

Copyright is owned by the Author of the thesis. Permission is given for a copy to be downloaded by an individual for the purpose of research and private study only. The thesis may not be reproduced elsewhere without the permission of the Author.

**The Secreted Aspartic Proteinases of *Candida albicans***

A thesis submitted in partial fulfillment of the requirements for the degree of Doctor  
of Philosophy in the Institute of Molecular BioSciences at  
Massey University, New Zealand

**Michelle E. Sullivan**

October 1999

To my parents

# ACKNOWLEDGEMENTS

---

I would like to thank Professor Pat Sullivan, not only for providing an excellent lab environment, but for his wisdom and his unfailing enthusiasm throughout this project. Thanks also to my second supervisor Associate Professor John Tweedie, for his timely support and advice, and for always having an open door.

I would also like to thank all the past and present members of the Mainland lab who not only contributed advice and support, but who gave the lab spirit and made it a great place to work; Rochelle Ramsay who taught me the effectiveness of the 'hands on hips' approach, and who will always be a friend (but I'm not scared of you, even if the whippersnappers are!); Rebecca Sanders for fashion advice, friendship and fun; Linda Giblin for handy shortcuts and raucous Irish laughter; Donna Cooper for her endless kindness; and to Paul McJarrow for always taking the time to explain about tricky carbohydrates!

Thanks also to all the members of the institute who provided advice and inspiration throughout - particularly Catherine Day for her encouragement and direction when everything looked pear-shaped, Carolyn Young for cloning hints and tips, Rachel Page for hot-room help, and Carol Flyger for seeds and gardening advice. For technical support I would like to thank Gill Norris, Diana Carne, Liz Nickless, the staff at SAPU, and of course the ever-present David McSweeney for all the things that no-one else knew about.

Special thanks must go to Liz Hutton who always believed in me, even when I didn't believe in myself, and who will always be a source of inspiration. I hope you rest in peace my darling friend. Thanks also to my parents for their constant love and friendship. Lastly, I would like to thank Shaun Lott, for love, patience, kindness and for making me happier than I ever dreamed I could be.

# ABSTRACT

---

The common human fungal pathogen, *Candida albicans*, possesses at least nine genes encoding secreted aspartic proteinases (*SAPs*). Saps are widely regarded as virulence factors, despite historical controversy surrounding their actual roles in the onset and development of candidosis. While Sap1, Sap2 and Sap3 had been previously studied at the biochemical level, Sap4 and Sap5 had not been detected, purified or characterised. To facilitate analysis of the proteins, *SAP4* and *SAP5* were amplified by PCR and cloned. The nucleotide changes in *SAP4* were silent, and *SAP5* contained one conserved amino acid substitution, compared with the published sequences. The methylotrophic yeast *Pichia pastoris* was used as a host for heterologous expression of *SAP4* and *SAP5*, and the respective proteins were purified to homogeneity. Purification of Sap4 involved Mono Q anion exchange chromatography at pH 7.0, while Sap5 was purified by cation exchange chromatography on Resource S at pH 7.0. *C. albicans SAP1* was also over-expressed in *P. pastoris* as a control. Biochemical analysis of the recombinant proteins revealed that Sap4 and Sap5 were optimally active at pH 4.5, 1-2 pH units higher than the optima of Saps1, 2 and 3. At optimum pH, the specific activities of Sap4 and Sap5 were 239 and 33  $\mu\text{g}$  tyrosine equivalents/min, respectively. These isoenzymes also retained significant activity at pH 7.0, which suggested roles in the disease process at host sites of neutral pH. Sap4 and Sap5 showed decreased specific activity towards denatured globin, and increased specific activity towards a fluorocasein substrate, when compared with Saps1-3. Substrate specificity analyses (performed using a peptide substrate, glucagon, and MALDI-TOF of the purified peptide fragments), showed that Sap4 and Sap5 hydrolysed the glucagon at the same sites, but the analysis did not reveal a consensus cleavage sequence. The deduced masses of Sap1, Sap4 and Sap5 were 36,179, 36,995 and 37,256, respectively. However, ES-MS indicated the masses of the recombinant Sap1 and Sap4 were larger than expected, by 2232 and 2041 respectively. Glycopeptide fragment ion analysis suggested the additional mass was due to attached sugar residues. Carbohydrate chromatography confirmed the presence of mannose and N-acetyl glucosamine. The presence of N-acetyl glucosamine species, and the lack of consensus *N*-linked glycosylation sites in the Sap1, Sap4 and Sap5 proteins

suggests a novel pattern of *O*-linked glycosylation in *P. pastoris*. The purified enzymes were subjected to crystallisation trials and some promising crystals were produced. Previous studies showed that *SAP4*, *5* and *6* are expressed during serum-induced germ-tube formation, but this was not confirmed in this study.

