Copyright is owned by the Author of the thesis. Permission is given for a copy to be downloaded by an individual for the purpose of research and private study only. The thesis may not be reproduced elsewhere without the permission of the Author.

Is There Sex-Specific DNA In The Tuatara, A Reptile With Temperature-Dependent Sex Determination?

Alex Quinn

A thesis submitted in partial fulfillment of the requirements of a Master of Science degree in the Institute of Natural Resources, Massey University

October 1999

Abstract

It is widely viewed that there is a dichotomy of sex-determining mechanisms within the reptiles: species either exhibit genotypic sex determination or temperature-dependent sex determination (TSD). However, very few species have been examined for both modes. Although it is often considered that the two mechanisms are mutually exclusive, there is evidence that there may be a weak genetic sex-determining mechanism in species in which the primary sex-determining mode is temperature-dependent sex determination. This infers that some TSD individuals may be sex-reversed; that is, their sexual genotype is discordant with their sexual phenotype. This hypothesis of an underlying genotypic system may also be linked to the question of the evolution of sex-determination within the reptiles. The discovery of sex-specific DNA within a TSD reptile could suggest that genotypic sex determination is ancestral and TSD has evolved many times over within independent reptile lineages.

This study tested the hypothesis that there is a genetic component to sex determination in TSD species. This was accomplished by searching for sex-specific DNA in the tuatara, a reptile with temperature-dependent sex determination, using two different molecular genetic techniques.

The major undertaking of the experimental programme was the completion of a comprehensive minisatellite DNA profiling survey. This incorporated 14 restriction enzymes and five different polycore DNA probes; in total, 66 different probe/enzyme combinations were tested for tuatara genomic DNA. None of the DNA profiles revealed sex-specific fragments. Furthermore, a significant difference in mean fragment numbers for males and females was not detected for any of the probe/enzyme combinations.

In addition, a RAPD analysis was conducted in a search for a molecular sex marker in the tuatara. A total of 27 random-sequence oligonucleotide primers were used to successfully amplify anonymous products from the genomic DNA of male and female tuatara. Again, no sex-specific fragments were detected.

Thus, evidence of sex-specific genetic differences in the tuatara was not found. This result fails to refute the null hypothesis that there are underlying sexual genotypes in the tuatara. This finding may reflect the absence of genetic sex differences in the tuatara. Alternatively, it might also be the result of accidental inclusion of sex-reversed individuals within the analyses, a situation which could have obscured the sex-specific nature of any sex-linked fragments. It would appear that the key to solving the question of sex-specific DNA within TSD reptiles such as the tuatara lies with the problem of ensuring sex-reversed individuals are excluded from molecular analysis.

Acknowledgements

For a research project that unfortunately failed to produce the desired result, there seems to be a large number of people I need to thank for their assistance and encouragement.

First of all, I would like to express my appreciation to my supervisor, Dr Steve Sarre, without whom this thesis would not have been possible. Steve has been constantly enthusiastic, patient, and optimistic about the progress of the research, despite many disappointments. I could not have hoped for a better supervisor. I wish Steve all the best for the future of this research, and hope that he discovers the elusive molecular sex marker in the tuatara sooner rather than later.

I would also like to thank Prof Dave Lambert, for allowing me to work in his lab for my thesis, for his contributions to the research, and, along with Steve, for giving me the opportunity to work on such an interesting research project.

A special thank you to Niccy Aitken for the endless assistance she has given me, especially during the early stages of the lab work. I wish her success as she continues the research with Steve.

I am very grateful to Dr Leon Huynen, for his patience in teaching me the intricacies of PCR and RAPD analysis, but also for acting as a springboard, usually against his will, for all my theories about genetic mechanisms of TSD. I would also like to take this opportunity to thank him for all the alcohol he consistently lost to me in numerous rugby and cricket result wagers.

Thanks also to the other members of the Molecular Ecology group for their friendship and assistance throughout the time I have spent in the lab, especially Peter Ritchie and James Bower, particularly for their constant reminders that I needed to work harder (which was usually true).

I would like to convey my appreciation to all those people who at some stage were involved in the collection of the tuatara blood samples that were used in this project, especially Prof Charles Daugherty and Nicki Nelson of Victoria University (also for the information they provided at various times), Graham Ussher of Auckland University, and also Michelle Finch, formerly of Auckland University. Thank you also to Shirley Pledger of Victoria University for her advice on statistical analysis.

To the members of the 3 Ayr Place Institution, past and present, thanks for the memories. You know as well as I do that I don't have many of them. And that poster looked good, you know it did. Opening with tail-enders would work too, I might add. Thank you also to Johnathan and Mike, my fellow thesis sufferers, for their mutual support during this dark period of our lives. I would also like to thank the All Blacks: if it were not for the sure knowledge that they will win back the World Cup this year in Wales, and my long-standing pledge to witness the event, then I perhaps would never have completed this thesis.

