concentrations in birds after a single injection of saline, 2.5 or 5 μ g D-Lys⁶Trp⁷Tyr⁸-GnRH/kg. Results are shown as mean \pm standard error.....299
- Figure 5.3** Individual LH concentrations in birds after a second injection of saline (A), 2.5 (B) or 5 (C) μ g D-Lys⁶Trp⁷Tyr⁸-GnRH/kg given 6 hours after the first injection.....301
- Figure 5.4** Mean LH concentrations in birds after a second injection of saline, 2.5 or 5 μ g D-Lys⁶Trp⁷Tyr⁸-GnRH/kg given 6 hours after the first injection. Results are shown as mean \pm standard error.....302
- Figure 5.5** Individual LH concentrations in birds after a third injection of saline (A), 2.5 (B) or 5 (C) μ g D-Lys⁶Trp⁷Tyr⁸-GnRH/kg given 6 hours after the second injection and 12 hours after the first injection.....305

- Figure 5.6** Mean LH concentrations in birds after a third injection of saline, 2.5 or 5 μg D-Lys⁶Trp⁷Tyr⁸-GnRH/kg given 6 hours after the second injection and 12 hours after the first injection. Results are shown as mean \pm standard error.....306
- Figure 5.7** Individual LH concentrations in birds after a fourth injection of saline (A), 2.5 (B) or 5 (C) μg D-Lys⁶Trp⁷Tyr⁸-GnRH/kg given 6 hours after the third injection, 12 hours after the second injection and 18 hours after the first injection.....308
- Figure 5.8** Mean LH concentrations in birds after a fourth injection of saline, 2.5 or 5 μg D-Lys⁶Trp⁷Tyr⁸-GnRH/kg given 6 hours after the third injection, 12 hours after the second injection and 18 hours after the first injection. Results are shown as mean \pm standard error.....309
- Figure 5.9** Changes in mean LH concentrations in birds after one, two, three or four injections of 2.5 (A) or 5 (B) μg D-Lys⁶Trp⁷Tyr⁸-GnRH/kg given 6 hours apart. Results are shown as mean \pm standard error.....311
- Figure 5.10** Mean LH concentrations in birds before treatment (0 h) and at the end of treatment (24 h). Results are shown as mean \pm standard error.....314
- Figure 6.1** Mean width of cloacal opening in female quail held on short days (8L:16D), long days (16L:8D) and short days with treatment of 5, 10, 15 or 20 μg D-Lys⁶Trp⁷Tyr⁸-GnRH/kg/day for 14 days by osmotic pump. Results are shown as mean \pm standard error.....324
- Figure 6.2** Individual LH concentrations in female quail held on short days (8L:16D), long days (16L:8D) and short days with treatment of 5, 10, 15 or 20 μg D-Lys⁶Trp⁷Tyr⁸-GnRH/kg/day for 14 days by osmotic pump. Note the use of a different y axis scale for the birds transferred to long days.....327
- Figure 6.3** Mean LH concentrations in female quail held on short days (8L:16D), long days (16L:8D) and short days with treatment of 5, 10, 15 or 20 μg D-Lys⁶Trp⁷Tyr⁸-GnRH/kg/day (A) and in treated and non-treated quail held on short days only (B) for 14 days by osmotic pump. Results are shown as mean \pm standard error.....328
- Figure 6.4** Mean ovary weights of female quail held on short days (8L:16D), long days (16L:8D) and short days with treatment of 5, 10, 15 or 20 μg D-Lys⁶Trp⁷Tyr⁸-GnRH/kg/day for 14 days by osmotic pump. Results are shown as mean \pm standard error.....333
- Figure 6.5** Mean oviduct weights of female quail held on short days (8L:16D), long days (16L:8D) and short days with treatment of 5, 10, 15 or 20 μg D-Lys⁶Trp⁷Tyr⁸-GnRH/kg/day for 14 days by osmotic pump. Results are shown as mean \pm standard error.....335

List of Tables

Table 1.1	Homologies (percentage) for amino acid sequence of mature GnRH peptides including the processing site9
Table 2.1	Legband identification of quail maintained in a semi-natural environment under natural conditions from 11 January 1999 to 6 January 2000.....36
Table 2.2	Social groups of quail maintained outdoors in a semi-natural environment from 11 January 1999 to 6 January 2000.....45
Table 2.3	One-way repeat measures ANOVA for the width of the cloacal opening of female quail maintained outdoors in a semi-natural environment.... 59