# TABLE OF CONTENTS

---

Abstract.....	i
Table of contents.....	iii
List of figures.....	ix
List of tables.....	xii
Amino acid abbreviations and colour coding.....	xiii
Abbreviations.....	xiv
<b>Chapter 1.0 Introduction.....</b>	<b>1</b>
1.1 <i>Candida albicans</i> .....	1
1.2 Pathogenicity.....	3
1.2.1 Adhesion.....	4
1.2.2 Cell wall.....	4
1.2.3 Polymorphism.....	5
1.2.4 Phenotypic variation.....	6
1.2.5 Secreted hydrolases.....	6
1.2.6 Hydrophobicity.....	7
1.2.7 Signal transduction networks.....	8
1.3 Other <i>Candida</i> species.....	9
1.4 <i>C. albicans</i> and host immunity.....	9
1.5 Antifungal therapies.....	10
1.6 Phenotypic switching.....	12
1.7 Molecular mechanisms of phenotypic switching.....	14
1.8 Molecular mechanisms of polymorphism.....	17
1.9 Aspartic proteinases.....	21
1.9.1 Evolution.....	21
1.9.2 Substrate specificity.....	22
1.9.3 Structural features.....	23
1.9.4 Catalytic mechanism.....	23
1.10 Retroviral aspartic proteinases.....	24
1.11 Fungal aspartic proteinases.....	25
1.12 Secreted aspartic proteinases.....	26
1.13 Secretion of Sap.....	27
1.14 Regulation of Sap.....	28
1.15 Sap and pathogenicity.....	32
1.16 Sap4, Sap5 and Sap6.....	35
1.17 Sap structure.....	38
1.18 Project aims.....	39

<b>2.0 Materials and Methods</b> .....	<b>40</b>
2.1 Enzymes .....	40
2.2 Chemicals .....	40
2.3 Miscellaneous products .....	41
2.4 <i>Escherichia coli</i> phenotypes .....	41
2.5 Yeast strains.....	42
2.6 Plasmids utilised during this study .....	42
2.7 Plasmids developed during this study.....	42
2.8 Growth media .....	43
2.8.1 Yeast media .....	43
2.8.2 Bacterial media.....	43
2.8.3 General buffers .....	44
2.9 General methods.....	44
2.9.1 Restriction endonuclease DNA digestion.....	44
2.9.2 Agarose gel electrophoresis of DNA .....	44
2.9.3 DNA ligations.....	44
2.9.4 Creating blunt-ended DNA.....	45
2.9.5 Preparation and transformation of <i>E. coli</i> competent cells .....	45
2.9.5.1 Preparation .....	45
2.9.5.2 Transformation .....	45
2.9.6 Selection of transformants .....	46
2.9.7 Plasmid DNA preparation – small scale .....	46
2.9.8 Plasmid DNA preparation – large scale.....	46
2.9.9 DNA purification from a TAE agarose gel.....	46
2.9.10 Genomic DNA purification from <i>C. albicans</i> .....	47
2.9.11 PCR.....	47
2.9.12 Southern blotting and hybridisation analyses .....	47
2.9.13 Radiolabelling DNA .....	48
2.9.14 Total RNA extraction from <i>C. albicans</i> .....	48
2.9.15 Agarose gel electrophoresis of RNA .....	48
2.9.16 Northern blotting and hybridisation analyses .....	49
2.9.17 Oligonucleotide synthesis .....	49
2.9.18 Dideoxy chain termination sequencing.....	49
2.10 Yeast and protein methods.....	49
2.10.1 Preparation and transformation of yeast spheroplasts.....	49
2.10.2 Screening yeast transformants .....	50
2.10.3 <i>P. pastoris</i> growth and induction.....	50
2.10.4 TCA precipitation of protein samples.....	51
2.10.5 SDS-PAGE analysis .....	51
2.10.6 Native PAGE analysis .....	51

