Copyright is owned by the Author of the thesis. Permission is given for a copy to be downloaded by an individual for the purpose of research and private study only. The thesis may not be reproduced elsewhere without the permission of the Author.

A study of natural lytic *Listeria* phages with decontaminating properties for use in seafood processing plants

A thesis presented in partial fulfilment of the requirements for the degree of

Doctor of Philosophy

in

Food Technology

Institute of Food, Nutrition and Human Health

Massey University

Auckland, New Zealand

Geevika J Ganegama Arachchi

Dedication

The thesis is dedicated to my loving mother and late father, my four siblings and aunties,

Acknowledgements

The success of this study is a result of the support given by many people and organizations at various stages. It would be therefore appropriate for me to acknowledge all the persons and organizations that contributed to the study, but due to limited space, I am only able to name a few. I therefore apologies to those individuals and organizations that have been not mentioned here.

First and foremost, I would like to express my gratitude to Massey University, *alma mater*, for giving me the opportunity to study for the Doctor of Philosophy in Food Technology. I also thank the NZ King Salmon Ltd, Nelson, for funding the practical work; Foundation for Research, Science and Technology (FRST) New Zealand [Contract No.: NZKX0902] for providing a stipend; Cawthron Institute, Nelson; Environmental and Science Research (ESR) Ltd, Christchurch; and New Zealand Plant and Food Research Ltd (PFR), Auckland; The University of Auckland, for various technical support. My sincere gratitude also goes to National Aquatic Resources Research and Development Agency (NARA), Sri Lanka, my employer for granting me study leave.

I am indebted to my dedicated supervisors for their assistance in transforming numerous crude drafts into a fine document present here: Professor Steve Flint for providing overall guidance, Dr Tony Mutukumira, Dr Cristina Cruz, Dr Lynn McIntyre and Dr Beatrice Manel Dias-Wanigasekera, for their support and impacting their respective knowledge during my stay at Massey University. I especially wish to acknowledge Drs Dias-Wanigasekera and Mutukumira for soliciting funding from NZ King Salmon Ltd and FRST. It is also imperative that I accordingly acknowledge Dr Mutukumira for his guidance on the planning experiments of and analysis of data, sourcing for materials, managing the budget and overall social support during my stay at Albany campus. I am also very thankful to Dr McIntyre for nurturing me in phage science at ESR Laboratories in Christchurch, which now forms a foundation for my career. I also thank Dr Cruz for her expert guidance in conducting good microbiological experiments and ensuring that all analyses are accurate. My sincere thanks go to Rachel Liu and Jarod Young for providing technical support.

The following persons also played an important role in the course of my research work: Dr. S. On, ESR, Christchurch, for providing the three phages and a *L. monocytogenes* strain; Graham Fletcher at PFR Ltd; Professor P. Bremer at Otago University, Dunedin, for providing the *L. monocytogenes* strains; and Prof. Dr. Martin J. Loessner, Institute of Food Science and Nutrition, Zurich, Switzerland.

I express my sincere thanks to Richard Smith, (formerly with NZ King Salmon Limited); Karen Wittington and Neil Graham, NZ King Salmon Limited; Dr. Andrew Hudson, Dr Craig Billington, A. Premaratne, Dr. G. Greening, Anne Glennie at ESR Limited; Ron Fyfe, Anneke van Laanen and other staff at Cawthron Institute, Nelson; Graeme Fox and other staff at Sealord Group Ltd; Gavin Kouwenhoven, Future Cuisine; Leon Antunvich, Westpac Mussels Distributors Ltd and various other seafood processing plants in New Zealand for their technical support. My sincere thanks also goes to Dr. R. Lau, Dr. Andrew Cridge, Dr Marie Wong, Dr. Lara Matia-Merino, Eddie Stonehouse, Maryln Mabon, Margaret Allison, Margaret McNeill, Helne Mathew, Yan Wang, Sue Pearce, Dmitri Roukin, Rashmi Ramesh, Michael Bolech, Su Jung Lee, Michelle Tamehana, Anne-Marie Jackson, Jane Leighton, Carine Marais and all library staff, Sally Iwikau and other staff of Massey University; Adrian Turner and Carlos

Cheung, School of Biological Sciences, The University of Auckland, Stuart Blakeney, George Weston Foods Limited (Jasol), Auckland, NZ; Paul Harrison, Hawkins Watts, New Zealand; Mr Rasheed Iraninejad, Sifa Pty Ltd, Australia for various support during my study period.