Table 2.4	One-way repeat measures ANOVA for mean LH concentrations in female quail maintained outdoors in a semi-natural environment.....61
Table 2.5	Friedmans non-parametric ANOVA for mean FSH concentrations in female quail maintained outdoors in a semi-natural environment.....63
Table 2.6	One-way single measures ANOVA for mean prolactin concentrations in female quail maintained outdoors in a semi-natural environment.....68
Table 3.1	Two-way single measures ANOVA for body weight of male quail transferred from short days to long days, quail transferred from long days to short days and quail held on short days at 10 °C (short day control) and long days at 20 °C (long day control).....100
Table 3.2	One-way repeated measures ANOVA for body weight of male quail held on long days at 20 °C (long day control) and held on short days at 10 °C (short day control).....101
Table 3.3	One-way single measures ANOVA for cloacal gland area of male quail transferred from short days to long days and quail held on long days at 20 °C (long day control).....107
Table 3.4	One-way single measures ANOVA for cloacal gland area of male quail transferred from long days to short days and quail held on short days at 10 °C (short day control).....107
Table 3.5	One way repeated measures ANOVA for cloacal gland area of male quail held on long days at 20 °C and male quail held on short days at 10 °C.....108
Table 3.6	Two-way single measures ANOVA for cloacal gland area on day 35 of male quail transferred from short days to long days, quail transferred from long days to short days and quail held on short days at 10 °C (SD - short day control) and long days at 20 °C (LD - long day control).....108

Table 3.7	Regression analyses for male quail transferred from short days to long days (A) and for male quail transferred from long days to short days (B).....	109
Table 3.8	Cloacal foam production and corresponding measurements for cloacal gland area and testosterone concentrations of male quail transferred from short days to long days, quail transferred from long days to short days and quail held on short days at 10 °C (short day control) or long days at 20 °C (long day control).....	111
Table 3.9	Kruskal-Wallis (K-W) non-parametric ANOVA and Mann-Whitney (M-W) U test contrasts for paired testis weight of quail transferred from short days to long days and quail held on long days at 20 °C (long day control).....	117
Table 3.10	Kruskal-Wallis (K-W) non-parametric ANOVA and Mann-Whitney (M-W) U test contrasts for paired testis weight of quail transferred from long days to short days and quail held on short days at 10 °C (short day control).....	117
Table 3.11	Two-way single measures ANOVA for paired testis weight on day 35 of male quail transferred from short days to long days, quail transferred from long days to short days and quail held on short days at 10 °C (SD - short day control) and long days at 20 °C (LD - long day control).....	118
Table 3.12	Two-way single measures ANOVA for LH concentrations of male quail transferred from short days to long days, quail transferred from long days to short days and quail held on short days at 10 °C (short day control) and long days at 20 °C (long day control).....	121
Table 3.13	Two-way single measures ANOVA for FSH concentrations of male quail transferred from short days to long days, quail transferred from long days to short days and quail held on short days at 10 °C (short day control) and long days at 20 °C (long day control).....	126
Table 3.14	Two-way single measures ANOVA for T concentrations of male quail transferred from short days to long days, quail transferred from long days to short days and quail held on short days at 10 °C (short day control) and long days at 20 °C (long day control).....	132