2.10.7	Western blotting of proteins .....	51
2.10.8	N-terminal amino acid sequence analysis of proteins.....	52
2.10.9	Sap enzyme assays.....	53
2.10.9.1	Azocollagen assay .....	53
2.10.9.2	BSA-based assay .....	53
2.10.9.3	Globin-based assay .....	53
2.10.9.4	Fluorocasein-based assay .....	54
2.10.10	Substrate specificity of Sap4 and Sap5.....	54
2.10.11	Column chromatography .....	55
2.10.12	Carbohydrate cleavage.....	55
2.10.12.1	$\beta$ -elimination reaction.....	55
2.10.12.2	Acid hydrolysis.....	56
2.10.13	DIG Glycan/protein analysis .....	56
2.10.13.1	Labelling.....	56
2.10.13.2	Detection .....	56
2.11	Antibody production .....	57
2.12	Crystallisation trials .....	57
2.13	Microscopy .....	58
2.14	Computing programs .....	58
<b>3.0</b>	<b>Cloning and Vector Construction.....</b>	<b>59</b>
3.1	Introduction .....	59
3.2	PCR amplification of <i>SAP4</i> , <i>SAP5</i> and <i>SAP6</i> .....	60
3.3	Optimising PCR amplification of <i>SAPs</i> .....	61
3.4	Restriction endonuclease analysis of PCR products .....	61
3.5	Analysis of <i>SAP4</i> and <i>SAP5</i> clones .....	66
3.6	DNA sequence analysis of <i>SAP4</i> and <i>SAP5</i> clones .....	70
3.7	Construction of heterologous expression vectors for Sap4 and Sap5.....	75
3.8	Construction of a Sap1 heterologous expression vector .....	78
<b>4.0</b>	<b>Transformation and Expression .....</b>	<b>80</b>
4.1	Introduction .....	80
4.2	Transformation of KM71 with heterologous expression constructs.....	81
4.3	Selection of KM71 transformants.....	81
4.4	Confirmation of plasmid integration.....	83
4.5	Preliminary expression of recombinant Sap proteins .....	83
4.6	Identification of the recombinant proteins.....	90
4.6.1	Identification by western blotting .....	90
4.6.2	Identification by enzyme assay.....	90
4.7	Large scale expression of Sap1, Sap4 and Sap5 .....	92

4.7.1	Optimising strain storage conditions .....	92
4.7.2	Optimising starter culture conditions for Sap expression .....	95
4.7.3	Effect of methanol concentration on Sap expression.....	95
4.7.4	Effect of added buffer on Sap expression .....	95
4.7.5	Effect of induction culture size on Sap expression .....	97
4.7.6	Effect of culture aeration on Sap expression .....	97
4.8	Fermentation scale induction of Sap.....	99
4.9	Development of a new system for Sap expression .....	100
4.10	<i>E. coli</i> expression of Sap4 .....	102
4.10.1	PCR primer design.....	102
4.10.2	Transformation of host strains .....	107
4.10.3	Expression of recombinant Sap4 .....	107
<b>5.0 Purification of Sap .....</b>		<b>108</b>
5.1	Introduction .....	108
5.2	Purification of recombinant Sap1 .....	109
5.3	Purification of recombinant Sap4 .....	110
5.3.1	Anion exchange chromatography at pH 6.0 .....	110
5.3.2	Cation exchange chromatography at pH 6.0.....	113
5.3.3	Anion exchange chromatography at pH 7.0 .....	113
5.3.4	Anion exchange chromatography at pH 7.0, low ionic strength.....	114
5.3.5	Optimising the anion exchange gradient.....	114
5.3.6	Batchwise purification trials with DEAE sepharose.....	116
5.3.7	DEAE sepharose column chromatography.....	117
5.4	Large-scale purification of mature Sap4.....	118
5.5	Purification of recombinant Sap5 .....	124
5.5.1	Anion exchange chromatography .....	124
5.5.2	Cation exchange chromatography .....	124
5.5.3	Gel filtration of Sap5 .....	125
5.6	Sap 5 purification protocol .....	125
<b>6.0 Biochemical Characterisation of Recombinant Sap Isoenzymes.....</b>		<b>126</b>
6.1	Introduction .....	126
6.2	Sap enzyme activity assays.....	127
6.2.1	Sap4 substrate preference .....	127
6.2.2	Enzyme kinetics.....	127
6.2.2.1	Globin assay .....	127
6.2.2.2	Fluorocasein assay.....	129
6.3	pH optima of the Sap isoenzymes.....	131
6.4	Specific activity of the Sap isoenzymes .....	133

