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Identification and characterization of an 8.4 kDa protein antigen of *Mycobacterium bovis*.

A thesis presented in partial fulfilment of the requirements for the degree of

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Alexander Dougal McLachlan

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ABSTRACT.

The culture filtrate (CF) derived from a *M. smegmatis* subclone transformed with the mycobacteria/*E. coli* plasmid shuttle vector pSU4511 containing a 4.3 kb fragment of *M. bovis* DNA (*M. smegmatis* pSU151.43), was observed to stimulate PBMC from a steer vaccinated with *M. bovis* BCG to proliferate and produce IFN-γ. To identify the source of immunoreactivity, the proteins in CF derived from *M. smegmatis* pSU151.43 were separated by fast protein liquid chromotography (FPLC) and the fractions were screened in whole blood IFN-γ assays. A stimulatory protein was purified that had a molecular mass of 8335 Da and the N-terminal amino acid sequence: DPVDAVINTT. Polyclonal antisera were raised against the purified recombinant antigen in rabbits and used for Western blotting.

The nucleotide sequence of the 4.3 kb insert of *M. bovis* DNA was determined, and the open reading frame (ORF) coding for the 8.4 kDa protein was identified. Computer analysis of the deduced amino acid sequence with the programme PSORT predicted that the nascent protein consisted of a 28 amino acid export signal sequence followed by an 82 amino acid mature protein. It was also found that *M. avium* possesses a nucleotide sequence that potentially codes for a protein with a high degree of homology to the 8.4 kDa antigen of *M. bovis*.

A segment of the 4.3 kb insert of *M. bovis* DNA adjacent to the gene coding for the 8.4 kDa antigen was found to be polymorphic between the strain of *M. bovis* from which the cosmid library was constructed and the published sequence of *M. tuberculosis* H37Rv (Cole *et al.* 1998). The *M. bovis* sequence contained 1.7 copies of a 62 bp exact tandem repeat and the *M. tuberculosis* sequence contained 2.7 copies. The species distribution of the 62 bp exact tandem repeat (ETR) locus was characterized by polymerase chain reaction (PCR) and Southern blotting. The 62 bp ETR was found to occur only in *M. tuberculosis* complex species and may be a useful genetic marker for differentiating between *M. bovis* and *M. tuberculosis*.

Lymphocyte proliferation and IFN- γ assays were used to measure the responses of ten BCG vaccinated and ten unvaccinated calves the 8.4 kDa antigen, PPD-B and PPD-A tuberculins, both before and after intratracheal challenge infection with virulent M. bovis.

The results provided evidence that vaccination of cattle with *M. bovis* BCG but not infection with *M. bovis* appeared to elicit an immune response to the 8.4 kDa antigen of *M. bovis*.

To obtain greater quantities of recombinant 8.4 kDa antigen, the gene that codes for the protein was cloned into $E.\ coli$ and $M.\ smegmatis$ expression plasmids. The 8.4 kDa antigen was overexpressed and secreted with an N-terminal 6 x Histidine tag by $M.\ smegmatis$. Approximately 500 µg of 6 x Histidine tagged 8.4 kDa Ag were purified / litre of CF in one step by metal chelate affinity chromatography. The recombinant protein was shown to elicit specific IFN- γ responses $in\ vitro$.

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Much of the work reported in this thesis was only made possible by the assistance of collaborators and commercial service providers. The nucleotide sequence of the 4.3 kb insert of *M. bovis* and other DNA sequences was determined by Dr. Kathryn Stowell and Lorraine Berry at the Massey University DNA Analysis Service. The species distribution of the 62 bp ETR locus was investigated in many more isolates of mycobacteria than available at Massey University by Dr. Cristina Gutierrez at the Pasteur Institute, Paris. Mass spectrometry and N-terminal sequencing was performed by Dr. Gill Norris and Mr. Trevor Loo at Massey University MasSpec, and Protein Sequencing Services. The experimental animals used at Massey University were grazed at the Large Animal Teaching Unit, and thanks must be extended to Mr. Robin Whitson and Odine Johnstone for their willing assistance with blood sampling.