I express my deep gratitude to Erika J. Lingohr, Research Public Health Agency of Canada; Emeritus Professor Andrew M. Kropinski, Queen's University; Professor Xiao-Lun Wu, University of Pittsburgh; Dr. Jochen Klumpp, Institute of Food, Nutrition and Health, Switzerland; Associate Professor Kalmokoff, Martin, Agriculture and Agri-Food, Canada; Ing-Nang Wang and Romain Gallet, University at Albany, NW; Prof. Alexander Sulakvelidze, University of Florida; Abraham Eisenstark, Cancer Research Centre, Columbia; Professor David A. Hodgson, University of Warwick; Assistant Professor Mark Liles, Auburn University, Alabama; Assistant Professor Laura Kasman, Department of Microbiology and Immunology, Charleston; Associate Professor Joseph D. Eifert, Department of Food Science & Technology Virginia Tech, Blacksburg, and Associate Professor Stephen Abedon, Ohio State University for their support with literature, and advice through discussion on the subject of bacteriophage.

I wish to extend my thanks to Dr. Champa Amarasiri and Nishahmini Karunasinghe who were in NARA, for guiding me to find a placement for my studies in New Zealand. I am ever so grateful to my friends and relatives: Krishathi Marambe, Pushpika Edirisinghe, Mahinda Jayaweera, Dayanthi Nugawela, Samantha Rathnayeka, Sujeewani Rathnayeka and all other friends who encouraged and made me feel at home while in New Zealand.

Abstract

Listeria monocytogenes is a major cause of illness, associated with seafood, therefore it is important to control this pathogen in seafood processing environments. Sporadic listeriosis outbreaks and seafood recalls indicate that current treatments to control this pathogen may be inadequate. The ability to adapt to harsh environmental conditions, develop resistance and form biofilms makes this environmental pathogen difficult to control using regular disinfectants. Bacteriophages (phages) could serve as effective alternative biocontrol agents. The main objective of this study was to isolate and characterize natural lytic Listeria specific phages and examine their effectiveness against L. monocytogenes under conditions mimicking those found in seafood processing environments.

Among a group of phages isolated from a seafood waste treatment unit, three phages (LiMN4L, LiMN4p and LiMN17) were selected based on plaque morphology and their source. The three phages were distinguished by morphology, efficiency of plating (EOP) in citrate agar and differences in EOP using different *L. monocytogenes* host strains. Three phages which were found as strictly virulent by whole genome sequence analysis, had broad host ranges at 15 °C and each phage also infected either *L. ivanovii* or *L. innocua*. These phages were unstable at 60 °C for 10 min suggesting psychrotrophic properties. The three phages showed low burst sizes indicating their potential suitability as passive biocontrol agents.

Low counts of *L. monocytogenes* strains (1909, 19DO3 and 19EO3) in late exponential phase, metabolically injured/stressed by heat and salt, lysed by the three phages at 15 °C in 30 min. The results suggested that the three virulent phages may be good candidates as biocontrol agents against *L. monocytogenes* under conditions commonly found in seafood processing plants.

The phages LiMN4L, LiMN4p and LiMN17, used as single phage or a cocktail of three phages, lysed cells adhered to stainless steel conditioned with soluble fish protein and on clean stainless steel coupons (SSC). The phage cocktail also eradicated low cell counts of about 2 log CFU/cm² adhered to SSC surfaces in the presence of fish proteins at 15 °C in 15 min and no re-growth of cells was observed from phage infected surfaces. This study suggested that a biofilm matrix shielded the bacterial cells from phage infection as three consecutive repeat applications of phages did not efficiently lyse undisturbed biofilm cells. Biofilm cells, once removed from the surface, showed

similar to sensitivity to that of exponential phase planktonic cells. Therefore, disruption of the biofilm structure may be required for effective phage treatments.

Phages suspended in phosphate buffered saline survived refrigeration for at least twelve months and were stable for at least 6 h under likely application conditions such as ambient temperatures and under fluorescence lighting. The three phages, either individually or as a cocktail, showed a high lytic efficacy indicating their potential to serve as bio-decontaminating agents in seafood processing environments.