6.5	Substrate specificity of Sap .....	135
6.6	Thermal stability of the Sap isoenzymes .....	143
6.7	SDS-PAGE analysis of the Saps1-5 .....	144
6.8	Native PAGE analysis of Saps1-5 .....	144
6.9	Autodegradation of Sap isoenzymes.....	146
6.10	Pepstatin inhibition of Sap.....	146
6.11	Glycosylation of Sap .....	149
	6.11.1 Electrospray mass spectroscopy .....	149
	6.11.2 FACE analysis of Sap.....	153
	6.11.3 Carbohydrate chromatography .....	155
6.12	Interaction of recombinant Saps with SQAPI.....	162
6.13	Antibody production.....	164
6.14	Production of anti-Sap4 .....	164
6.15	Purification of the IgG fraction.....	165
6.16	Titre of anti-Sap4.....	165
	6.16.1 Comparison of anti-Sap2 and anti-Sap4 .....	165
	6.16.2 Anti-Sap4 titre .....	167
6.17	Specificity of anti-Sap4 .....	167
<b>7.0</b>	<b>Expression of Sap .....</b>	<b>171</b>
7.1	Introduction .....	171
7.2	Expression of <i>SAPs 4, 5 and 6</i> .....	172
7.3	Northern analysis of <i>SAP4-6</i> expression: strains ATCC 10231 and A72 .....	176
7.4	Northern analysis of <i>SAP4-6</i> expression: strain ATCC 10261 .....	179
7.5	<i>In vitro</i> expression analyses.....	181
<b>8.0</b>	<b>Crystallisation trials.....</b>	<b>183</b>
8.1	Introduction .....	183
8.2	Crystallisation of Sap1.....	184
8.3	Crystallisation of Sap4.....	188
8.4	Crystallisation of Sap5.....	189
8.5	Homology modelling of Saps 1, 4 and 5 .....	193
<b>9.0</b>	<b>Discussion.....</b>	<b>198</b>
9.1	Future work .....	208
<b>10.0</b>	<b>Appendix.....</b>	<b>210</b>
10.1	<i>Sequence alignment of SAPI-SAP9 of C. albicans</i> .....	211
10.2	Dionex analysis of Sap isoenzymes.....	214

10.2.1	Dionex calculations .....	214
10.2.2	Dionex analysis of Sap5 carbohydrate moieties .....	215
10.3	Purification and analysis of Sap5 .....	216
10.3.1	Purification of Sap5 .....	216
10.3.2	Anti Sap5 antiserum .....	218
10.4	ESMS data .....	219
10.5	MALDI-TOF analysis of glucagon peptide fragments .....	220
<b>11.0</b>	<b>References .....</b>	<b>224</b>