Table 3.15	Two-way single measures ANOVA for prolactin concentrations of male quail transferred from short days to long days, quail transferred from long days to short days and quail held on short days at 10 °C (short day control) and long days at 20 °C (long day control).....	136
Table 3.16	Two-way single measures ANOVA for T4 concentrations of male quail transferred from short days to long days, quail transferred from long days to short days and quail held on short days at 10 °C (short day control) and long days at 20 °C (long day control).....	141

Table 3.17	Kruskal-Wallis (K-W) non-parametric ANOVA and Mann-Whitney (M-W) U test contrasts for T3 concentrations of male quail transferred from short days to long days and quail held on long days at 20 °C (long day control).....	147
Table 3.18	Kruskal-Wallis (K-W) non-parametric ANOVA and Mann-Whitney (M-W) U test contrasts for T3 concentrations of quail transferred from long days to short days and quail held on short days at 10 °C (short day control).....	147
Table 3.19	Two-way single measures ANOVA for T3 concentrations on day 35 of male quail transferred from short days to long days, quail transferred from long days to short days and quail held on short days at 10 °C (SD - short day control) and long days at 20 °C (LD - long day control).....	148
Table 3.20	Summary of the timing of significant changes in variables measured in male quail transferred from short days to long days and in male quail transferred from long days to short days (first significant change compared with day 0 indicated in bold type for each variable).....	151
Table 3.21	Two-way single measures ANOVA for body weight of female quail transferred from short days to long days, quail transferred from long days to short days and quail held on short days at 10 °C (short day control) and long days at 20 °C (long day control).....	154
Table 3.22	One-way repeated measures ANOVA for body weight of female quail held on long days at 20 °C (long day control) and held on short days at 10 °C (short day control).....	155
Table 3.23	One-way single measures ANOVAs for the width of the cloacal opening of female quail transferred from short days to long days and quail held on long days at 20 °C (long day control).....	160
Table 3.24	One-way single measures ANOVAs for the width of the cloacal opening of female quail transferred from long days to short days and quail held on short days at 10 °C (short day control).....	160
Table 3.25	One-way repeated measures ANOVA for the width of the cloacal opening of female quail held on long days at 20 °C (long day control) and held on short days at 10 °C (short day control).....	161
Table 3.26	Two-way single measures ANOVA for the width of the cloacal opening on day 35 of female quail transferred from short days to long days, quail transferred from long days to short days and quail held on short days at 10 °C (SD - short day control) and long days at 20 °C (LD - long day control).....	161
Table 3.27	Regression analyses for female quail transferred from short days to long days (A) and for female quail transferred from long days to short days (B).....	162

Table 3.28	Kruskal-Wallis (K-W) non-parametric ANOVA and Mann-Whitney (M-W) U test contrasts for ovary weight of female quail transferred from short days to long days and quail held on long days at 20 °C (long day control).....	169
Table 3.29	Kruskal-Wallis (K-W) non-parametric ANOVA and Mann-Whitney (M-W) U test contrasts for ovary weight of female quail transferred from long days to short days and quail held on short days at 10 °C (short day control).....	169
Table 3.30	Two way single measures ANOVA for ovary weight on day 35 of female quail transferred from short days to long days, quail transferred from long days to short days and quail held on short days at 10 °C (SD - short day control) and long days at 20 °C (LD - long day control).....	170