Table of Contents

Acknowl	edgements	ii
Abstract		iv
Table of	Contents	vi
List of M	anuscripts	xi
List of Ta	ibles	xii
List of Fi	gures	xiv
Chapter 1	1: General Introduction	1
Referenc	es	5
Chapter 2	2: Literature Review	6
2.1	Introduction	6
2.2	Bacteriophage ecology	7
2.2.	Historical background of bacteriophages	7
2.2.2	2 Existence in nature	7
2.2.3	Behaviour of phages and interactions with bacteria	8
2.3	Different applications of phage and phage products	10
2.3.	1 Typing phage	10
2.3.2	Use of phages in rapid detection of <i>L. monocytogenes</i> in food	11
2.3.3	3 Lysins	11
2.3.4	4 Monocins	12
2.4	L. monocytogenes as a food-borne pathogen	13
2.4.	Prevalence of <i>L. monocytogenes</i>	13
2.4.2	Fish industry and food safety	14
2.4.3	Types of <i>Listeria monocytogenes</i> isolates linked to seafood contaminations	15
2.5	Isolation of phages from environments	17
2.5.	I Isolation of natural lytic phages from the environments	17
2.5.2	2 Impact of the protocol used for the phage isolation	18
2.5.3	3 Effect of temperature of phage isolation	19
2.6	Characterisation of phage isolates	23
2.6.	Different parameters to distinguish new phage isolates	23
2.7	Phage application strategies.	28
2.7.	Different approaches of phage application	28
2.7.2	2 Kinetic modelling of phage application	29

2.7.3	Commercial phage products for the food industry	30
2.7.4	Listeria phage-based biocontrol in food systems	31
2.7.5	Phage treatment of abiotic surfaces.	34
2.7.6	Effectiveness of phage in biofilms	35
2.7.7	Stability of phages	36
2.7.8	Development of phage resistant mutants	37
2.8 Con	clusion	38
References (C	Chapter 1 and 2)	39
	ost range and in vitro lysis of <i>Listeria monocytogenes</i> seafood isolates by	59
Abstract		60
3.1 Intr	oduction	61
3.2 Ma	terials and methods	63
3.2.1	L. monocytogenes strains and listeriaphages	63
3.2.2	Preparation of phage stocks	63
3.2.3	Host range of phages	64
3.2.4	In vitro lysis of L. monocytogenes by phage FWLLm3	65
3.2.5	Inactivation of low counts of <i>L. monocytogenes</i> by FWLLm3 over 24 h	65
3.2.6	Determination of adsorption rate constant (k value) of phage FWLLm3	65
3.2.7	Estimation of MOI _{input} and minimum phage FWLLm3 titre	66
3.2.8	Data analysis	67
3.3 Res	sults and discussion	67
3.3.1	Host range of three phages	67
3.3.2	In vitro lysis of L. monocytogenes cells by phage FWLLm3	72
3.3.3	The k value of phage FWLLm3	75
3.3.4	Estimation of MOI _{input} and minimum dose of phage FWLLm3	77
3.3.5	Conclusion	78
Authors' cont	ributions to the manucript	78
References		79
Chapter 4: Chenvironment	naracteristics of three listeriaphages isolated from New Zealand seafood	85
Abstract		86
4.1 Intr	oduction	87
4.2 Ma	terials and methods	87
4.2.1	Listeria strains and reference bacteriophage	88
4.2.2	Isolation and purification of phages from environmental samples	88
4.2.3	Plaque morphology at different gel concentrations.	90
	vii	

4.2	Preparation of purified high titre phage stocks	90
4.2	2.5 Efficiency of plating (EOP) of phages in citrate agar	90
4.2	2.6 Heat tolerance of phages at 50 and 60°C	91
4.2	2.7 Electron microscopy of phages	91
4.2	2.8 Restriction enzyme digestion of phage DNA	91
4.2	2.9 Host range of phages	91
4.2	2.10 Adsorption rate constant (k) of phages	92
4.2	Plaque formation temperature range of phages	92
4.2	2.12 Single-step replication (growth) of phages	93
4.2	2.13 Statistical analysis	93
4.3	Results	93
4.3	Isolation of listeriaphages based on plaque characteristics	93
4.3	EOP of phages in citrate agar	95
4.3	Heat treatment of phages at 50 and 60°C	95
4.3	8.4 Electron microscope images of phages	96
4.3	Phage DNA restriction analysis	98
4.3	Host ranges of the phages	100
4.3	The k values of phages	102
4.3	Plaque formation temperature range of phages	103
4.3	Single-step replication of phages	103
4.4	Discussion	104
4.5	Conclusion	107
Authors	s' contributions to the manuscript	108
Referen	nces	108
Support	ting Information	114
	\pm 5: Preliminary investigation of bacteriophage-lysis of physiologically stressed L .	
-	togenes in seafood processing environments	
	.t	
5.1	Introduction	
5.2	Materials and methods	
5.2	7 6 1 6	
5.2		
5.2	1	
5.2	<i>y</i>	
5.2		
5.2	Preparation and phage-based lysis of salt-stressed <i>L. monocytogenes</i>	125