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ABBREVIATIONS.

2D-PAGE two dimensional polyacrylamide gel electrophoresis.

2-ME 2-mercaptoethanol.

2xSLB 2 x sample loading buffer.

6 x His 6 x Histidine.

A₂₈₀ absorbance at 280 nm.

Ag antigen.

AHB Animal Health Board.

APC antigen presenting cell.

APS ammonium persulfate.

ATCC American Type Culture Collection.

BCG M. bovis bacillus Calmette-Guérin.

BLAST basic local alignment search tool.

bp nucleotide base pairs
BSA bovine serum albumin.
CCT comparative cervical test.
CD cluster of differentiation.

CF culture filtrate.

Cos151 *M. smegmatis* cosmid library clone 151.

ConA concanavalin A.

dH₂O double distilled water. c.p.m counts per minute.

 Δ c.p.m difference in counts per minute.

 $\Delta \text{ OD}_{450}$ difference in optical density at 450 nm.

DAB 3, 3'-Diaminobenzidine.

DIG-dUTP Digoxigenin-11-2'-deoxy-uridine-5'-triphosphate.

DMSO dimethyl sulfoxide.

DNA deoxyribonucleic acid.

dsDNA double stranded DNA.

dNTP deoxynucleoside triphosphate.

DR direct repeat.

DTH delayed type hypersensitivity.

DTT dithiothreitol.

EIA enzyme immunoassay.

ELISA enzyme linked immunosorbent assay.

ERMA Environmental Risk Management Authority.

ETR exact tandem repeat.

FAO Food and Agriculture Organization of the United Nations.

FCS foetal calf serum.

FPLC fast protein liquid chromatography.

g gravity (a force of ~ 10 N).
GST glutathione-S-transferase.
HRP horesradish peroxidase.

ICAM intercellular adhesion molecule.
IFA Incomplete Freund's Adjuvant.

IFN interferon.

IFN-γ interferon gamma.

IL interleukin.

IPTG isopropylthio- β -galactoside.

IS insertion sequence.
IU international units.

IUATLD International Union Against Tuberculosis and Lung Disease.

IUPAC International Union of Pure and Applied Chemists.

IWGMT International Working Group on Mycobacterial Taxonomy.

kb kilobase pairs. LB Lauria-Bertani.

LøP lymphocyte proliferation.

MHC major histocompatability complex.

MIRU mycobacterial interspersed repetitive unit.

MPTR major polymorphic tandem repeat.

MW molecular weight.

MWCO molecular weight cut-off.

NCBI National Center for Biotechnology Information.

NK natural killer T-lymphocyte.

NVL no visible lesions.
OD optical density.

OIE Office International des Epizooties.

ORF open reading frame.

PBMC peripheral blood mononuclear cells.

PBS phosphate buffered saline.
PCR polymerase chain reaction.

PGRS polymorphic GC rich repetitive sequence.

PMSF phenylmethylsulfonyl flouride.

PO₄SB phosphate start buffer.
PO₄SB phosphate wash buffer.
PPD purified protein deriviative.
PPD-A PPD derived from *M. avium*.
PPD-B PPD derived from *M. bovis*.

RFLP restriction fragment length polymorphism.

RNA ribosomal nucleic acid. r.p.m revolutions per minute.

SDS-PAGE sodium dodecyl sulphate - polyacrylamide electrophoresis.

SIT single intradermal test.

TAE Tris-acetate.

TB complex *Mycobacterium tuberculosis* complex.

TBE Tris-borate.

TIGR The Institute for Genomic Research.

TEMED N, N, N', N'-tetramethylethylenediamine.
Th 1/Th 2 T-helper cell phenotype Type 1/Type 2.

TNF tumour necrosis factor.

TTBS Tween Tris-buffered saline.

U units.
UV ultraviolet.
V volts.

VNTR variable number of tandem repeats.

WHO World Health Organization.

w/v weight for volume.

X-Gal 5-bromo-4-chloro-3-indoyl-β-D-galactoside.