Table 3.31	Number and size of large yellow follicles present on the ovaries and corresponding ovary weights of quail transferred from short days to long days and quail maintained on long days at 20 °C (long day control) (A) and quail transferred from long days to short days and quail maintained on short days at 10 °C (short day control) (B). Numbers in parentheses denote the number of birds with follicles in the diameter range. Follicles were divided into six classes based on follicle diameter data recorded for the New Zealand strain of Japanese quail (Bennett, 2002).....	172
Table 3.32	Kruskal-Wallis (K-W) non-parametric ANOVA and Mann-Whitney (M-W) U test contrasts for oviduct weight of female quail transferred from short days to long days and quail held on long days at 20 °C (long day control).....	178
Table 3.33	One way single measures ANOVA for oviduct weight of female quail transferred from long days to short days and quail held on short days at 10 °C (short day control).....	178
Table 3.34	Two-way single measures ANOVA for oviduct weight on day 35 of female quail transferred from short days to long days, quail transferred from long days to short days and quail held on short days at 10 °C (SD - short day control) and long days at 20 °C (LD - long day control).....	179
Table 3.35	One way single measures ANOVA for LH concentrations of female quail transferred from short days to long days and quail held on long days at 20 °C (long day control).....	182
Table 3.36	One way single measures ANOVA for LH concentrations of female quail transferred from long days to short days and quail held on short days at 10 °C (short day control).....	182
Table 3.37	Two way single measures ANOVA for LH concentrations on day 35 of female quail transferred from short days to long days, quail transferred from long days to short days and quail held on short days at 10 °C (SD - short day control) and long days at 20 °C (LD - long day control).....	183

Table 3.38	Two-way single measures ANOVA for FSH concentrations of female quail transferred from short days to long days, quail transferred from long days to short days and quail held on short days at 10 °C (short day control) and long days at 20 °C (long day control).....	187
Table 3.39	Two-way single measures ANOVA for estradiol concentrations of female quail transferred from short days to long days, quail transferred from long days to short days and quail held on short days at 10 °C (short day control) and long days at 20 °C (long day control).....	193
Table 3.40	Two-way single measures ANOVA for P ₄ concentrations of female quail transferred from short days to long days, quail transferred from long days to short days and quail held on short days at 10 °C (short day control) and long days at 20 °C (long day control).....	199
Table 3.41	One way single measures ANOVA for prolactin concentrations of female quail transferred from short days to long days and quail held on long days at 20 °C (long day control).....	203
Table 3.42	One way single measures ANOVA for prolactin concentrations of female quail transferred from long days to short days and quail held on short days at 10 °C (short day control).....	203
Table 3.43	Two-way single measures ANOVA for prolactin concentrations on day 35 of female quail transferred from short days to long days, quail transferred from long days to short days and quail held on short days at 10 °C (SD - short day control) and long days at 20 °C (LD - long day control).....	204
Table 3.44	Two-way single measures ANOVA for T4 concentrations of female quail transferred from short days to long days, quail transferred from long days to short days and quail held on short days at 10 °C (short day control) and long days at 20 °C (long day control).....	208