5.2.7	Fluorescence microscopy of stressed <i>L. monocytogenes</i> cells stained with Acridine Orange	125
5.2.8	Phage- insensitive cells of high count <i>L. monocytogenes</i> culture	125
5.2.9	Determination of N-acetylglucosamine (GLcNAc) in cell wall of L.	
	monocytogenes	126
5.2.10	Statistical analysis	126
5.3 Re	esults and discussion	127
5.3.1	Phage-lysis of low count cells of <i>L. monocytogenes</i> in fish broth	127
5.3.2	Phage-mediated lysis of heat-injured L. monocytogenes	129
5.3.3	Phage-based lysis of starved L. monocytogenes cells	132
5.3.4	Phage-based lysis of salt-stressed L. monocytogenes	133
5.3.5	Phage-insensitive cells of exponential phase <i>L. monocytogenes</i> 19CO9 culture	. 135
5.3.6	Emergence of phage-resistant L. monocytogenes 19CO9 cells over 48 h	136
5.3.7	Presence of GLcNAc in the cell wall of <i>L. monocytogenes</i>	137
5.3.8	Conclusion	139
Authors' cor	ntributions to the manuscript	140
References		140
Chapter 6: E	Effectiveness of phages in the decontamination of Listeria monocytogenes adh	ered
to clean stair	nless steel, stainless steel coated with fish protein, and as a biofilm	145
Abstract		146
6.1 In	troduction	147
6.2 M	aterials and Methods	148
6.2.1	L. monocytogenes strains and Listeria phages	148
6.2.2	Test surfaces	148
6.2.3	Attachment of <i>L. monocytogenes</i> on surfaces	149
6.2.4	Phage-lysis of <i>L. monocytogenes</i> cells adhered on surfaces	150
6.2.5	Enumeration of cells adhered to surfaces.	150
6.2.6	Preparation of seven-day old <i>L. monocytogenes</i> biofilm on stainless steel coupon	150
6.2.7	Infection of seven-day old biofilm with <i>Listeria</i> phages	151
6.2.8	Preparation of dislodged seven-day old biofilm cells	151
6.2.9	Phage-lysis of dislodged seven-day old biofilm cells	152
6.2.10	Fluorescence microscopy of cells on surfaces	152
6.2.11	Data analysis	152
6.3 Re	esults	153
6.3.1	Adhesion of <i>L. monocytogenes</i> strains to FBSSC and SSC	153

6.3	.2 Phage-lysis of <i>L. monocytogenes</i> adhered to surfaces	155
6.3	.3 Phage-lysis of seven-day old biofilms on stainless steel	156
6.3	.4 Phage-lysis of dislodged seven-day old biofilm cells	156
6.4	Discussion	158
6.5	Conclusion	161
Authors	contributions to the manuscript	162
Referen	ces	162
Electron	ic supplementary material (ESM)	169
	7: Bacteriophages for the control of low levels of <i>Listeria monocytogenes</i> in a d fish processing environment	170
Abstract		171
7.1	Introduction	172
7.2	Materials and methods	175
7.2	.1 L. monocytogenes strains and Listeria phages	175
7.2	.2 Preparation of stainless steel coupon (SSC)	175
7.2	.3 Fish broth (FB)	176
7.2	.4 Phage-based lysis of low count cells on SSC	176
7.2	.5 Stability of <i>Listeria</i> phages under different ambient conditions	177
7.2	.6 Statistical analysis	179
7.3	Results	179
7.3	.1 Phage-lysis of low counts of <i>L monocytogenes</i> adhered on SSC	179
7. 3	3.2 Stability of <i>Listeria</i> phages under different conditions	181
7.4	Discussion	187
7.5	Conclusion	189
Authors	contributions to the manuscript	190
Referen	ces	190
Highligh	nts	195
Chapter	8: Summary and future directions of study	196
8.1	Summary	196
8.2	Future directions of study	198
Append	x 1	200

List of Manuscripts

The chapters 3, 4, 5, 6 and 7 were presented as manuscripts. Each manuscript has been formatted according to the specifications of each journal.