# LIST OF FIGURES

---

1.1	Alternative morphologies of <i>C. albicans</i> .....	2
1.2	Organisation of <i>C. albicans</i> cell wall.....	5
1.3	Variant colony phenotypes in the switching repertoire of WO-1 .....	14
1.4	A model for the regulation of white- and opaque-phase specific genes.....	16
1.5	A model for the regulation of pseudohyphal growth in <i>C. albicans</i> .....	20
1.6	Schematic representation of the substrate binding cleft of an aspartic proteinase .....	22
1.7	Expression of the <i>SAP</i> gene family in <i>C. albicans</i> .....	31
3.1	Protocol for generating <i>SAP</i> clones .....	62
3.2	Optimised PCR amplification of <i>SAP4</i> , <i>SAP5</i> and <i>SAP6</i> .....	64
3.3	Schematic of <i>SAP4</i> , <i>5</i> & <i>6</i> showing relative positions of PCR primers and restriction sites..	65
3.4	Restriction endonuclease analysis of <i>SAPs</i> 4, 5 and 6 PCR products .....	67
3.5	Restriction digest analysis of <i>SAP4</i> and <i>SAP5</i> sequences cloned into pBluescript.....	69
3.6	Nucleotide sequence of <i>SAP4</i> from <i>C. albicans</i> strain ATCC 10261 .....	71
3.7	Nucleotide sequence of <i>SAP5</i> from <i>C. albicans</i> strain ATCC 10261 .....	72
3.8	Alignment of the sequence containing the amino acid substitutions in Sap5 with other members of the <i>C. albicans</i> Sap family .....	74
3.9	Schematic of expression vector construction.....	76
3.10	Screening pPIC3K clones for the correct orientation of <i>SAP4</i> and <i>SAP5</i> genes.....	77
3.11	Schematic of the <i>SAP1</i> cloning strategy .....	79
4.1	G418 screen of KM71 transformants.....	84
4.2	Schematic representation of pMS1 and pMS5 plasmids integrated into the KM71 genome..	85
4.3	Southern blot of <i>P. pastoris</i> (KM71) transformants MS4A and MS5A, containing pMS4 and pMS5.....	86
4.4	SDS-PAGE of preliminary Sap1, Sap4 and Sap5 expression trials .....	88
4.5	N-terminal sequence of recombinant Sap proteins .....	89
4.6	Western analysis of Sap expression .....	91
4.7	Comparative induction of Sap4 and Bgl2.....	93
4.8	Effect of storage medium on Sap expression .....	94
4.9	Effect of starter culture medium on Sap expression .....	94
4.10	Effect of methanol concentration on Sap expression.....	96
4.11	Effect of buffer on Sap expression.....	96
4.12	Effect of induction culture volume on Sap expression.....	98
4.13	Effect of culture aeration on Sap expression.....	98
4.14	Sparger system for induction of recombinant Sap .....	101
4.15	PCR primers for <i>E. coli</i> expression.....	103
4.16	Strategy for cloning <i>SAP4</i> into pRSETa, an <i>E. coli</i> expression vector .....	104

4.17	SDS-PAGE and western blot analysis of <i>E. coli</i> expression of Sap4 .....	107
5.1	Purification of Sap1 using ion-exchange chromatography .....	111
5.2	SDS-PAGE of Sap1 fractions from ion exchange chromatography .....	112
5.3	SDS-PAGE of Sap4 fractions from ion exchange chromatography (pH 7.0).....	112
5.4	SDS-PAGE of Sap4 fractions from anion exchange chromatography at low ionic strength (pH 7.0).....	115
5.5	Separation of the misprocessed species of Sap4 using Resource Q ion exchange chromatography .....	115
5.6	SDS-PAGE of Sap4 fractions from optimised anion exchange gradient.....	116
5.7	Batchwise DEAE Sepharose purification trial results.....	117
5.8	DEAE Sepharose chromatography at pH 7.0.....	119
5.9	Graph showing protein and Sap activity fractions from the DEAE Sepharose column.....	121
5.10	SDS-PAGE analysis of fractions from the DEAE Sepharose column .....	121
5.11	h.p.l.c. trace of Mono Q purification of Sap4 .....	122
5.12	Sap4 at each stage of purification .....	123
5.13	Purified recombinant Sap4.....	123
6.1	Relative Sap4 activity toward various protein substrates.....	128
6.2	Time course of Sap activity in the globin assay.....	128
6.3	Plot of reaction velocity vs. substrate concentration.....	130
6.4	Hanes plot for Sap4 activity using the globin assay.....	130
6.5	The fluorocasein Sap assay .....	132
6.6	pH activity profiles of the Sap isoenzymes.....	132
6.7	Specific activities of Sap isoenzymes in the globin (A) and fluorocasein (B) assays .....	134
6.8	Reversed phase h.p.l.c of glucagon hydrolysates with Sap4.....	136
6.9	Reversed phase h.p.l.c of glucagon hydrolysis by Sap5 .....	137
6.10	Model of cleavage of glucagon by Saps 1-5 .....	140
6.11	Thermal stability profiles of Sap isoenzymes .....	143
6.12	SDS-PAGE analysis of Saps 1-5 .....	145
6.13	Native PAGE analysis of Saps 1-5 .....	145
6.14	Sap autodegradation.....	147
6.15	Pepstatin inhibition of Sap .....	148
6.16	ES mass spectra of Sap1, Sap4 and Sap5.....	149
6.17	Glycopeptide fragment ion analysis.....	150
6.18	DIG glycan/protein analysis of Saps 1, 2, 3 and 4 .....	154
6.19	Sap Dionex analysis.....	158
6.20	Standard curve for quantitation of carbohydrate species .....	159
6.21	O-linked oligosaccharides of <i>S. cerevisiae</i> and <i>P. pastoris</i> .....	161
6.22	Biacore analysis of Sap interactions with SQAPI.....	163
6.23	Antibody titre after second booster.....	166
6.24	Western analysis of anti-Sap4 vs. <i>P. pastoris</i> supernatant.....	166