Table 3.45	Kruskal-Wallis (K-W) non-parametric ANOVA and Mann-Whitney (M-W) U test contrasts for T3 concentrations of female quail transferred from short days to long days and quail held on long days at 20 °C (long day control).....	214
Table 3.46	Kruskal-Wallis (K-W) non-parametric ANOVA and Mann-Whitney (M-W) U test contrasts for T3 concentrations of female quail transferred from long days to short days and quail held on short days at 10 °C (short day control).....	214
Table 3.47	Two-way single measures ANOVA for T3 concentrations on day 35 of female quail transferred from short days to long days, quail transferred from long days to short days and quail held on short days at 10 °C (SD - short day control) and long days at 20 °C (LD - long day control).....	215
Table 3.48	Summary of the timing of significant changes in variables measured in female quail transferred from short days to long days and in female quail	

	transferred from long days to short days. First significant change from day 0 in each variable is indicated in bold type.....	218
Table 3.49	Testicular growth rates for male Japanese quail transferred to photoperiods ≥ 16 h light per day.....	219
Table 4.1	Experimental groups for experiment one.....	235
Table 4.2	Experimental groups for experiment two.....	235
Table 4.3	Experimental groups for experiment three.....	236
Table 4.4	Two way repeated measures ANOVA for LH concentrations in female quail treated with saline and 0.25, 0.5, 1, 2.5 and 5 μg cGnRH-I/kg body weight.....	245
Table 4.5	Two way repeated measures ANOVA for LH concentrations in female quail treated with saline and 0.25, 0.5, 1, 2.5 and 5 μg cGnRH-II/kg body weight.....	250
Table 4.6	Two way repeated measures ANOVA for LH concentrations in female quail treated with saline or 0.5, 1, 2.5 or 4 μg buserelin/kg body weight.....	255
Table 4.7	Two way repeated measures ANOVA for LH concentrations in female quail treated with saline and 0.2, 1, 2.5 and 5 μg D-Lys ⁶ Trp ⁷ Tyr ⁸ -GnRH/kg body weight.....	260
Table 4.8	Two way repeated measures ANOVA for LH concentrations in female quail treated with 0.5 μg cGnRH-I, cGnRH-II or buserelin/kg body weight.....	264
Table 4.9	Two way repeated measures ANOVA for LH concentrations in female quail treated with 1 μg cGnRH-I, cGnRH-II, buserelin or D-Lys ⁶ Trp ⁷ Tyr ⁸ -GnRH /kg body weight.....	265
Table 4.10	Two way repeated measures ANOVA for LH concentrations in female quail treated with 2.5 μg cGnRH-I, cGnRH-II, buserelin or D-Lys ⁶ Trp ⁷ Tyr ⁸ -GnRH /kg body weight.....	266
Table 4.11	Two way repeated measures ANOVA for LH concentrations in female quail treated with 5 μg cGnRH-I, cGnRH-II or D-Lys ⁶ Trp ⁷ Tyr ⁸ -GnRH /kg body weight.....	267
Table 4.12	Two way repeated measures ANOVA for cloacal diameter in female quail injected daily with saline, PVP or transferred to long days for 7 days.....	269
Table 4.13	Single measures ANOVAs for ovary weights (A), oviduct weights (B) and LH concentrations (C) in female quail before treatment (reference),	

	quail injected daily with saline, PVP or transferred to long days for 7 days.....	271
Table 4.14	Two way repeated measures ANOVA for cloacal diameter in female quail injected daily with 0.5, 1, 2.5, 5 μg cGnRH-II /kg body weight in saline and birds transferred to long days for 7 days.....	273
Table 4.15	Two way repeated measures ANOVA for cloacal diameter in female quail injected daily with 0.5, 1, 2.5, 5 μg cGnRH-II /kg body weight in PVP and birds transferred to long days for 7 days.....	274
Table 4.16	One way ANOVA for ovary weights of female quail injected daily with 0.5, 1, 2.5, 5 μg cGnRH-II /kg or 0.5, 1, 2.5, 5 μg D-Lys ⁶ Trp ⁷ Tyr ⁸ -GnRH /kg in saline and birds transferred to long days for 7 days.....	277
Table 4.17	One way ANOVA for oviduct weights of female quail injected daily with 0.5, 1, 3, 5 μg cGnRH-II /kg or 0.5, 1, 2.5, 5 μg D-Lys ⁶ Trp ⁷ Tyr ⁸ -GnRH /kg in saline and birds transferred to long days for 7 days.....	278