Table 1. List of manuscripts

Chapter number	Title of manuscript (Journal)			
3	Host range and in vitro lysis of Listeria monocytogenes seafood			
	isolates by bacteriophages (Submitted to Food Science and			
	Technology International)			
4	Characteristics of three listeriaphages isolated from New Zealand			
	seafood environment (Submitted to Journal of Applied			
	Microbiology)			
5	Preliminary investigation of bacteriophages mediated lysis of			
	physiologically stressed <i>L. monocytogenes</i> (To be submitted to Food			
	Science and Technology International)			
6	Effectiveness of phages in the decontamination of Listeria			
	monocytogenes adhered to clean stainless steel, stainless steel coated			
	with fish protein, and as a biofilm (Submitted to Journal of Industrial			
	Microbiology & Biotechnology)			
7	Bacteriophages for the control of low levels of Listeria			
	monocytogenes in a simulated fish processing environment (To be			
	submitted to International Journal of Food Microbiology)			

The following three manuscripts published with some amendments are included in the Appendix 1.

- 1. Host range and *in vitro* lysis of *Listeria monocytogenes* seafood isolates by bacteriophages (Submitted to Food Science and Technology International)
- 2. Characteristics of three listeriaphages isolated from New Zealand seafood environment (Submitted to Journal of Applied Microbiology)
- 3. Effectiveness of phages in the decontamination of *Listeria monocytogenes* adhered to clean stainless steel, stainless steel coated with fish protein, and as a biofilm (Submitted to Journal of Industrial Microbiology & Biotechnology)

List of Tables

Table 1 List of manuscripts	xi
Table 2.1 Characteristics of lytic <i>Listeria</i> phages belonging to the order <i>Caudovirale</i> and families <i>Myoviridae</i> and <i>Siphoviridae</i>	
Table 3.1 Host ranges of phages FWLLm1, FWLLm3 and FWLLm5 across 36 <i>L. monocytogenes</i> strains isolated from fish, mussel and environmental swabs	70
Table 3. 2 Host ranges of listeriaphages FWLLm1, FWLLm3 and FWLLm5 using 14 <i>L. monocytogenes</i> strains isolated from one salmon processing plant	71
Table 3.3 Relative efficiency of plating (%EOP) of listeriaphages FWLLm1, FWLLm3 and FWLLm5 on exponential phase <i>L. monocytogenes</i> strains	72
Table 3.4 Estimates of input multiplicity of infection (MOI _{input}) and minimum practical (input) listeriaphage FWLLm3 titres (doses)	77
Table 4.1 Dimensions of listeriaphage LiMN4L, LiMN4p and LiMN17 determined on TEM images	98
Table 4.2 Percentage efficiency of plating (% EOP) of listeriaphages LiMN4L, LiMN4p and LiMN17	. 101
Table 4.3 Adsorption rate constants (k) for phages LiMN4L, LiMN4p and LiMN17 with different <i>L. monocytogenes</i> strains	. 103
Table 4.4 Single-step replication kinetics of listeriaphages LiMN4L, LiMN4p and LiMN17 grown with <i>L. monocytogenes</i> 19CO9	. 104
Table 4.S1 Listeriaphage (LiMN4L, LiMN4p and LiMN17) plaques on lawn of <i>L. monocytogenes</i> 19CO9 on trypticase soy yeast extract double layer agar	. 114
Table 4.S2 Percentage survival of listeriaphages LiMN4L, LiMN4p and LiMN17 in lysates	114

Table 4.S3 Percentage efficiency of plating (%EOP) and formation of lytic zones (LZ) by listeriaphages LiMN4L, LiMN4p and LiMN17 on <i>L. monocytogenes</i> 19CO9 115
Table 5.1 <i>L. monocytogenes</i> strains assayed with fluorescently labeled wheat germ agglutinin
Table 6.1 Initial counts (mean \pm standard error) of <i>L. monocytogenes</i> strains in 1/10 (v/v) trypticase soy broth (cell adhesion bath), on the surfaces
Table 6.2 Cells adhered on fish broth based conditioned coupons (FBSSC) and stainless steel coupons (SSC) with respect to the mean cell counts on the control coupons after infection with phages LiMN4L, LiMN4p, LiMN17 and three-phage cocktail