6.25	Comparison of the sensitivity of anti-Sap2 and anti-Sap4 .....	168
6.26	Anti-Sap4 titre.....	168
6.27	Comparison of the specificity of anti-Sap2 and anti-Sap4.....	169
7.1	Time course of serum-induced hyphal formation.....	173, 174
7.2	Clumped <i>C. albicans</i> strain ATCC 10231 hyphae.....	175
7.3	RNA extracted from strain ATCC 10231 .....	177
7.4	Northern hybridisations for <i>SAP4-6</i> and <i>ACT</i> .....	178
7.5	Northern hybridisations for <i>SAP4-6</i> and <i>LEU2</i> .....	180
8.1	Crystals of native Sap5 from the sparse matrix screen and crystal screen II .....	190
8.2	Crystals of Sap4 complexed with A70450 inhibitor .....	191
8.3	Crystals of Sap4 complexed with SQAPI inhibitor .....	192
8.4	Homology models using the Sap2 structure as a template.....	195
8.5	Active site models of Saps 1, 4 and 5 .....	196
8.6	Sap2 active site structure (in complex with A70450) overlaid with the Sap4 model.....	197
9.1	Model of proposed roles of Sap isoenzymes in the onset and development of candidosis ...	206
10.1	Alignment of the amino acid sequences of Sap1-Sap9 of <i>C. albicans</i> .....	211
10.2	Dionex chromatograms of Sap5.....	215
10.3	Purification of Sap5 using ion-exchange chromatography .....	216
10.4	SDS-PAGE of Sap5 following ion-exchange chromatography .....	217
10.5	Analysis of the specificity of anti-Sap5 .....	218

# LIST OF TABLES

---

1.1	Antifungal agents currently used for treatment of <i>Candida</i> infections .....	12
1.2	Table of identity between <i>SAP</i> sequences from <i>C. albicans</i> .....	27
3.1	Summary of the PCR conditions for <i>SAP4</i> , <i>SAP5</i> and <i>SAP6</i> PCR amplification reactions ....	63
3.2	Comparison of the expected and actual band sizes from restriction enzyme analysis of <i>SAP4</i> , <i>SAP5</i> and <i>SAP6</i> PCR products.....	68
5.1	Purification of recombinant Sap1 .....	111
5.2	Purification of mature Sap4 .....	122
6.1	Glucagon fragments produced by Sap4 and Sap5.....	139
6.2	Consensus sequences of Saps 1-5 cleavage sites in glucagon.....	141
6.3	Amino acid side chains which form subsites S <sub>1</sub> and S <sub>1</sub> ' .....	142
6.4	Predicted and experimental masses of Saps.....	152
6.5	Carbohydrate destruction during acid hydrolysis.....	156
8.1	Sparse matrix crystal screen.....	185
8.2	Crystal screen II.....	186
8.3	Additional crystallisation conditions for Sap1 and Sap4 .....	187

## AMINO ACID ABBREVIATIONS & COLOUR

---

Amino acid	Abbreviation	Symbol
Alanine	Ala	A
Arginine	Arg	R
Asparagine	Asn	N
Aspartic acid	Asp	D
Asparagine or Aspartic acid	Asx	B
Cysteine	Cys	C
Glutamine	Gln	Q
Glutamic acid	Glu	E
Glutamine or Glutamic acid	Glx	Z
Glycine	Gly	G
Histidine	His	H
Isoleucine	Ile	I
Leucine	Leu	L
Lysine	Lys	K
Methionine	Met	M
Phenylalanine	Phe	F
Proline	Pro	P
Serine	Ser	S
Threonine	Thr	T
Tryptophan	Trp	W
Tyrosine	Tyr	Y
Valine	Val	V

Grey	General non-polar	A,M,I,L,V
Green	General polar	S,T,N,Q
Blue	Positively charged	H,K,R
Red	Negatively charged	D,E
Purple	Aromatic	F,W,Y
Brown	“Structure breakers”	G,P
Yellow	Cysteine	C