Table 4.18	One way ANOVA for LH concentrations of female quail injected daily with 0.5, 1, 2.5, 5 μg cGnRH-II /kg or 0.5, 1, 2.5, 5 μg D-Lys ⁶ Trp ⁷ Tyr ⁸ -GnRH /kg in saline and birds transferred to long days for 7 days.....	279
Table 4.19	One way ANOVA for ovary weights of female quail injected daily with 0.5, 1, 2.5, 5 μg cGnRH-II /kg or 0.5, 1, 3, 5 μg D-Lys ⁶ Trp ⁷ Tyr ⁸ -GnRH /kg in PVP and birds transferred to long days for 7 days.....	280
Table 4.20	One way ANOVA for oviduct weights of female quail injected daily with 0.5, 1, 2.5, 5 μg cGnRH-II /kg or 0.5, 1, 3, 5 μg D-Lys ⁶ Trp ⁷ Tyr ⁸ -GnRH /kg in PVP and in birds transferred to long days for 7 days.....	281
Table 4.21	One way ANOVA for LH concentrations of female quail injected daily with 0.5, 1, 2.5, 5 μg cGnRH-II /kg or 0.5, 1, 3, 5 μg D-Lys ⁶ Trp ⁷ Tyr ⁸ -GnRH /kg in PVP and birds transferred to long days for 7 days.....	282
Table 4.22	Two way repeated measures ANOVA for cloacal diameter in female quail injected daily with 0.2, 1, 2.5, 5 μg D-Lys ⁶ Trp ⁷ Tyr ⁸ -GnRH /kg body weight in saline and birds transferred to long days for 7 days....	284
Table 4.23	Two way repeated measures ANOVA for cloacal diameter in female quail injected daily with 0.2, 1, 2.5, 5 μg D-Lys ⁶ Trp ⁷ Tyr ⁸ -GnRH/kg body weight in PVP and birds transferred to long days for 7 days.....	285
Table 5.1	Experimental groups.....	295
Table 5.2	Two way repeat measures ANOVA for LH concentrations in female quail after a single injection of saline or 2.5 or 5 μg D-Lys ⁶ Trp ⁷ Tyr ⁸ – GnRH /kg body weight.....	300

Table 5.3	Two way repeat measures ANOVA for LH concentrations in female quail after two subcutaneous injections (one injection every six hours) of saline, 2.5 or 5 µg D-Lys ⁶ Trp ⁷ Tyr ⁸ -GnRH/kg body weight.....	303
Table 5.4	Two way repeat measures ANOVA for LH concentrations in female quail after three subcutaneous injections (one injection every six hours) of saline, 2.5 or 5 µg D-Lys ⁶ Trp ⁷ Tyr ⁸ -GnRH/kg body weight.....	307
Table 5.5	Two way repeat measures ANOVA for LH concentrations in female quail after four subcutaneous injections (one injection every six hours) of saline, 2.5 or 5 µg D-Lys ⁶ Trp ⁷ Tyr ⁸ -GnRH/kg body weight.....	310
Table 5.6	Two way single measures ANOVA for LH concentrations in female quail after one, two, three or four subcutaneous injections (one injection every six hours) of 2.5 µg D-Lys ⁶ Trp ⁷ Tyr ⁸ -GnRH/kg body weight....	312
Table 5.7	Two way single measures ANOVA for LH concentrations in female quail after one, two, three or four subcutaneous injections (one injection every six hours) of 5 µg D-Lys ⁶ Trp ⁷ Tyr ⁸ -GnRH/kg body weight.....	313
Table 5.8	Two way single measures ANOVA for LH concentrations in female quail after four subcutaneous injections (one injection every six hours) of 5 µg D-Lys ⁶ Trp ⁷ Tyr ⁸ -GnRH/kg body weight.....	315
Table 6.1	Experimental groups.....	320
Table 6.2	Two way repeat measures ANOVA for the width of the cloacal opening in female quail held on short days (8L:16D), long days (16L:8D) and short days with treatment of 5, 10, 15 or 20 µg D-Lys ⁶ Trp ⁷ Tyr ⁸ -GnRH /kg/day for 14 days by osmotic pump.....	325
Table 6.3	Two way repeat measures ANOVA for LH concentrations in female quail held on short days (8L:16D), long days (16L:8D) and short days with treatment of 5, 10, 15 or 20 µg D-Lys ⁶ Trp ⁷ Tyr ⁸ -GnRH /kg/day for 14 days by osmotic pump.....	329
Table 6.4	Single measures ANOVA for ovary weights in female quail held on short days (8L:16D), long days (16L:8D) and short days with treatment of 5, 10, 15 or 20 µg D-Lys ⁶ Trp ⁷ Tyr ⁸ -GnRH /kg/day for 14 days by osmotic pump.....	324
Table 6.5	Single measures ANOVA for oviduct weights in female quail held on short days (8L:16D), long days (16L:8D) and short days with treatment of 5, 10, 15 or 20 µg D-Lys ⁶ Trp ⁷ Tyr ⁸ -GnRH /kg/day for 14 days by osmotic pump.....	326