List of Figures

Figure 3.1 Host ranges of three listeriaphages were determined by the formation of either lytic zones or discreet plaques on host lawns
Figure 3.2 Viable cell counts of <i>L. monocytogenes</i> after infection with a series of listeriaphage FWLLm3 titres
Figure 3.3 Viable cell counts of <i>L. monocytogenes</i> after infection with ~8.7 log ₁₀ PFU/ml of listeriaphage FWLLm3 in trypticase soy broth
Figure 3.4 The adsorption kinetics of phage FWLLm3 on <i>L. monocytogenes</i> 18GO1, 19EO1, 19EO3 and 22BO7
Figure 4.1 Listeriaphage plaques on trypticase soy yeast extract double layer agar (0.1% agarose overlay and 1% agar base)
Figure 4.2 Efficiency of plating of listeriaphages LiMN4L, LiMN4p and LiMN1796
Figure 4.3 Transmission Electron Microscope images of listeriaphages
Figure 4.4 Listeriaphage DNA profiles on 0.7% agarose gel. Enzyme-digested (ClaI) profiles of phage DNA with λ HindIII marker
Figure 4.5 Enzyme-digested (SacI) profiles of phage DNA with λ HindIII marker99
Figure 4.S1 Single-step growth curves of phages LiMN4L
Figure 4.S2 Single-step growth curves of phages LiMN4p at 15, 25 and 35°C 117
Figure 4.S3 Single-step growth curves of phages LiMN17
Figure 5.1 Viable counts of phage infected and control cultures of late exponential phase three <i>L. monocytogenes</i> strains.
Figure 5.2 Viable counts of control and phage-infected cultures of heat-treated <i>L. monocytogenes</i> strains
Figure 5.3 Coccoid-form cells of <i>L. monocytogenes</i> strains starved in MilliQ water.
Figure 5.4 Viable counts of control and phage-infected cultures of starved <i>L</i> .

Figure 5.5 Stressed cells of <i>L monocytogenes</i> strains in 9% (w/v) NaCl
Figure 5.6 Viable counts of control and phage-infected cultures of salt-stressed <i>L. monocytogenes</i> strains
Figure 5.7 Viable counts of exponential phase <i>L. monocytogenes</i> 19CO9 during reinfection with phages LiMN4L, LiMN4p and LiMN17
Figure 5.8 Viable counts of <i>L. monocytogenes</i> 19CO9 after exposure to phages LiMN4L, LiMN4p and LiMN17
Figure 5.9 <i>L. monocytogenes</i> 19CO9 cells stained with fluorescently labelled wheat germ agglutinin (Alexa Fluor 488 WGA)
Figure 6.1 Fluorescent microscopy images of a cell-mix of three <i>L. monocytogenes</i> strains (19CO9, 19DO3 and 19EO3) cocktail adhered to stainless steel
Figure 6.2 Counts of a three strain mix of a seven-day old <i>L. monocytogenes</i> (19CO9, 19DO3 and 19EO3) biofilm on stainless steel coupon after three phage applications. 157
Figure 6.3 Counts of dislodged seven-day old biofilm cell cultures of individual <i>L. monocytogenes</i> strains (19CO9, 19DO3, 19EO3) after infection with individual phages.
ESM Figure 6.1. Degradation of seven-day biofilm of mixed cells of <i>L. monocytogenes</i> strains
Figure 7.1 Viable counts of a mixture of three <i>L. monocytogenes</i> strains (19CO9, 19DO3 and 19EO3) adhered with 10% fish proteins broth on stainless steel
Figure 7.2 Viable counts of a mixture of three <i>L. monocytogenes</i> strains adhered with 10% fish protein broth on stainless steel
Figure 7.3 Titre of <i>Listeria</i> phage LiMN4L in QAC exposed at 15±1 °C
Figure 7.4 Titre of listeriaphage LiMN4p in QAC exposed at 15 °C
Figure 7.5 Titre of listeriaphage LiMN17 in QAC exposed at 15 °C
Figure 7.6 Titre of three <i>Listeria</i> phages in fish broth (FB) and phosphate buffered saline (PBS) at 15±1 °C for 24 h
Figure 7.7 Titre of three <i>Listeria</i> phages in trypticase sov broth at pH 3-12

Figure 7.8 Surviving titres of three <i>Listeria</i> phages in low (487±30 lux) and high (1710±130 lux) intensity light	186
Figure 7.9 Surviving titres of three <i>Listeria</i> phages in the control (suspension) and attached on the stainless steel coupon	186
Figure 7.10 Surviving titres of three <i>Listeria</i> phages in phosphate buffered saline	187