# ABBREVIATIONS

---

Amp	Ampicillin
BCIG	5'-bromo-4-chloro-3-indolyl- $\gamma$ -D-galactopyranoside
BCIP	5'-bromo-4-chloro-3-indolyl phosphate
bp	base pair
BSA	Bovine serum albumin
CAPS	3-cyclohexylamino-1-propanesulphonic acid
cDNA	Complementary DNA
C-terminal	Carboxy-terminal
Da	Dalton
DEAE	Diethylaminoethyl
DIG	Digoxigenin
DMF	Dimethylformamide
DMSO	Dimethyl sulphoxide
DNA	Deoxyribonucleic acid
dATP	2'-deoxyadenosine-5'-triphosphate
dCTP	2'-deoxycytidine-5'-triphosphate
dGTP	2'-deoxyguanosine-5'-triphosphate
dTTP	2'-deoxythymidine-5'-triphosphate
dNTP	deoxynucleoside triphosphate
DTT	Dithiothreitol
EDTA	Ethylenediamine tetra-acetic acid
ES-MS	Electrospray mass spectrometry
G418	Geneticin – disulphide salt
GalNAc	N-acetyl galactosamine
GlcNAc	N-acetyl glucosamine
HEPES	N-2-hydroxyethylpiperazine-N'-ethanesulphonic acid
h.p.l.c.	High pressure liquid chromatography
IPTG	Isopropyl- $\gamma$ -D-thiogalactopyranoside
kb	kilobase
kDa	kiloDalton
MOPS	3-[N-morpholino]propanesulphonic acid

NBT	Nitro blue tetrazolium
mRNA	messenger RNA
N-terminal	Amino-terminal
PCR	Polymerase chain reaction
PEG	Polyethylene glycol
pI	Isoelectric point
PIPES	Piperazine-n,n'-bis(2-ethanesulphonic acid)
PVDF	Polyvinylidenedifluoride
RNA	Ribonucleic acid
<i>SAP</i>	SAP gene
Sap	Sap protein
SDS	Sodium dodecyl sulphate
SDS-PAGE	Sodium dodecyl sulphate polyacrylamide gel electrophoresis
TEMED	N,N,N',N'-tetramethylethylenediamine
TFA	Trifluoroacetic acid
Tris	Tris(hydroxymethyl)aminomethane
Tween 20	Polyoxyethylene-sorbitan monolaurate
v/v	volume/volume
w/v	weight/volume
w/w	weight/weight

## *C. albicans* GENETIC CODE

		<i>S E C O N D P O S I T I O N</i>								
		<b>T</b>	<b>C</b>	<b>A</b>		<b>G</b>				
<i>F</i>	<b>T</b>	TTT	Phe	TCT	Ser	TAT	Tyr	TGT	Cys	<b>T</b>
		TTC	Phe	TCC	Ser	TAC	Tyr	TGC	Cys	<b>C</b>
		TTA	Leu	TCA	Ser	TAA	*	TGA	*	<b>A</b>
		TTG	Leu	TCG	Ser	TAG	*	TGG	Trp	<b>G</b>
<i>S</i>	<b>C</b>	CTT	Leu	CCT	Pro	CAT	His	CGT	Arg	<b>T</b>
		CTC	Leu	CCC	Pro	CAC	His	CGC	Arg	<b>C</b>
		CTA	Leu	CCA	Pro	CAA	Gln	CGA	Arg	<b>A</b>
		CTG	Ser	CCG	Pro	CAG	Gln	CGG	Arg	<b>G</b>
<i>O</i>	<b>A</b>	ATT	Ile	ACT	Thr	AAT	Asn	AGT	Ser	<b>T</b>
		ATC	Ile	ACC	Thr	AAC	Asn	AGC	Ser	<b>C</b>
		ATA	Ile	ACA	Thr	AAA	Lys	AGA	Arg	<b>A</b>
		ATG	Met	ACG	Thr	AAG	Lys	AGG	Arg	<b>G</b>
<i>O</i>	<b>G</b>	GTT	Val	GCT	Ala	GAT	Asp	GGT	Gly	<b>T</b>
		GTC	Val	GCC	Ala	GAC	Asp	GGC	Gly	<b>C</b>
		GTA	Val	GCA	Ala	GAA	Glu	GGA	Gly	<b>A</b>
		GTG	Val	GCG	Ala	GAG	Glu	GGG	Gly	<b>G</b>

Codons are as they appear in the coding strand of DNA, reading 5'→3'. Termination codons are represented by \*. Note the unusual decoding of CTG as a Serine in *C. albicans*, rather than the standard amino acid Leucine.