Finally, I would like to express my deepest gratitude to my family, for their constant support and encouragement, not just throughout the duration of this degree, but throughout my entire education.

Thesis Structure And Format

I have arranged this thesis in the following manner. Chapter 1 gives a general introduction to the tuatara, describing its evolutionary position, taxonomic status, distribution, and general biology. The aspect of tuatara ecology that is the focus of this study, sex determination, is introduced. Finally, the aim of the thesis is explained, in terms of its relevance to scientific study and also its application to the conservation of this rare species.

In Chapter 2 I provide background material and theory relevant to this research. The phenomenon of temperature-dependent sex determination (TSD) is described in some detail, including a general review of its occurrence within the four major living groups of reptiles (including the tuatara). The concept of genetic sex differences in a species that exhibits temperature-dependent sex determination is examined. Current insights into the possible molecular mechanisms of TSD are discussed, particularly with respect to reconciling the concept of an underlying genotypic mode of sex determination interacting with the TSD mechanism. This discussion is used to give a context to the methodological approach of the study.

Chapter 3 presents a detailed account of the major experimental undertaking of the investigation; a comprehensive minisatellite DNA profiling survey aimed at testing the hypothesis that tuatara have sex-specific DNA. This survey is divided into three distinct phases, consistent with three different sets of tuatara blood samples. The results of a large number of probe/enzyme combinations are presented and discussed.

Chapter 4 is an account of a brief investigation employing RAPD (Randomly Amplified Polymorphic DNA) assays as a further attempt to detect a molecular marker for gender in this species. In this study, a large number of random-sequence oligonucleotide primers were used to amplify anonymous PCR products from male and female tuatara. Results are presented and discussed.

Chapter 5, the final chapter, presents a summary of the findings of the research. Following a general conclusion, there is a discussion of potential avenues of investigation for future research into the question of sex determination in the tuatara (and TSD reptiles in general).

Contents

Abstract		
Acknowledgements		
Thesis Structure and Format		
Contents List of Figures List of Tables		vii
		xi
		xii
Chap	ter 1	
Introd	uction: The Tuatara, Sphenodon spp.	1
1.0	Evolutionary Position	1
1.1	Past and Present Distribution	4
1.2	Taxonomy and Conservation Status	6
1.3	Biology and Ecology of the Tuatara	8
1.4	Sex Determination in the Tuatara	11
1.5	Aim and Description of Thesis	12
1.5.1	Sex-specific DNA in the tuatara	12
1.5.2	Application to conservation	14
Chap	iter 2	
Temp	erature-Dependent Sex Determination In Reptiles: An Overview	16
2.1	Introduction	16
2.1.1	Sex determination and sexual differentiation	17
2.2	Temperature-Dependent Sex Determination in Reptiles	18
2.2.1	Sex ratio biases and pivotal incubation temperatures	19
2.2.1		21
2.2.2	Patterns of temperature-dependent sex determination in reptiles The effect of incubation temperature upon sex determination	23
2.2.3	The effect of incubation temperature upon sex determination Tayonomic Distribution of Say Determining Mechanisms in Pentiles	24
2.3.1	Taxonomic Distribution of Sex-Determining Mechanisms in Reptiles General distribution	24
2.3.1	General distribution	24

2.4	Genetic Involvement in Temperature-Dependent Sex Determination	28
2.4.1	The concept of sex-specific genetic differences in reptiles with	28
	temperature-dependent sex determination	
2.4.2	Examples of the apparent co-occurrence of genotypic and	31
	temperature-dependent sex determination within a species	
2.5	The Molecular Mechanism of Temperature-Dependent	42
	Sex Determination	
2.5.1	Estrogen: the critical factor in reptilian sex determination	42
2.5.2	The role of aromatase in temperature-dependent	43
	sex determination	
2.5.3	Reconciling the hypothesis of an underlying genotypic sex	45
	determination with the aromatase model of TSD	
2.6	An Explanation of the Methodology	45
Chap	iter 3	
Minis	atellite DNA Profiling: Does It Detect Sex-Specific Variation	
In The	e Tuatara?	50
2.1		50
3.1	Introduction	50
3.1.1	Sex assignment in the tuatara: previous investigation	50
	and current objectives	
3.1.2	Molecular sex assignment and minisatellite DNA profiling	51
3.2	Methods and Materials	56
3.2.1	Strategies for detecting sex-specific DNA	56
	using minisatellite DNA profiling	
3.2.2	Sites and collection of blood samples	59
3.2.3	Extraction of genomic DNA	60
3.2.4	Digestion of genomic DNA	60
3.2.5	Gel electrophoresis and Southern blotting	62
3.2.6		
	DNA hybridisation and autoradiography	63
3.2.7	DNA hybridisation and autoradiography Analysis of DNA profiles: Checking for sex-specificity and band	63 66

3.3	Results	69
3.3.1	Probe-enzyme combinations	69
3.3.2	Examination for sex-specific fragments	71
3.3.3	Band-scoring as a test for sex bias in fragment numbers	80
3.4	Discussion	86
Chap	ter 4	
Rando	omly Amplified Polymorphic DNA (RAPD) Assays	
For Se	ex-Specific DNA In The Tuatara	90
4.1	Introduction	90
4.1.1	Molecular sex assignment in the tuatara: previous investigations	90
4.1.2	RAPDs: A useful tool for molecular sex assignment	91
4.2	Methods and Materials	92
4.2.1	Strategies for detecting sex-specific fragments by RAPD analysis	92
4.2.2	Site and collection of blood samples	95
4.2.3	DNA extraction	96
4.2.4	PCR amplification of RAPD products with 10-mer primers	96
4.3	Results	97
4.4	Discussion	102
Chap	ter 5	
Summ	nary, Conclusion, and Future Directions	105
5.1	Review of the Research Aims and Objectives	105
5.2	Summary of the Experimental Programme and Results	106
5.3	Conclusion	106
5.3.1	General implications	107
5.3.2	Implications for conservation	108
5.4	Future Directions for Research	109
5.5	Final Remarks	112

Appendix I	
Morphological data of the Lady Alice Island tuatara	113
Appendix II	
The patterns of temperature-dependent sex determination: two or three?	
Appendix III	
The Evolution of Sex Determination in Reptiles	
The case for an ancestry of temperature-dependent sex determination	116
The case for an ancestry of genotypic sex determination	119
The adaptive significance of temperature-dependent sex determination	122
£	
Appendix IV	
A statistical analysis of potential sex-specific fragments	126
Appendix V	
Sequences of the 10-bp primers used in the RAPD analysis	
References	

List of Figures

Figure 1.1	Map of the current distribution of tuatara	5
Figure 2.1	Hatchling sex ratio versus constant incubation temperature in the Ouachita Map Turtle	20
Figure 2.2	Plots of hatchling sex ratio as a function of constant incubation temperature, showing the three general patterns of TSD in reptiles	22
Figure 3.1	BamHI-, MspI-, and RsaI-digested DNA probed with Bkm (59°C)	70
Figure 3.2	AluI-digested DNA probed with Bkm (63°C)	72
Figure 3.3	HaeIII-digested DNA probed with Bkm (65°C)	73
Figure 3.4	HaeIII-digested DNA probed with pV47-2 (59°C)	74
Figure 3.5	MboII-digested DNA probed with pV47-2 (57°C)	75
Figure 3.6	HaeIII-digested DNA probed with 33.6 (61°C)	76
Figure 3.7	AluI-digested DNA probed with 33.15 (59°C)	77
Figure 3.8	AluI-digested DNA probed with per (63°C)	78
Figure 3.9	BstNI-digested DNA probed with per (57°C)	79
Figure 3.10	HaeIII-digested DNA probed with 33.15 (59°C)	81
Figure 4.1	RAPD band profiles for products amplified from cocktails of male and female DNA with primers OPA-02 and OPA-05	99
Figure 4.2	RAPD band profiles for products amplified from individual tuatara DNA with primers OPA-02, -04, -10 and -11	100
Figure 4.3	A. RAPD profile indicating potential female-specific band for primer OPF-15 B. RAPD profile showing amplification with OPF-15, repeated for eight individuals of each sex	101

List of Tables

Table 1.1	Numbers of male and female tuatara hatchlings for different constant temperature regimes (data of Cree et al, 1995)	13
Table 2.1	Theoretical classes of sex if TSD is superimposed upon GSD	30
Table 3.1	DNA probes used in the three phases of the minisatellite DNA profiling survey	57
Table 3.2	Restriction enzymes used in the minisatellite DNA profiling survey	61
Table 3.3 (a)	Restriction enzyme and probe combinations tested during PHASE 1 of the minisatellite DNA profiling survey	64
Table 3.3 (b)	Restriction enzyme and probe combinations tested during PHASE 2 of the minisatellite DNA profiling survey	65
Table 3.3(c)	Restriction enzyme and probe combinations tested during PHASE 3 of the minisatellite DNA profiling survey	65
Table 3.4 (a)	Mean numbers of DNA fragments for phenotypic males and females for PHASE 1 probe-enzyme combinations	82
Table 3.4 (b)	Mean numbers of DNA fragments for phenotypic males and females for PHASE 2 probe-enzyme combinations	83
Table 3.4(c)	Mean numbers of DNA fragments for phenotypic males and females for PHASE 3 probe-enzyme combinations	84
Table 3.5	Comparison of band counts from current study with the data of Finch (1994): HaeIII-digested DNA probed with 33.15	85